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(54) Title: ASPERGILLUS OCHRACEUS 11 ALPHA HYDROXYLASE AND OXIDOREDUCTASE

(57) Abstract: The present invention relates to a novel cytochrome P450-like enzyme (*Aspergillus ochraceus*) isolated from cDNA library generated from the mRNA of *Aspergillus ochraceus* spores. When the cDNA encoding the 11 alpha hydroxylase was co-expressed in *Spodoptera frugiperda* (Sf-9) insect cells with the cDNA encoding human oxidoreductase as an electron donor, it successfully catalyzed the conversion of the steroid substrate 4-androstene-3,17-dione (AD) to 11 alpha-hydroxy-AD as determined by HPLC analysis. The invention also relates to nucleic acid molecules associated with or derived from these cDNAs including complements, homologues and fragments thereof, and methods of using these nucleic acid molecules, to generate, for example, polypeptides and fragments thereof. The invention also relates to the generation of antibodies that recognizes the *A. ochraceus* 11 alpha hydroxylase and oxidoreductase and methods of using these antibodies to detect the presence of these native and recombinant polypeptides within unmodified and transformed host cells, respectively. The invention also provides methods of expressing the *Aspergillus* 11 alpha hydroxylase gene separately, or in combination with human or *Aspergillus* oxidoreductase, in heterologous host cells, to facilitate the bioconversion of steroid substances to their 11 alpha hydroxy-counterparts.

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Aspergillus ochraceus 11 alpha hydroxylase and oxidoreductase

Priority

The present application claims priority under Title 35, United States Code,
5 § 119 of United States Provisional Application Serial No. 60/244,300, filed October
30, 2000.

Field of the invention

The present invention relates to a novel cytochrome P450-like enzyme
(*Aspergillus ochraceus* 11 alpha hydroxylase) and an oxidoreductase (*Aspergillus*
10 *ochraceus* oxidoreductase) isolated from cDNA library generated from the mRNA of
Aspergillus ochraceus spores. When the cDNA encoding the 11 alpha hydroxylase
was co-expressed in *Spodoptera frugiperda* (Sf-9) insect cells with the cDNA
encoding human oxidoreductase as an electron donor, it successfully catalyzed the
conversion of the steroid substrate 4-androstene-3,17-dione (AD) to 11 alpha-
15 hydroxy-AD as determined by HPLC analysis. The invention also relates to nucleic
acid molecules associated with or derived from these cDNAs including
complements, homologues and fragments thereof, and methods of using these
nucleic acid molecules, to generate, for example, polypeptides and fragments
thereof. The invention also relates to the generation of antibodies that recognize
20 the *A. ochraceus* 11 alpha hydroxylase and oxidoreductase and methods of using
these antibodies to detect the presence of these native and recombinant
polypeptides within unmodified and transformed host cells, respectively. The
invention also provides methods of expressing the *Aspergillus* 11 alpha
hydroxylase gene separately, or in combination with human or *Aspergillus*
25 oxidoreductase, in heterologous host cells, to facilitate the bioconversion of steroid
substrates to their 11 alpha hydroxy-counterparts.

Background of the invention

Microbial transformation or bioconversion reactions have long been used to
facilitate the chemical synthesis of a wide variety of pharmaceutical products.
30 Stereospecific reactions carried out under mild enzymatic conditions frequently
offer advantages over comparable chemical processes which result in undesirable

side products. Microorganisms also have the ability to carry out simultaneous independent or sequential reactions on a substrate molecule, minimizing the number of distinct steps in a synthesis and reducing the total cost of the desired intermediate or end product.

5 General features of microbial systems used as biocatalysts for the transformation of organic compounds has been reviewed (See e.g., Goodhue, Charles T., *Microb. Transform. Bioact. Compd.*, 1: 9-44, 1982). Biotransformations can be carried out, for example, in continuous cultures or in batch cultures. Enzymes secreted from the microorganism react with a substrate, and the product 10 can be recovered from the medium. Intracellular enzymes can also react with a substrate if it is able to enter cells by an active or a passive diffusion process. Immobilized, dried, permeabilized, and resting cells, and spores have also been used for microbial transformations. The use of cell extracts and purified enzymes 15 in solution, or immobilized on carriers, may eventually offer significant cost or control advantages over traditional fermentation methods.

Bioconversion reactions have been widely used in the field of steroids (Kieslich, K.; Sebek, O. K. *Annu. Rep. Ferment. Processes* 3: 275-304, 1979; Kieslich, Klaus. *Econ. Microbiol.*, 5 (Microb. Enzymes Bioconvers.): 369-465, 1980). A variety of reactions have been characterized, including hydroxylation, 20 epoxidation, oxidation, dehydrogenation, ring and side chain degradation, reduction, hydrolysis, and isomerization reactions. Many types of microorganisms have also been used including species as diverse, for example, as *Acremonium*, *Aspergillus*, *Rhizopus*, *Fusarium*, *Penicillium*, *Streptomyces*, *Actinomyces*, *Nocardia*, *Pseudomonas*, *Mycobacterium*, *Arthrobacter* and *Bacillus*.

25 A variety of approaches have been used to facilitate the hydroxylation of intermediates used in the synthesis of commercially-important steroid compounds. US patent 4,588,683, for example, describes a method of preparing 11 beta, 17 alpha, 20, 21 tetrahydroxy steroids by incubating substrate compounds in a medium comprising a fungal culture of the genus *Curvularia* capable of effecting 30 11 beta hydroxylation. *Aspergillus ochraceus* cultures and preparations of mycelia have also been used to convert progesterone and other steroids to their corresponding 11 alpha hydroxy forms (Tan, L. and Falardeau, P., 1970; Tan L., and Falardeau P., *J. Steroid Biochem.* 1: 221-227, 1970; Samanta, T.B. et al., *Biochem. J.* 176, 593-594, 1978; Jayanthi, C.R. et al., *Biochem. Biophys. Res. Commun.* 106: 1262-1268, 1982).

The advent of new and expanded clinical uses of steroids for the treatment of a wide variety of disorders has created a need for improved methods for the production of steroid compounds and their intermediates on a commercial scale. U.S. patent 4,559,332, for example, describes a number of methods for the preparation of 20-spiroxane series of steroid compounds, including methods for the preparation of eplerenone methyl hydrogen 9,11 α -epoxy-17 α -hydroxy-3-oxopregn-4-ene-7 α ,21-dicarboxylate, γ -lactone (also referred to as eplerenone or epoxymexrenone) and related compounds. WO 98/25948 and U.S. application 09/319,673 describe novel processes for the preparation of 9,11-epoxy steroid compounds, especially those of the 20-spiroxane series and their analogs, novel intermediates useful in the preparation of steroid compounds, and processes for the preparation of such novel intermediates. United States Patent 6,046,023 discloses improved methods for the microbial transformation of canrenone or estr-4-ene-3,17-dione into its 11 α -hydroxy analogue using microorganisms of the genus *Aspergillus*, *Rhizopus*, and *Pestelotia*, using steroid substrates having a purity of less than 97% and more than 90% at a concentration greater than 10 g/L.

Many modern, systematic approaches needed to optimize bioconversion of particular steroid intermediates are often hindered by insufficient biochemical knowledge of the enzymes involved in their synthesis and degradation. Eukaryotic cytochromes P450 appear to be associated with the endoplasmic reticulum (ER) or mitochondrial membranes. The electron donor for ER-associated cytochrome P450 enzymes is often an FAD/FMN-dependent NADPH-cytochrome P450 oxidoreductase. Electron transfer in the mitochondrial cytochromes P450 is usually mediated by an NADPH-ferredoxin oxidoreductase and ferrodoxin. The specific electron donors known to be involved in mammalian steroidogenesis, are also called adrenodoxin reductase and adrenodoxin, respectively.

While fungal biotransformations are known to be mediated by cytochrome P450 enzymes, many of these enzymes are extremely difficult to purify in an enzymatically-active form (van den Brink et al., *Fungal Genetics and Biology* 23, 1-17, 1998). Many fungal P450 enzymes appear to be associated with the endoplasmic reticulum (van den Brink et al., *Fungal Genetics and Biology* 23, 1-17, 1998). Yeast have an adrenodoxin reductase homologue which was shown to couple with a mammalian 11 beta hydroxylase *in vitro*. (Lacour et al., *Journal of Biological Chemistry* 273, 23984-23992, 1998). In contrast, the electron donor which couples with *Aspergillus ochraceus* 11 alpha hydroxylase was predicted to be an NADPH-cytochrome P450 oxidoreductase (Samanta and Ghosh, *J Steroid Biochem* 28, 327-32, 1987). The steroid 11 alpha hydroxylation complex in

5 *Rhizopus nigricans* also appears to require an NADPH-cytochrome p450 oxidoreductase (Makovec and Breskvar, *Arch Biochem Biophys.* **357**, 310-6, 1998). Amplification of cytochrome *R. nigricans* P450 and NADPH-cytochrome P450 reductase activities in preparations of progesterone-induced fungal mycelia may facilitate biochemical characterization of both enzymes (Makovec and Breskvar, *Pflugers Arch - Eur J. Physiol* 439(Suppl): R111-R112, 2000).

10 *Aspergillus ochraceus* spores have been shown to catalyze the 11 alpha hydroxylation of steroid substrates such as progesterone (Dutta TK, Datta J, Samanta TB, *Biochem. Biophys. Res. Commun.* **192**:119-123, 1993). *A. fumigatus* is also known to exhibit a steroid 11 alpha hydroxylase activity (Smith et al., *J Steroid Biochem Mol Biol* **49**: 93-100, 1994). The *A. fumigatus* enzyme is distinguished from the *A. ochraceus* enzyme, in that it appears to be a cytochrome P450 with dual site-specificity for 11 alpha and 15 beta hydroxylation and, unlike the *A. ochraceus* hydroxylase, appears to be non-inducible.

15 Despite recent advances in sequencing technologies, detailed knowledge about the structural relationships of fungal cytochrome P450s gleaned from nucleotide sequence data remains primitive. Breskvar et al., (*Biochem. Biophys. Res. Commun* 1991; **178**, 1078-1083, 1991) have described a genomic DNA sequence from *Rhizopus nigricans* for a putative P-450 encoding an 20 11 α -hydroxylase for progesterone. This sequence may not be complete, however, since the predicted amino acid sequence lacks the canonical heme-binding motif, FxxGxxxCxG, which is common to almost all known cytochrome P-450 enzymes. (Nelson et al, *Pharmacogenetics* **6**: 1-42, 1996).

25 The cloning and characterization of the NADPH cytochrome P450 oxidoreductase (cprA) gene of *Aspergillus niger* has been described (van den Brink, J., et al., Genbank accession numbers Z26938, CAA81550, 1993, unpublished). The primary structure of *Saccharomyces cerevisiae* NADPH-cytochrome P450 reductase has also been deduced from the nucleotide sequence of its cloned gene (Yabusaki et al., *J. Biochem.* **103**(6): 1004-1010, 1988).

30 Several other approaches have been used to facilitate the cloning and analysis of steroid enzymes. U.S. patents 5,422,262, 5,679,521, and European patent EP 0 528 906 B1, for example, describes the expression cloning of steroid 5 alpha reductase, type 2. U.S. patent 5,869,283, for example, describes an expression cassette comprising heterologous DNAs encoding two or more enzymes, 35 each catalyzing an oxidation step involved conversion of cholesterol into

hydrocortisone, including the conversion of cholesterol to pregnenolone; the conversion of pregnenolone to progesterone; the conversion of progesterone to 17 α -hydroxy-progesterone; the conversion of 17 α -hydroxyprogesterone to cortexolone; and the conversion of cortexolone to hydrocortisone.

5 The sequences of *Aspergillus ochraceus* 11 alpha hydroxylase and *A. ochraceus* oxidoreductase have not been reported. Knowledge about their sequence could greatly facilitate the development of expression vectors and recombinant host strains that can carry out more efficient bioconversions of steroid intermediates and the synthesis of end products on a commercial scale without the problems
10 associated with partially-characterized host strains or an incomplete understanding of the enzymes involved in steroidogenesis. The present invention overcomes many of the limitations discussed above by identifying enzymes capable of carrying out the 11 alpha hydroxylation of steroids. This approach not only greatly facilitates the use of 11 alpha hydroxylation, but also permits the
15 development of new strategies for the identification of similar enzymes from other fungi, the cloning of other enzymes involved in steroidogenesis from *Aspergillus ochraceus* and other microorganisms, and the development of improved host strains or methods using free cells or immobilized cells or enzymes in bioconversion reactions. Similar approaches could also be developed to aid in the construction of
20 expression vectors and recombinant host strains that are more amenable to propagation and control than wild-type microorganisms now commonly used for bioconversion in large scale bioreactors.

Summary of the invention

25 In its broadest scope, the present invention provides a method to clone enzymes involved in steroid metabolism and use of these enzymes to produce novel steroid intermediates and end-products. One aspect of the claimed invention is to provide a novel enzyme 11 alpha hydroxylase and oxidoreductase, and their nucleic acids, proteins, peptides, fragments, and homologues. The invention also relates to methods of identifying and cloning other enzymes involved in steroid metabolism.
30 The invention also covers novel vectors and host cells, a novel method for making heterologous proteins by using the above vectors, and a method for identifying the substrate specificity of the cloned enzymes.

35 The invention provides a means for determining the substrate specificity of the cloned 11 alpha hydroxylase, allelic variants, muteins, and fusion proteins thereof, permitting evaluation of a broad array of steroid substrates including 3

keto delta 4,5 steroids (3 keto delta 4 steroids); 3 keto delta 4, 5 delta 6, 7 steroids (3 keto delta 4 delta 6 steroids); 3 keto delta 6, 7 steroids (3 keto delta 6 steroids); or 3 keto delta 1, 2 delta 4, 5 steroids (3 keto delta 1 delta 4 steroids). Preferred substrates for testing include (a) canrenone; (b) androstenedione; (c) aldonia; (d) ADD (1,4 androstenedione) (e) mexrenone; (f) 6 beta mexrenone; (g) 9 alpha mexrenone; (h) 12 beta mexrenone; (i) delta 12 mexrenone; (j) testosterone; (k) progesterone; (l) mexrenone 6,7-bis-lactone; and (m) mexrenone 7,9-bislactone. Preferably the cloned 11 alpha hydroxylase, allelic variants, muteins, and fusion proteins thereof do not also catalyze a second hydroxylation selected from the group consisting of 15 alpha or beta hydroxylation, 6 alpha or beta hydroxylation, 7 alpha or beta hydroxylation, 9 alpha or beta hydroxylation, 12 alpha or beta hydroxylation, and 17 alpha or beta hydroxylation of substrates selected from the group consisting of 3 keto delta 4,5 steroids; 3 keto delta 4, 5 delta 6, 7 steroids; or 3 keto delta 6, 7 steroids. Most preferably the cloned 11 alpha hydroxylase, allelic variants, muteins, and fusion proteins thereof do not catalyze the 15 beta hydroxylation of substrates selected from the group consisting of 3 keto delta 4,5 steroids; 3 keto delta 4, 5 delta 6, 7 steroids; or 3 keto delta 6, 7 steroids.

The invention provides an isolated and purified nucleic acid, encoding *Aspergillus ochraceus* 11 alpha hydroxylase. It also provides an isolated DNA, cDNA, gene, and an allele of the gene encoding *Aspergillus ochraceus* 11 alpha hydroxylase. Preferably the isolated and purified nucleic acid is as set forth in SEQ ID NO: 01. Preferably the isolated DNA, cDNA, gene, and an allele of the gene is as set forth in SEQ ID NO: 01.

The invention provides an isolated protein having the amino acid sequence of *Aspergillus ochraceus* 11 alpha hydroxylase. It also provides an isolated variant of *Aspergillus ochraceus* 11 alpha hydroxylase, and a fusion protein comprising this hydroxylase. Preferably the protein is as set forth in SEQ ID NO: 2. It also provides for variant of the protein set forth in SEQ ID NO: 2.; a polypeptide which comprises SEQ ID NO: 2 with at least one conservative amino acid substitution; polypeptides, with an amino acid sequence at least 99%, 95%, 90%, 75%, and 50% identical to SEQ ID NO: 2.

The invention provides an isolated and purified nucleic acid, encoding *Aspergillus ochraceus* 11 alpha oxidoreductase. It also provides an isolated DNA, cDNA, gene, and allele of the gene encoding *Aspergillus ochraceus* oxidoreductase. Preferably, the isolated and purified nucleic acid, wherein said nucleic acid

sequence is as set forth in SEQ ID NO: 5. It also provides for an isolated DNA, cDNA, gene, and allele of the gene set forth in SEQ ID NO: 5.

The invention provides an isolated protein having the amino acid sequence of *Aspergillus ochraceus* oxidoreductase. It also provides an isolated variant of the 5 protein having the amino acid sequence of *Aspergillus ochraceus* oxidoreductase, and a fusion protein comprising the amino acid sequence of *Aspergillus ochraceus* oxidoreductase. Preferably the isolated protein has the amino acid sequence set forth in SEQ ID NO: 6. It also provides an isolated variant of a protein set forth in 10 SEQ ID NO: 6, a purified polypeptide, the amino acid sequence of which comprises SEQ ID NO: 6 with at least one conservative amino acid substitution; and a polypeptides with an amino acid sequence at least 99%, 95%, 90%, 75%, and 50% identical to SEQ ID NO: 6.

The invention provides an isolated and purified nucleic acid encoding an 15 enzyme that can catalyze the 11 alpha hydroxylation of 3 keto delta 4,5 steroids (3 keto delta 4 steroids); 3 keto delta 4, 5 delta 6, 7 steroids (3 keto delta 4 delta 6 steroids); 3 keto delta 6, 7 steroids (3 keto delta 6 steroids); or 3 keto delta 1, 2 delta 4, 5 steroids (3 keto delta 1 delta 4 steroids). Preferably the enzyme does not catalyze the 15 beta hydroxylation of 3 keto delta 4,5 steroids; 3 keto delta 4, 5 delta 6, 7 steroids; or 3 keto delta 6, 7 steroids. More preferably, the hydroxylation 20 is selected from the group consisting of: (a) canrenone to 11 alpha hydroxy canrenone; (b) androstenedione to 11 alpha hydroxy androstenedione; (c) aldona to 11 alpha hydroxy aldona; (d) ADD (1,4 androstenedienedione) to 11 alpha hydroxy ADD; (e) mexrenone to 11 alpha hydroxy mexrenone; (f) 6 beta mexrenone to 11 alpha hydroxy 6 beta mexrenone; (g) 9 alpha mexrenone to 11 alpha hydroxy 9 alpha mexrenone; (h) 12 beta mexrenone to 11 alpha hydroxy 12 beta mexrenone; (i) delta 12 mexrenone to 11 alpha hydroxy delta 12 mexrenone; (j) testosterone to 11 alpha hydroxy testosterone; (k) progesterone to 11 alpha hydroxy progesterone; (l) mexrenone 6,7-bis-lactone to 11 alpha hydroxy mexrenone 6,7-bis-lactone; and (m) mexrenone 7,9-bislactone to 11 alpha hydroxy mexrenone 7,9-bislactone. More 25 preferably, the hydroxylation is selected from the group consisting of: (a) canrenone to 11 alpha hydroxy canrenone; (b) androstenedione to 11 alpha hydroxy androstenedione; (c) aldona to 11 alpha hydroxy aldona; and (d) ADD (1,4 androstenedienedione) to 11 alpha hydroxy ADD. Most preferably the hydroxylation is from canrenone to 11 alpha hydroxy canrenone.

35 The invention also provides a method of expressing a protein that can catalyze the 11 alpha hydroxylation of 3 keto delta 4,5 steroids; 3 keto delta 4, 5

delta 6, 7 steroids; 3 keto delta 6, 7 steroids; or 3 keto delta 1, 2 delta 4, 5 steroids comprising; (a) transforming or transfecting host cells with an expression cassette comprising a promoter operably linked to a nucleic acid that encodes said protein, and (b) expressing said protein in said host cells. The invention also provides for a 5 method of producing the protein further comprising the step of recovering said protein. Preferably, this protein is *Aspergillus ochraceus* 11 alpha hydroxylase. More preferably, this method further comprises expressing an electron donor protein, wherein said electron donor protein can donate electrons to said protein that can catalyze the 11 alpha hydroxylation of 3 keto delta 4,5 steroids; 3 keto 10 delta 4, 5 delta 6, 7 steroids; 3 keto delta 6, 7 steroids; or 3 keto delta 1, 2 delta 4, 5 steroids. Preferably, the electron donor protein is selected from the group consisting of human oxidoreductase and *Aspergillus ochraceus* oxidoreductase. More preferably the electron donor protein is *Aspergillus ochraceus* oxidoreductase. More preferably, the nucleic acid encoding said steroid 11 alpha hydroxylase and 15 said electron donor protein are on separate expression cassettes. More preferably, the nucleic acid encoding said steroid 11 alpha hydroxylase and said electron donor protein are on the same expression cassettes. Even more preferably, the steroid 11 alpha hydroxylase is *Aspergillus ochraceus* 11 alpha hydroxylase and said electron donor protein is human oxidoreductase. Even more preferably, the steroid 11 alpha hydroxylase is *Aspergillus ochraceus* 11 alpha hydroxylase and said electron 20 donor protein is *Aspergillus ochraceus* oxidoreductase. Preferably, the expression cassette is on an expression vector. More preferably, the expression vector is a baculovirus. Even more preferably, the baculovirus is a nuclear polyhedrosis virus is selected from the group consisting of *Autographa californica* nuclear 25 polyhedrosis virus and *Bombyx mori* nuclear polyhedrosis virus. Most preferably, the nuclear polyhedrosis virus is *Autographa californica* nuclear polyhedrosis virus. Preferably, the host cells are insect cells. More preferably, the insect cells are selected from the group consisting of *Spodoptera frugiperda*, *Trichoplusia ni*, *Autographa californica*, and *Manduca sexta* cells. Most preferably the insect cells 30 are *Spodoptera frugiperda* cells. The invention also provides for a method of expressing a protein wherein the *Aspergillus ochraceus* 11 alpha hydroxylase is SEQ ID NO: 2; the human oxidoreductase is SEQ ID NO: 4; and the *Aspergillus ochraceus* oxidoreductase is SEQ ID NO: 6.

The invention also provides for an isolated and purified polypeptide that 35 can catalyze the 11 alpha hydroxylation of 3 keto delta 4,5 steroids (3 keto delta 4 steroids); 3 keto delta 4, 5 delta 6, 7 steroids (3 keto delta 4 delta 6 steroids); 3 keto delta 6, 7 steroids (3 keto delta 6 steroids); or 3 keto delta 1, 2 delta 4, 5 steroids (3

keto delta 1 delta 4 steroids). Preferably, the polypeptide does not catalyze the 15 beta hydroxylation of 3 keto delta 4,5 steroids; 3 keto delta 4, 5 delta 6, 7 steroids; or 3 keto delta 6, 7 steroids. More preferably, the hydroxylation is selected from the group consisting of: (a) canrenone to 11 alpha hydroxy canrenone; (b) androstenedione to 11 alpha hydroxy androstenedione; (c) aldona to 11 alpha hydroxy aldona; (d) ADD (1,4 androstenedienedione) to 11 alpha hydroxy ADD; (e) mexrenone to 11 alpha hydroxy mexrenone; (f) 6 beta mexrenone to 11 alpha hydroxy 6 beta mexrenone; (g) 9 alpha mexrenone to 11 alpha hydroxy 9 alpha mexrenone; (h) 12 beta mexrenone to 11 alpha hydroxy 12 beta mexrenone; (i) 10 delta 12 mexrenone to 11 alpha hydroxy delta 12 mexrenone; (j) testosterone to 11 alpha hydroxy testosterone; (k) progesterone to 11 alpha hydroxy progesterone; (l) mexrenone 6,7-bis-lactone to 11 alpha hydroxy mexrenone 6,7-bis-lactone; and (m) mexrenone 7,9-bislactone to 11 alpha hydroxy mexrenone 7,9-bislactone. More preferably, the hydroxylation is selected from the group consisting of: (a) 15 canrenone to 11 alpha hydroxy canrenone; (b) androstenedione to 11 alpha hydroxy androstenedione; (c) aldona to 11 alpha hydroxy aldona; and (d) ADD (1,4 androstenedienedione) to 11 alpha hydroxy ADD. Most preferably the hydroxylation is from canrenone to 11 alpha hydroxy canrenone.

The invention also provides for an expression cassette comprising a promoter operably linked to an isolated and purified nucleic acid encoding a polypeptide that can catalyze the 11 alpha hydroxylation of 3 keto delta 4,5 steroids (3 keto delta 4 steroids); 3 keto delta 4, 5 delta 6, 7 steroids (3 keto delta 4 delta 6 steroids); 3 keto delta 6, 7 steroids (3 keto delta 6 steroids); or 3 keto delta 1, 2 delta 4, 5 steroids (3 keto delta 1 delta 4 steroids). More preferably, the hydroxylation is selected from the group consisting of: (a) canrenone to 11 alpha hydroxy canrenone; (b) androstenedione to 11 alpha hydroxy androstenedione; (c) aldona to 11 alpha hydroxy aldona; (d) ADD (1,4 androstenedienedione) to 11 alpha hydroxy ADD; (e) mexrenone to 11 alpha hydroxy mexrenone; (f) 6 beta mexrenone to 11 alpha hydroxy 6 beta mexrenone; (g) 9 alpha mexrenone to 11 alpha hydroxy 9 alpha mexrenone; (h) 12 beta mexrenone to 11 alpha hydroxy 12 beta mexrenone; (i) 30 delta 12 mexrenone to 11 alpha hydroxy delta 12 mexrenone; (j) testosterone to 11 alpha hydroxy testosterone; (k) progesterone to 11 alpha hydroxy progesterone; (l) mexrenone 6,7-bis-lactone to 11 alpha hydroxy mexrenone 6,7-bis-lactone; and (m) mexrenone 7,9-bislactone to 11 alpha hydroxy mexrenone 7,9-bislactone. More preferably, the hydroxylation is selected from the group consisting of: (a) canrenone to 11 alpha hydroxy canrenone; (b) androstenedione to 11 alpha hydroxy androstenedione; (c) aldona to 11 alpha 35 hydroxy aldona; and (d) ADD (1,4 androstenedienedione) to 11 alpha hydroxy ADD.

hydroxy aldon; and (d) ADD (1,4 androstenedienedione) to 11 alpha hydroxy ADD. Most preferably the hydroxylation is from canrenone to 11 alpha hydroxy canrenone.

The invention also provides for an expression cassette comprising a promoter operably linked to an isolated and purified nucleic acid encoding *Aspergillus ochraceus* oxidoreductase. Preferably the nucleic acid is SEQ ID NO: 6.

The invention also provides for an expression cassette comprising a heterologous DNA encoding an enzyme from the metabolic pathway for the synthesis of sitosterol to eplerenone wherein said enzyme catalyzes at least one conversion selected from the group consisting of: (a) canrenone to 11 alpha hydroxy canrenone; (b) androstenedione to 11 alpha hydroxy androstenedione; (c) aldon to 11 alpha hydroxy aldon; (d) ADD (1,4 androstenedienedione) to 11 alpha hydroxy ADD; (e) mexrenone to 11 alpha hydroxy mexrenone; (f) 6 beta mexrenone to 11 alpha hydroxy 6 beta mexrenone; (g) 9 alpha mexrenone to 11 alpha hydroxy 9 alpha mexrenone; (h) 12 beta mexrenone to 11 alpha hydroxy 12 beta mexrenone; (i) delta 12 mexrenone to 11 alpha hydroxy delta 12 mexrenone; (j) testosterone to 11 alpha hydroxy testosterone; and (k) progesterone to 11 alpha hydroxy progesterone; (l) mexrenone 6,7-bis-lactone to 11 alpha hydroxy mexrenone 6,7-bis-lactone; and (m) mexrenone 7,9-bislactone to 11 alpha hydroxy mexrenone 7,9-bislactone and wherein the heterologous DNA is operably linked to control sequences required to express the encoded enzymes in a recombinant host. Preferably the heterologous DNA coding sequences in the expression cassette are selected from the group consisting of the following genus and species: *Aspergillus ochraceus*, *Aspergillus ochraceus*, *Aspergillus niger*, *Aspergillus nidulans*, *Rhizopus oryzae*, *Rhizopus stolonifer*, *Streptomyces fradiae*, *Bacillus megaterium*, *Pseudomonas cruciviae*, *Trichothecium roseum*, *Fusarium oxysporum* *Rhizopus arrhizus*, *Absidia coerulea*, *Absidia glauca*, *Actinomucor elegans*, *Aspergillus flavipes*, *Aspergillus fumigatus*, *Beauveria bassiana*, *Botryosphaeria obtusa*, *Calonectria decora*, *Chaetomium cochlioides*, *Corynespora cassiicola*, *Cunninghamella blakesleeana*, *Cunninghamella echinulata*, *Cunninghamella elegans*, *Curvularia clavata*, *Curvularia lunata*, *Cylindrocarpon radicicola*, *Epicoccum humicola*, *Gongronella butleri*, *Hypomyces chrysospermus*, *Monosporium olivaceum*, *Mortierella isabellina*, *Mucor mucedo*, *Mucor griseocyanus*, *Myrothecium verrucaria*, *Nocardia corallina*, *Paecilomyces carneus*, *Penicillium patulum*, *Pithomyces atroolivaceus*, *Pithomyces cynodontis*, *Pycnosporium* sp., *Saccharopolyspora erythrae*, *Sepedonium chrysospermum*, *Stachyliodium bicolor*, *Streptomyces hygroscopicus*, *Streptomyces purpurascens*,

5 *Syncephalastrum racemosum*, *Thamnostylum piriforme*, *Thielavia terricola*, and
 Verticillium theobromae, *Cephalosporium aphidicola*, *Cochliobolus lunatas*,
 Tieghemella orchidis, *Tieghemella hyalospora*, *Monosporium olivaceum*,
 Aspergillus ustus, *Fusarium graminearum*, *Verticillium glaucum*, and *Rhizopus*
10 *nigricans*. More preferably, the genus and species are selected from the group
 consisting of *Aspergillus ochraceus*, *Aspergillus ochraceus*, *Aspergillus niger*,
 Aspergillus nidulans, *Rhizopus oryzae*, *Rhizopus stolonifer*, *Streptomyces fradiae*,
 Bacillus megaterium, *Pseudomonas cruciviae*, *Trichothecium roseum*, *Fusarium*
 oxysporum, *Rhizopus arrhizus*, and *Monosporium olivaceum*. Most preferably,
15 genus and species is *Aspergillus ochraceus*.

15 Preferably, the recombinant host cell and progeny thereof comprise at least
 one expression cassette. More preferably, the host is a microorganism. Most
 preferably, the host is a bacterium. The invention also provides for a process for
 making one or more enzymes from the metabolic pathway for the transformation of
 sitosterol to eplerenone comprising incubating the recombinant host cell in a
20 nutrient medium under conditions where the one or more enzymes encoded by the
 heterologous DNA are expressed and accumulate. More preferably the process
 comprises the steps of: (a) incubating the compound to be oxidized in the presence
 of the recombinant host cells under conditions where the compound is hydroxylated
25 and the hydroxylated product accumulates, and (b) recovering the hydroxylated
 product. Most preferably, the process comprises the steps of: (a) incubating the
 compound to be oxidized in the presence of the enzymes produced under conditions
 where the compound is hydroxylated and the hydroxylated product accumulates,
 and (b) recovering the hydroxylated product. The invention also provides for a host
30 cells harboring an expression cassette. More preferably the expression cassette is
 integrated into the chromosome of said host cell. More preferably, the expression
 cassette is integrated into an expression vector.

30 The invention also provides for a method of determining the specific
 activity of a cloned 11 alpha hydroxylase comprising the steps of; (a) transforming
 host cells with an expression vector comprising a nucleic acid that encodes said 11
 alpha hydroxylase, (b) expressing said 11 alpha hydroxylase in said host cells; (c)
 preparing subcellular membrane fractions from said cells, (d) incubating said
 subcellular membrane fractions with a steroid substrate, and (e) monitoring
 conversion of the steroid substrate to its 11 alpha hydroxy steroid counterpart.
35 Preferably, the further comprises transforming host cells with an expression vector
 nucleic acid that encodes an oxidoreductase, and expressing said oxidoreductase in
 said host cells. More preferably, the oxidoreductase is human or *Aspergillus*

ochraceus. Most preferably the oxidoreductase is human oxidoreductase. Most preferably the oxidoreductase is *Aspergillus ochraceus* oxidoreductase.

The invention also provides for a protein having SEQ ID NO: 2 and variants thereof that are at least 95% identical to SEQ ID NO: 2 and catalyze the 5 11 alpha hydroxylation of 3 keto delta 4,5 steroids; 3 keto delta 4, 5 delta 6, 7 steroids; 3 keto delta 6, 7 steroids; or 3 keto delta 1, 2 delta 4, 5 steroids, wherein said hydroxylation is selected from the group consisting of: (a) canrenone to 11 alpha hydroxy canrenone; (b) androstenedione to 11 alpha hydroxy androstenedione; (c) aldonia to 11 alpha hydroxy aldonia; (d) ADD (1,4 10 androstenedienedione) to 11 alpha hydroxy ADD; (e) mexrenone to 11 alpha hydroxy mexrenone; (f) 6 beta mexrenone to 11 alpha hydroxy 6 beta mexrenone; (g) 9 alpha mexrenone to 11 alpha hydroxy 9 alpha mexrenone; (h) 12 beta mexrenone to 11 alpha hydroxy 12 beta mexrenone; (i) delta 12 mexrenone to 11 alpha hydroxy delta 12 mexrenone; (j) testosterone to 11 alpha hydroxy 15 testosterone; and (k) progesterone to 11 alpha hydroxy progesterone. Preferably the enzyme does not catalyze the 15 beta hydroxylation of 3 keto delta 4,5 steroids; 3 keto delta 4, 5 delta 6, 7 steroids; or 3 keto delta 6, 7 steroids.

The invention provides an isolated and purified nucleic acid encoding an 20 enzyme that can catalyze the 11 alpha hydroxylation of 3 keto delta 4,5 steroids (3 keto delta 4 steroids); 3 keto delta 4, 5 delta 6, 7 steroids (3 keto delta 4 delta 6 steroids); 3 keto delta 6, 7 steroids (3 keto delta 6 steroids); or 3 keto delta 1, 2 delta 4, 5 steroids (3 keto delta 1 delta 4 steroids) wherein the hydroxylation is 25 selected from the group consisting of: (a) canrenone to 11 alpha hydroxy canrenone; (b) androstenedione to 11 alpha hydroxy androstenedione; (c) aldonia to 11 alpha hydroxy aldonia; (d) ADD (1,4 androstenedienedione) to 11 alpha hydroxy ADD; (e) mexrenone to 11 alpha hydroxy mexrenone; (f) 6 beta mexrenone to 11 alpha hydroxy 6 beta mexrenone; (g) 9 alpha mexrenone to 11 alpha hydroxy 9 alpha mexrenone; (h) 12 beta mexrenone to 11 alpha hydroxy 12 beta mexrenone; (i) delta 12 mexrenone to 11 alpha hydroxy delta 12 mexrenone; (j) testosterone to 11 alpha hydroxy 30 testosterone; and (k) progesterone to 11 alpha hydroxy progesterone. Preferably the enzyme does not catalyze the 15 beta hydroxylation of 3 keto delta 4,5 steroids; 3 keto delta 4, 5 delta 6, 7 steroids; or 3 keto delta 6, 7 steroids.

The invention also provides for a purified polypeptide, the amino acid 35 sequence of which is selected from the group consisting of SEQ ID NO: 23, SEQ ID NO: 24, SEQ ID NO: 25.

The invention provides for a purified immunogenic polypeptide, the amino acid sequence of which comprises at least ten consecutive residues of SEQ ID NO: 2.

5 The invention provides for an isolated and purified antibody having a binding specificity for 11 alpha hydroxylase having an amino acid sequence as shown in SEQ ID NO: 2. Preferably the antibody binds to a protein region selected from the group consisting of (a) the N-terminal amino acids 1-10 of SEQ ID NO: 2; (b) the last 10 C-terminal amino acids of SEQ ID NO: 2; (c) amino acids SEQ ID NO: 23; (d) amino acids SEQ ID NO: 24; and (e) amino acids SEQ ID NO: 25.
10 Preferably the antibody is purified on a peptide column, wherein said peptide is selected from the group consisting of: (a) the N-terminal amino acids 1-10 of SEQ ID NO: 2; (b) the last 10 C-terminal amino acids of SEQ ID NO: 2; (c) amino acids SEQ ID NO: 23; (d) amino acids SEQ ID NO: 24; and (e) amino acids SEQ ID NO: 25.

15 The invention also provides for a purified polypeptide, the amino acid sequence of which is selected from the group consisting of SEQ ID NO: 26.

The invention also provides for a purified immunogenic polypeptide, the amino acid sequence of which comprises at least ten consecutive residues of SEQ ID NO: 6.

20 The invention also provides for an isolated and purified antibody having a binding specificity for 11 alpha hydroxylase having an amino acid sequence as shown in SEQ ID NO: 6. Preferably the antibody binds to a protein region selected from the group consisting of (a) the N-terminal amino acids 1-10 of SEQ ID NO: 6; (b) the last 10 C-terminal amino acids of SEQ ID NO: 6; and (c) amino acids SEQ
25 ID NO: 26. More preferably, the antibody is purified on a peptide column, wherein said peptide is selected from the group consisting of: (a) the N-terminal amino acids 1-10 of SEQ ID NO: 6; (b) the last 10 C-terminal amino acids of SEQ ID NO: 6; and (c) amino acids SEQ ID NO: 26.

30 The invention also provides for a composition comprising an antibody described above in an effective carrier, vehicle, or auxiliary agent. It also provides for a composition comprising such an antibody and a solution. The antibody may be a polyclonal antibody. The antibody may also be a monoclonal antibody. The antibody may be conjugated to an immunoaffinity matrix. The invention also provides for a method of using an immunoaffinity matrix to purify a polypeptide from a biological fluid or cell lysate. Preferably the immunoaffinity matrix is
35

SEPHAROSE 4B. More preferably the method of using an immunoaffinity matrix to purify a polypeptide from a biological fluid or cell lysate uses SEPHAROSE 4B as an immunoaffinity matrix. More preferably, the method of using an immunoaffinity matrix to purify a polypeptide from a biological fluid or cell lysate. 5 uses SEPHAROSE 4B as an immunoaffinity matrix.

The invention also provides for a method of using a peptide column to purify an antibody, wherein said peptide is selected from the group consisting of: (a) the N-terminal amino acids 1-10 of SEQ ID NO: 2; (b) the last 10 C-terminal amino acids of SEQ ID NO: 2; (c) amino acids SEQ ID NO: 23; (d) amino acids SEQ 10 ID NO: 24; and (e) amino acids SEQ ID NO: 25.

The invention also provides for a method of using a peptide column to purify an antibody, wherein said peptide is selected from the group consisting of: (a) the N-terminal amino acids 1-10 of SEQ ID NO: 6; (b) the last 10 C-terminal amino acids of SEQ ID NO: 6; and (c) amino acids SEQ ID NO: 26.

15 The invention also provides for a method of detecting a first polypeptide in a biological fluid, wherein said first polypeptide is selected from the group consisting of 11 alpha hydroxylase and oxidoreductase, comprising the following steps: (a) contacting said fluid with a second polypeptide, having a binding specificity for said first polypeptide, and (b) assaying the presence of said second 20 polypeptide to determine the level of said first polypeptide. Preferably, the second polypeptide is an antibody. More preferably, the second polypeptide is radiolabeled.

25 The invention also provides for a process for producing an isolated nucleic acid comprising hybridizing SEQ ID NO: 1 to genomic DNA in 6XSSC and 65°C and isolating the nucleic acid detected with SEQ ID NO: 1. The invention also provides for an isolated DNA nucleic acid prepared according to this process.

The invention also provides for an isolated nucleic acid that specifically hybridizes under highly stringent conditions to the complement of the sequence set forth in SEQ ID NO: 1.

30 The invention also provides for a process for producing an isolated nucleic acid comprising hybridizing SEQ ID NO: 5 to genomic DNA in 6XSSC and 65°C and isolating the nucleic acid detected with SEQ ID NO: 5. The invention also provides for an isolated DNA nucleic acid prepared according to this process.

The invention also provides for an isolated nucleic acid that specifically hybridizes under highly stringent conditions to the complement of the sequence set forth in SEQ ID NO: 5.

5 The invention also provides for a DNA construct which alters the expression of a 11 alpha hydroxylase gene not normally expressed in a cell when said DNA construct is inserted into chromosomal DNA of the cell, said DNA construct comprising: (a) a targeting sequence; (b) a regulatory sequence; and (c) the structural gene for a steroid 11 alpha hydroxylase. The invention also provides for a host cell harboring this DNA construct.

10 The invention also provides for a DNA construct which alters the expression of a 11 alpha hydroxylase gene not normally expressed in a cell when said DNA construct is inserted into chromosomal DNA of the cell, said DNA construct comprising: (a) a targeting sequence; (b) a regulatory sequence; and (c) the structural gene for a steroid oxidoreductase. The invention also provides for a host cell harboring this DNA construct.

15 The invention also provides for use of a host cell harboring a cloned 11 alpha hydroxylase for the manufacture of a medicament for therapeutic application to treat heart disease, inflammation, arthritis, or cancer.

20 The invention also provides for a composition comprising from about 0.5-to about 500 g/L molasses, 0.5-50 g/L cornsteep liquid, 0.5-50 g/L KH₂PO₄, 2.5-250 g/L NaCl, 2.5-250 g/L glucose, and 0.04-4 g/L progesterone, pH 3.5-7. Preferably, this composition is comprised of from about 10-250 g/L molasses, 1-25 g/L cornsteep liquid, 1-25 g/L KH₂PO₄, 5-125 g/L NaCl, 5-125 g/L glucose, and 0.08-2 g/L progesterone, pH 4.5-6.5. More preferably, the composition is comprised of from 25 about 25-100 g/L molasses, 2.5-10 g/L cornsteep liquid, 2.5-10 g/L KH₂PO₄, 12.5-50 g/L NaCl, 12.5-50 g/L glucose, and 0.2-0.8 g/L progesterone, pH 5.5-6.0. Most preferably the composition comprises about 50 g/L molasses, 5 g/L cornsteep liquid, 5 g/L KH₂PO₄, 25 g/L NaCl, 25 g/L glucose, 20 g/L agar, and 0.4 g/L progesterone, pH 5.8.

30 The invention also provides for a semisolid formulation of any of the compositions described above, further comprising from about 4-100 g/L agar. Preferably the agar is at a concentration of from about 10-40 g/L agar. More preferably, the agar is about 20 g/L agar.

The invention also provides for the use of any of the compositions described above to produce spores from the microorganism selected from the group consisting of *Aspergillus ochraceus*, *Aspergillus niger*, *Aspergillus nidulans*, *Rhizopus oryzae*, *Rhizopus stolonifer*, and *Trichothecium roseum*, *Fusarium oxysporum*, *Rhizopus arrhizus*, *Monosporium olivaceum*, *Penicillium chrysogenum*, and *Absidia coerulea*.
5 Preferably, the composition is used to produce spores from *Aspergillus ochraceus*.

Definitions

The following is a list of abbreviations and the corresponding meanings as used interchangeably herein:

10 11 alpha hydroxycanrenone = 11 alpha hydroxy-4-androstene-3,17-dione
(C₂₂H₂₈O₄, MW 356.46)

AcNPV = *Autographa californica* nuclear polyhedrosis virus, a member of the Baculoviridae family of insect viruses

AD = androstanedione or 4-androstene-3,17-dione (C₂₂H₂₈O₃, MW 340.46)

15 aldadiene = canrenone

Amp = ampicillin

attTn7 = attachment site for Tn7 (a preferential site for Tn7 insertion into bacterial chromosomes)

bacmid = recombinant baculovirus shuttle vector isolated from *E. coli*

20 Bluo-gal = halogenated indolyl-β-D-galactoside

bp = base pair(s)

Cam = chloramphenicol

cDNA = complementary DNA

DMF = N,N-dimethylformamide

25 ds = double-stranded

eplerenone or epoxymexrenone = methyl hydrogen 9,11α-epoxy-17α-hydroxy-3-oxopregn-4-ene-7α,21-dicarboxylate, γ-lactone (MW 414.5)

g = gram(s)

Gen = gentamicin

30 hoxr = human oxidoreductase

HPLC = high performance liquid chromatography

hydroxycanrenone = 11 alpha- or 11 beta-hydroxycanrenone

IPTG = isopropyl-β-D-thiogalactopyranoside

Kan = kanamycin

35 kb = kilobase(s), 1000 bp(s)

mb = megabase(s)

Me = methyl
mg = milligram(s)
ml or mL = milliliter(s)
mm = millimeter
5 mM = millimolar
NMR = nuclear magnetic resonance
oxr = oxidoreductase
PCR = polymerase chain reaction
r = resistant or resistance
10 RP-HPLC = reverse phase high performance liquid chromatography
RT = room temperature
RT-PCR = reverse transcriptase polymerase chain reaction
s = sensitive
SDS-PAGE = sodium dodecyl sulfate polyacrylamide gel electrophoresis
15 Spc/Str = spectinomycin/streptomycin
Tet = tetracycline
Tn = transposon
ts = temperature-sensitive
U = units
20 ug or μ g = microgram(s)
ul or μ l = microliter(s)
X-gal = 5-bromo-3-chloro-indolyl- β -D-galactopyranoside
X-gluc = 5-bromo-3-chloro-indolyl- β -D-glucopyranoside

The following is a list definitions of various terms used herein:

25 The species "Aspergillus ochraceus NRRL 405" means the filamentous fungus *Aspergillus ochraceus* NRRL 405, accession number 18500, obtained from the American Type Culture Collection (ATCC). *A. ochraceus* NRRL 405 and *A. ochraceus* ATCC 18500 are the same strain, catalogued differently.

30 The term "amino acid(s)" means all naturally occurring L-amino acids, including norleucine, norvaline, homocysteine, and ornithine.

The term "degenerate" means that two nucleic acid molecules encode for the same amino acid sequences but comprise different nucleotide sequences.

The term "fragment" means a nucleic acid molecule whose sequence is shorter than the target or identified nucleic acid molecule and having the identical,

the substantial complement, or the substantial homologue of at least 10 contiguous nucleotides of the target or identified nucleic acid molecule.

The term "fusion protein" means a protein or fragment thereof that comprises one or more additional peptide regions not derived from that protein.

5 The term "probe" means an agent that is utilized to determine an attribute or feature (e.g. presence or absence, location, correlation, etc.) of a molecule, cell, tissue, or organism.

10 The term "promoter" is used in an expansive sense to refer to the regulatory sequence(s) that control mRNA production. Such sequences include RNA polymerase binding sites, enhancers, etc.

15 The term "protein fragment" means a peptide or polypeptide molecule whose amino acid sequence comprises a subset of the amino acid sequence of that protein.

15 The term "recombinant" means any agent (e.g., DNA, peptide, etc.), that is, or results from, however indirectly, human manipulation of a nucleic acid molecule.

15 The term "selectable or screenable marker genes" means genes whose expression can be detected by a probe as a means of identifying or selecting for transformed cells.

20 The term "specifically bind" means that the binding of an antibody or peptide is not competitively inhibited by the presence of non-related molecules.

15 The term "specifically hybridizing" means that two nucleic acid molecules are capable of forming an anti-parallel, double-stranded nucleic acid structure.

25 The term "substantial complement" means that a nucleic acid sequence shares at least 80% sequence identity with the complement.

15 The term "substantial fragment" means a nucleic acid fragment which comprises at least 100 nucleotides.

15 The term "substantial homologue" means that a nucleic acid molecule shares at least 80% sequence identity with another.

The term "substantially hybridizing" means that two nucleic acid molecules can form an anti-parallel, double-stranded nucleic acid structure under conditions (e.g., salt and temperature) that permit hybridization of sequences that exhibit 90% sequence identity or greater with each other and exhibit this identity for at least about a contiguous 50 nucleotides of the nucleic acid molecules.

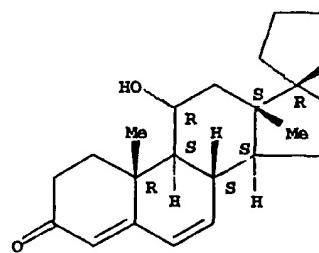
The term "substantially-purified" means that one or more molecules that are or may be present in a naturally-occurring preparation containing the target molecule will have been removed or reduced in concentration.

The following is a list of steroids, corresponding terms, and their structures, as used interchangeably herein:

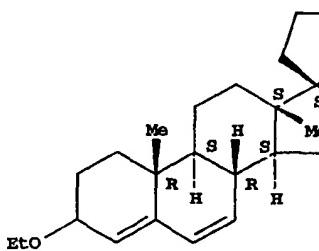
#	Name	CA Index Name:	Other Names	Formula	Structure
1	Eplerenone	Pregn-4-ene-7,21-dicarboxylic acid, 9,11-epoxy-17-hydroxy-3-oxo-, γ -lactone, methyl ester, (7 α ,11 α ,17 α)- (9CI)	Spiro[9,11-ene-7,21-epoxy-9H-cyclopenta[a]phenanthrene-17(2H),2'(3'H)-furan], pregn-4-ene-7,21-dicarboxylic acid deriv.; CGP 30083; Eplerenone; SC 66110	C24H30O6	
2	Aldadiene; Canrenone	Pregna-4,6-diene-21-carboxylic acid, 17-hydroxy-3-oxo-, γ -lactone, (17 α)-(9CI)	17 α -Pregna-4,6-diene-21-carboxylic acid, 17-hydroxy-3-oxo-, γ -lactone (6CI, 7CI, 8CI); Spiro[17H-cyclopenta[a]phenanthrene-17,2'(5'H)-furan], pregn-4,6-diene-21-carboxylic acid deriv.; 11614 R.P.; 17 β -Hydroxy-3-oxopregna-4,6-diene-21-carboxylic acid; 20-Spiroxa-4,6-diene-3,21-dione;	C22H28O3	

Aldadiene;
Canrenone;
Phanurane; SC
9376;
Spirolactone SC
14266

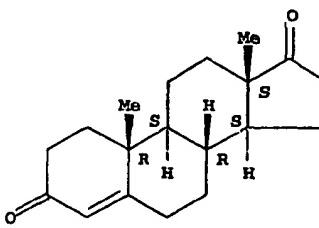
- 3 11 α -Hydroxycanrenone Pregna-4,6-diene-21-carboxylic acid, 11,17-dihydroxy-3-oxo-, γ -lactone, (11 α ,17 α)-(9CI) 11 α -Hydroxycanreno ne C22 H28 O4



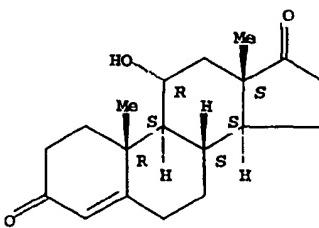
- 5 Aldona ethyl enol ether Pregna-4,6-diene-21-carboxylic acid, 3-ethoxy-17-hydroxy-, γ -lactone (9CI) Spiro[17H-cyclopenta[a]phenanthrene-17,2'(5'H)-furan], pregna-4,6-diene-21-carboxylic acid deriv.; Aldona ethyl enol ether C24 H34 O3



- 6 Androstenedione Androst-4-ene-3,17-dione (8CI, 9CI) $\Delta 4$ -Androstene-3,17-dione; 17-Ketotestosterone; 3,17-Dioxoandrost-4-ene; Androstenedione ; Fecundin; SKF 2170 C19 H26 O2



- 7 11 α -Hydroxyandrostenedione Androst-4-ene-3,17-dione, 11 α -hydroxy-, (11 α)-(9CI) Androst-4-ene-3,17-dione, 11 α -hydroxy- (8CI); 11 α -Hydroxyandrostenedione; 11 α -Hydroxyandrostenedione C19 H26 O3



8	Mexrenone	Pregn-4-ene-7,21-dicarboxylic acid, 17-hydroxy-3-oxo-, γ -lactone, methyl ester, (7 α ,17 α)-(9CI)	Spiro[17H-cyclopenta[a]phenanthrene-17,2'(5'H)-furan], pregn-4-ene-7,21-dicarboxylic acid deriv.; Mexrenone; SC 25152; ZK 32055	C24 H32 O5	
9	11 β -Hydroxymexrenone	Pregn-4-ene-7,21-dicarboxylic acid, 11,17-dihydroxy-3-oxo-, γ -lactone, methyl ester, (7 α ,11 β ,17 α)-(9CI)	11 β -Hydroxymexrenone	C24 H32 O6	
10	12 β -Hydroxymexrenone	Pregn-4-ene-7,21-dicarboxylic acid, 12,17-dihydroxy-3-oxo-, γ -lactone, methyl ester, (7 α ,12 β ,17 α)-(9CI)	12 β -Hydroxymexrenone	C24 H32 O6	
11	9 α -Hydroxymexrenone	Pregn-4-ene-7,21-dicarboxylic acid, 9,17-dihydroxy-3-oxo-, 21,17-lactone, 7-methyl ester, (7 α ,17 α)-(9CI)	9 α -Hydroxymexrenone	C24 H32 O6	

12	6β -Hydroxymexrenone	Pregn-4-ene-7,21-dicarboxylic acid, 6,17-dihydroxy-3-oxo-, γ -lactone, methyl ester, ($6\beta,7\alpha,17\alpha$)- (9CI)	Spiro[17H-cyclopenta[a]phenanthrene-17,2'(3'H)-furan], pregn-4-ene-7,21-dicarboxylic acid deriv.; 6β -Hydroxymexrone	C24 H32 O6	
13	Progesterone	Pregn-4-ene-3,20-dione (9CI)	Progesterone (8CI); $\Delta 4$ -Pregnene-3,20-dione; and >70 other names	C21 H30 O2	
14	Estr-4-ene-3,17-dione	Estr-4-ene-3,17-dione (6CI, 8CI, 9CI)	(+)-19-Norandrost-4-ene-3,17-dione; $\Delta 4$ -Estrone-3,17-dione; 19-Norandrost-4-ene-3,17-dione	C18 H24 O2	
15	delta 1,4-androstadiene-3,17-dione (ADD)	Androsta-1,4-diene-3,17-dione (7CI, 8CI, 9CI)	$\Delta 1,4$ -Androstadiene-3,17-dione; 1-Dehydroandrostenedione; Androstadienedione; Androstan-1,4-diene-3,17-dione	C19 H24 O2	
16	11 α -Hydroxyandrost-1,4-diene-3,17-dione (11 alpha-hydroxy) (11 α)-ADD	Androsta-1,4-diene-3,17-dione, 11 α -hydroxy-, (11 α)- (9CI)	Androsta-1,4-diene-3,17-dione, 11 α -hydroxy- (6CI, 7CI, 8CI); 11 α -Hydroxyandrost-1,4-diene-3,17-dione; Kurchinin	C19 H24 O3	
17	aldona		Compound 5 (aldona ethyl enol ether with O= in place of EtO- at position		

18	mexrenone 6,7-bislactone	Compound 12 with cyclic bis-lacto ring (-O-C=O-) formed between carbons at positions 6 and 7 (See U 5,981,744 for discussion of similar lactone rings)
19	11 alpha hydroxy mexrenone 6,7-bislactone	11 alpha hydroxy version of Compound 18
20	mexrenone 7,9-bislactone	Compound 11 with cyclic bis-lacto ring (-O-C=O-) formed between carbons at positions 7 and 9 (See U 5,981,744 for discussion of similar lactone rings)
21	11 alpha hydroxy mexrenone 7,9-bislactone	11 alpha hydroxy version of Compound 20

Figure 1 - Nucleotide and protein sequence of *Aspergillus ochraceus* 11 alpha hydroxylase

The nucleotide and protein sequences of *Aspergillus ochraceus* 11 alpha hydroxylase (SEQ ID NO: 1, SEQ ID NO: 2, respectively) are displayed.

Figure 2 - Nucleotide and protein sequence of human oxidoreductase

The nucleotide and protein sequences of human oxidoreductase (SEQ ID NO: 3, SEQ ID NO: 4, respectively) are displayed. The predicted amino acid sequence of human oxidoreductase independently cloned from a cDNA library prepared by RT-PCR using the RNA from a human HepG2 cells as a template, as disclosed in this specification, matches that previously reported by three different laboratories. The GenBank accession numbers for these loci include A60557 (NADPH--ferrihemoprotein reductase (EC 1.6.2.4) - human); AAG09798 (NADPH-cytochrome P450 reductase [Homo sapiens]), and P16435 (NADPH-CYTOCHROME P450 REDUCTASE (CPR) (P450R)).

The amino acid sequence of AAB21814 (cytochrome P450 reductase {EC 1.6.2.4} [human, placenta, Peptide Partial, 676 aa]), differs from human oxidoreductase A60557 and P16435 at 4 residues: A→V at 500, F→L at 518, V→W at 537, and A→H at 538. The initial methionine is also missing from AAB21814. The cognate nucleic acid for AA21814 (S90469 [cytochrome P450 reductase

[human, placenta, mRNA Partial, 2403 nt]) lacks the ATG codon for the initial methionine and includes a C→T change at 1496, a C→A, change at 1551, and a frameshift due to a missing G at 1605 which is resolved by the addition of a T at 1616.

5 References for these loci are as follows: A60557 [Yamano,S., Aoyama,T., McBride,O.W., Hardwick,J.P., Gelboin,H.V. and Gonzalez,F.J. Human NADPH-P450 oxidoreductase: complementary DNA cloning, sequence and vaccinia virus-mediated expression and localization of the CYPOR gene to chromosome 7 Mol. Pharmacol. 36 (1), 83-88 (1989)]; AAG09798 [Czerwinski,M., Sahni,M., Madan,A. and Parkinson,A. Polymorphism of human CYPOR: Expression of new allele. Unpublished, Direct Submission], and P16435 [Haniu,M., McManus,M.E., Birkett,D.J., Lee,T.D. and Shively,J.E. Structural and functional analysis of NADPH-cytochrome P-450 reductase from human liver: complete sequence of human enzyme and NADPH-binding sites. Biochemistry 28 (21), 8639-8645 (1989)]; AAB21814 [Shephard,E.A., Palmer,C.N., Segall,H.J. and Phillips,I.R. Quantification of cytochrome P450 reductase gene expression in human tissues. Arch. Biochem. Biophys. 294 (1), 168-172 (1992)]; S90469 [Shephard,E.A., Palmer,C.N., Segall,H.J. and Phillips,I.R. Quantification of cytochrome P450 reductase gene expression in human tissues. Arch. Biochem. Biophys. 294 (1), 168-172 (1992)].
10
15
20

Figure 3 - Nucleotide and protein sequence of *Aspergillus ochraceus* oxidoreductase

The nucleotide and protein sequences of *Aspergillus ochraceus* 11 oxidoreductase (SEQ ID NO: 5, SEQ ID NO: 6, respectively) are displayed.

25 **Figure 4 - Amino acid homology alignment of *A. ochraceus* 11 alpha hydroxylase with the top 10 BLAST hits from GenBank**

30 'Aspergillus ochraceus' steroid 11 alpha hydroxylase (SEQ ID NO: 02), cloned into plasmid pMON45624 (SEQ ID NO: 01), was aligned with related enzymes found in GenBank using the BLASTP program that implements a heuristic matching algorithm (Altschul et al., *J Mol Biol* Oct 5;215(3):403-10, 1990). The GenBank accession numbers (its probable function, [genus and species]) for the top 10 matches are as follows: CAA75565 (cytochrome P450 monooxygenase [*Gibberella fujikuroi*]); CAB91316 (probable cytochrome P450 monooxygenase (*lva*) [*Neurospora crassa*]); CAB56503 (cytochrome P450 [*Catharanthus roseus*]); AAB94588 (CYP71D10p [*Glycine max*]); CAA75566 (cytochrome P450
35

monooxygenase [*Gibberella fujikuroi*]); AAD34552 (cytochrome P450 monooxygenase [*Aspergillus terreus*]); CAA75567 (cytochrome P450 monooxygenase [*Gibberella fujikuroi*]); CAA76703 (cytochrome P450 [*Gibberella fujikuroi*]); CAA57874 (unnamed protein product [*Fusarium oxysporum*]));
5 CAA91268 (similar to cytochrome P450-cDNA EST yk423b11.3 comes from this gene [*Caenorhabditis elegans*]).

References for these loci are as follows: CAA75565 [Tudzynski,B. and Holter,K., Gibberellin biosynthetic pathway in *Gibberella fujikuroi*: evidence for a gene cluster. *Fungal Genet. Biol.* **25** (3), 157-170 (1998)]; CAB91316 [Schulte,U.,
10 Aign,V., Hoheisel,J., Brandt,P., Fartmann,B., Holland,R., Nyakatura,G., Mewes,H.W. and Mannhaupt,G., Unpublished]; CAB56503 [Schroeder,G., Unterbusch,E., Kaltenbach,M., Schmidt,J., Strack,D. and Schroeder,J. Light-induced cytochrome P450-dependent enzyme in indole alkaloid biosynthesis: tabersonine 16-hydroxylase *FEBS Lett.* **458**, 97-102 (1999)]; AAB94588
15 [Siminszky,B., Corbin,F.T., Ward,E.R., Fleischmann,T.J. and Dewey,R.E. Expression of a soybean cytochrome P450 monooxygenase cDNA in yeast and tobacco enhances the metabolism of phenylurea herbicides. *Proc. Natl. Acad. Sci. U.S.A.* **96** (4), 1750-1755 (1999)]; CAA75566 [Tudzynski,B. and Holter,K. Gibberellin biosynthetic pathway in *Gibberella fujikuroi*: evidence for a gene cluster. *Fungal Genet. Biol.* **25** (3), 157-170 (1998)]; AAD34552 [Kennedy,J., Auclair,K., Kendrew,S.G., Park,C., Vedera,J.C. and Hutchinson,C.R. Accessory Proteins Modulate Polyketide Synthase Activity During Lovastatin Biosynthesis. *Science* (1999) In press]; CAA75567 [Tudzynski,B. and Holter,K. Gibberellin biosynthetic pathway in *Gibberella fujikuroi*: evidence for a gene cluster. *Fungal Genet. Biol.* **25** (3), 157-170 (1998)]; CAA76703 [Tudzynski,B. and Hoelter,K. Characterization of P450 monooxygenase genes from *Gibberella fujikuroi*. Unpublished]; CAA57874 [Mouyna,I. and Brygoo,Y. Disruption of a *Fusarium oxysporum* f.sp. *elaeidis* cytochrome P450 gene by a repetitive sequence. Unpublished]; and CAA91268 [No Authors. Genome sequence of the nematode *C. elegans*: a platform for investigating biology. The *C. elegans* Sequencing Consortium. *Science* **282** (5396), 2012-2018 (1998) [Published errata appear in *Science* 1999 Jan 1;**283**(5398):35 and 1999 Mar 26;**283**(5410):2103 and 1999 Sep 3;**285**(5433):1493]].

Figure 5 – Phylogenetic tree showing the relatedness of *Aspergillus ochraceus* 11 alpha hydroxylase to the top 10 BLAST hits from GenBank

A phylogenetic tree displaying the genetic relatedness of *Aspergillus ochraceus* steroid 11 alpha hydroxylase, cloned into plasmid pMON45624, was aligned with related enzymes found in GenBank. BLAST was used to find the related enzymes within GenBank, and ClustalW was used generate the multiple sequence alignment and phylogenetic tree depicted in this figure. Descriptions of the GenBank accession numbers used as labels in the figure are the same as that described above for the legend to Figure 4.

10 Figure 6 – Percent homology between *Aspergillus ochraceus* 11 alpha hydroxylase and the top 10 BLAST hits from GenBank

The percent homology between *Aspergillus ochraceus* steroid 11 alpha hydroxylase and the top 10 enzymes found in GenBank using BLAST was calculated using CLUSTAL (Thompson et al., *Comput. Appl. Biosci.* 10:19-29, 15 1994).

Figure 7 - Amino acid homology alignment of *Aspergillus ochraceus* and human oxidoreductase to NADPH cytochrome P450 reductases from *A. niger*, mouse, and *S. cerevisiae*

The amino acid sequences of *Aspergillus ochraceus* steroid oxidoreductase (SEQ ID NO: 06) cloned into plasmid pMON45632 (SEQ ID NO: 05), and human oxidoreductase (SEQ ID NO: 03), cloned into plasmid pMON45605 (SEQ ID NO: 04) were aligned with related enzymes from *A. niger*, mouse, and *S. cerevisiae*, as described above. The GenBank accession numbers (probable function, [genus and species]) are as follows: BAA02936 (NADPH-cytochrome P450 reductase precursor [*Saccharomyces cerevisiae*]); CAA81550 NADPH cytochrome P450 oxidoreductase [*Aspergillus niger*]; P16435 (NADPH-CYTOCHROME P450 REDUCTASE (CPR) (P450R) [human]); BAA04496 (NADPH-cytochrome P450 oxidoreductase [*Mus musculus*]).

30 References for these loci are as follows: BAA02936 [Yabusaki,Y., Murakami,H. and Ohkawa,H. Primary structure of *Saccharomyces cerevisiae* NADPH-cytochrome P450 reductase deduced from nucleotide sequence of its cloned gene. *J. Biochem.* 103 (6), 1004-1010 (1988)]; CAA81550 [van den Brink,J., van Zeijl,C., van den Hondel,C. and van Gorcom,R. Cloning and characterization of the NADPH cytochrome P450 oxidoreductase (cprA) gene of *Aspergillus niger*.

Unpublished]; P16435 [Haniu,M., McManus,M.E., Birkett,D.J., Lee,T.D. and Shively,J.E. Structural and functional analysis of NADPH-cytochrome P-450 reductase from human liver: complete sequence of human enzyme and NADPH-binding sites *Biochemistry* 28 (21), 8639-8645 (1989)]; BAA04496 [Ohgiya,S.,
5 Shinriki,N., Kamataki,T. and Ishizaki,K. Mouse NADPH-cytochrome P-450 oxidoreductase: molecular cloning and functional expression in yeast. *Biochim. Biophys. Acta* 1186 (1-2), 137-141 (1994)].

10 **Figure 8 – Amino acid homology alignment of *A. ochraceus* oxidoreductase to NADPH cytochrome P450 reductases from *A. niger*, yeast, and *S. cerevisiae***

The amino acid sequence of *Aspergillus ochraceus* steroid oxidoreductase (SEQ ID NO: 06) cloned into plasmid pMON45632 (SEQ ID NO: 05), was aligned with related fungal enzymes from *A. niger* and *S. cerevisiae*, as described above. Descriptions of the GenBank accession numbers used as labels in the figure are the same as that described above for the legend to Figure 7, above.
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Figure 9 - Phylogenetic tree showing the relatedness of *Aspergillus ochraceus* and human oxidoreductase to reductases from *A. niger*, yeast, and mouse.

20 A phylogenetic tree displaying the genetic relatedness of *Aspergillus ochraceus* oxidoreductase (SEQ ID NO: 06), cloned into plasmid pMON45632 (SEQ ID NO: 05), was aligned with related enzymes. BLAST was used to find the related enzymes within GenBank, and ClustalW was used generate the multiple sequence alignment and phylogenetic tree depicted in this figure. Descriptions of the GenBank accession numbers used as labels in the figure are the same as that
25 described above for the legend to Figure 7, above.

Figure 10 - Percent identity between *Aspergillus ochraceus* oxidoreductase and reductases from *A. niger*, yeast, and mouse.

30 The percent identity between *Aspergillus ochraceus* oxidoreductase and the oxidoreductases from *A. niger*, yeast, and mouse was calculated using Clustal W and Boxshade.

Figure 11 - Alignment of human oxidoreductase with top 4 hits from SwissProt

The amino acid sequences of human steroid oxidoreductase (SEQ ID NO: 04), cloned into plasmid pMON45605 (SEQ ID NO: 03), which corresponds to the 5 amino acid sequence of the corrected sequence reported for P16435 below, was aligned with the top 4 hits from the SWISSPROT protein sequence database, as described above. The SWISSPROT accession numbers [locus] [common name] and species]) probable function) are as follows: P16435 {NCPR_HUMAN} [human] NADPH-CYTOCHROME P450 REDUCTASE; P00389 {NCPR_RABIT} [rabbit] NADPH-CYTOCHROME P450 REDUCTASE; P00388 {NCPR_RAT} [rat] NADPH-CYTOCHROME P450 REDUCTASE; P37040 {NCPR_MOUSE} [mouse] NADPH-CYTOCHROME P450 REDUCTASE; P04175 {NCPR_PIG} [pig] (NADPH-CYTOCHROME P450 REDUCTASE.

References for these loci are as follows: P16435 [Haniu,M., McManus,M.E., Birkett,D.J., Lee,T.D. and Shively,J.E. Structural and functional analysis of NADPH-cytochrome P-450 reductase from human liver: complete sequence of 15 human enzyme and NADPH-binding sites. *Biochemistry* 28 (21), 8639-8645 (1989)]; P00389 [Katagiri,M., Murakami,H., Yabasaki,Y., Sugiyama,T., Okamoto,M., Yamano,T. and Ohkawa,H. Molecular cloning and sequence analysis of full-length 20 cDNA for rabbit liver NADPH-cytochrome P-450 reductase mRNA. *J. Biochem.* 100 (4), 945-954 (1986)]; P00388 [Porter,T.D. and Kasper,C.B. Coding nucleotide sequence of rat NADPH-cytochrome P-450 oxidoreductase cDNA and identification of flavin-binding domains. *Proc. Natl. Acad. Sci. U.S.A.* 82 (4), 973-977 (1985)]; P37040 [Ohgiya,S., Shinriki,N., Kamataki,T. and Ishizaki,K. Mouse NADPH-cytochrome P-450 oxidoreductase: molecular cloning and functional expression in 25 yeast. *Biochim. Biophys. Acta* 1186 (1-2), 137-141 (1994)]; P04175 [Haniu,M., Iyanagi,T., Miller,P., Lee,T.D. and Shively,J.E. Complete amino acid sequence of NADPH-cytochrome P-450 reductase from porcine hepatic microsomes. *Biochemistry* 25 (24), 7906-7911 (1986)].

30 **Figure 12 - Phylogenetic tree showing the relatedness of human oxidoreductases with top 4 hits from SwissProt**

A phylogenetic tree displaying the genetic relatedness of human oxidoreductase (SEQ ID NO: 04), cloned into plasmid pMON45604 (SEQ ID NO: 03), was aligned with related enzymes found in SWISSPROT. BLAST was used to 35 find the related enzymes within SWISSPROT, and ClustalW was used generate

the multiple sequence alignment and phylogenetic tree depicted in this figure. Descriptions of the SWISPROT accession numbers used as labels in the figure are the same as that described above for the legend to Figure 11, above.

Figure 13 – Percent identity between human oxidoreductase and top 4 hits from SwissProt

The percent identity between human oxidoreductase and the top 4 hits found in SWISSPROT was calculated using Clustal W and Boxshade.

Figure 14: Expression of *Aspergillus ochraceus* 11 alpha hydroxylase in transfected Sf9 insect cells

Baculovirus-infected insect cells expressing *Aspergillus ochraceus* 11 alpha hydroxylase were harvested at 25 and 48 hours post infection and microsomal membrane fractions were prepared and separated by SDS-polyacrylamide gel electrophoresis. The proteins in the gel were electrophoretically transferred to 0.2 um nitrocellulose membrane (Schleicher & Schuell Grimsehlstrasse 23 37574 Einbeck Germany) and probed with antibodies GN-1187 and GN-1188 prepared from peptide 11aOH peptide 2 CRQILTPYIHKRKSLKGTTD (SEQ ID NO: 24).

Figure 15: Expression of *Aspergillus ochraceus* P450 oxidoreductase in transfected Sf9 insect cells

Baculovirus-infected insect cells expressing *Aspergillus ochraceus* 11 oxidoreductase were harvested at 25 and 48 hours post infection and microsomal membrane fractions were prepared and separated by SDS-polyacrylamide gel electrophoresis. The proteins in the gel were electrophoretically transferred to 0.2 um nitrocellulose membrane (Schleicher & Schuell Grimsehlstrasse 23 37574 Einbeck Germany) and probed with antibodies GN-2023 and GN-12024 prepared from oxr peptide 1 CTYWAVAKDPYASAGPAMNG (SEQ ID NO: 26).

Figure 16 – Conversion of androstenedione to 11 alpha hydroxy androstenedione monitored by HPLC

Microsomal and mitochondrial subcellular fractions were prepared from insect cells co-infected with recombinant baculoviruses expressing recombinant *Aspergillus ochraceus* 11 alpha hydroxylase and human oxidoreductase cloned from HepG2 cell RNA. The subcellular fractions were incubated with 250 µM androstenedione (AD) in the presence of an NADPH-generating system for 120

minutes, and the resulting products were separated by HPLC and monitored by ultraviolet detection at 247 nm. Hydroxylase activity was found in the microsomal fraction, as expected, but also appeared in the mitochondrial fraction. These results suggest that the 11 alpha hydroxylase may have a tendency to stick to membranes in disrupted cells, or that the separation of the subcellular fractions in this experiment was insufficient. Panel A illustrates a reaction carried out using enzyme prepared from a mitochondrial fraction. The peak in panel A that elutes after AD appears to be testosterone. When a microsomal fraction was used, almost as much AD was converted to 11 alpha hydroxy AD, but relatively more testosterone was also produced. Panel B illustrates the same reaction carried out for 120 minutes without a source of enzyme. Panel C illustrates an HPLC tracing with 11 α -hydroxyandrostenedione standard added to incubation buffer.

Detailed Description of the Invention

The present invention encompasses enzymes that facilitate the biosynthesis of steroid molecules, particularly enzymes possessing cytochrome P450 or oxidoreductase activities. The present invention is directed, in part, to the isolation of a nucleic acid encoding *Aspergillus ochraceus* 11 alpha hydroxylase, which exhibits sequence homology to the highly conserved residues that correspond to cytochrome P450 enzymes. It also directed to the isolation of nucleic acids encoding human and *Aspergillus ochraceus* oxidoreductase. Biological activities of the cloned hydroxylases and oxidoreductases of the present invention can be determined by a variety of assays, including incubation of steroid substrates in the presence of microsomes prepared from recombinant baculovirus-infected insect cells and monitoring the conversion to their 11 alpha hydroxy-counterparts by high pressure liquid chromatography (HPLC). The present invention, comprising novel 11 alpha hydroxylase and oxidoreductase nucleic acids, proteins, peptides, homologues, and fragments of either, provides new and advantageous methods to convert steroid intermediates to their 11 alpha hydroxy counterparts.

The present invention also includes the DNA sequences which code for the 11 alpha hydroxylases and oxidoreductases, DNA sequences which are substantially similar and perform substantially the same function, and DNA sequences which differ from the DNAs encoding the hydroxylases and oxidoreductases of the invention only due to the degeneracy of the genetic code. Also included in the present invention are the oligonucleotide intermediates used to construct mutated versions of these DNAs and the polypeptides encoded by these oligonucleotides and mutant DNAs.

The present invention also includes antibodies which bind specifically to *A. ochraceus* 11 alpha hydroxylase or *A. ochraceus* oxidoreductase, including anti-peptide antibodies, methods of using these anti-peptide antibodies to purify these and other related polypeptides, methods of using the purified polypeptides to generate polyclonal or monoclonal antibodies to the full-length polypeptides, and methods of using antibodies to the full-length polypeptides to assess the presence of the polypeptides in recombinant and non-recombinant host cells. The antibodies can be used to identify related polypeptides in any of a variety of host organisms that possess the biological activities associated with these polypeptides.

Among the preferred organisms that can be used in this hydroxylation step are *Aspergillus ochraceus* NRRL 405, *Aspergillus ochraceus* ATCC 18500, *Aspergillus niger* ATCC 16888 and ATCC 26693, *Aspergillus nidulans* ATCC 11267, *Rhizopus oryzae* ATCC 11145, *Rhizopus stolonifer* ATCC 6227b, *Streptomyces fradiae* ATCC 10745, *Bacillus megaterium* ATCC 14945, *Pseudomonas cruciviae* ATCC 13262, and *Trichothecium roseum* ATCC 12543. Other preferred organisms include *Fusarium oxysporum f. sp. cepae* ATCC 11171 and *Rhizopus arrhizus* ATCC 11145.

Other organisms that have exhibited activity for this reaction include *Absidia coerula* ATCC 6647, *Absidia glauca* ATCC 22752, *Actinomucor elegans* ATCC 6476, *Aspergillus flavipes* ATCC 1030, *Aspergillus fumigatus* ATCC 26934, *Beauveria bassiana* ATCC 7159 and ATCC 13144, *Botryosphaeria obtusa* IMI 038560, *Calonectria decora* ATCC 14767, *Chaetomium cochlioides* ATCC 10195, *Corynespora cassiicola* ATCC 16718, *Cunninghamella blakesleeana* ATCC 8688a, *Cunninghamella echinulata* ATCC 3655, *Cunninghamella elegans* ATCC 9245, *Curvularia clavata* ATCC 22921, *Curvularia lunata* ACTT 12071, *Cylindrocarpon radicicola* ATCC 1011, *Epicoccum humicola* ATCC 12722, *Gongronella butleri* ATCC 22822, *Hypomyces chrysospermus*, *Mortierella isabellina* ATCC 42613, *Mucor mucedo* ATCC 4605, *Mucor griseocyanus* ATCC 1207A, *Myrothecium verrucaria* ATCC 9095, *Nocardia corallina*, *Paecilomyces carneus* ATCC 46579, *Penicillium patulum* ATCC 24550, *Pithomyces atroolivaceus* IFO 6651, *Pithomyces cynodontis* ATCC 26150, *Pycnosporium* sp. ATCC 12231, *Saccharopolyspora erythrae* ATCC 11635, *Sepedonium chrysospermum* ATCC 13378, *Stachylidium bicolor* ATCC 12672, *Streptomyces hygroscopicus* ATCC 27438, *Streptomyces purpurascens* ATCC 25489, *Syncephalastrum racemosum* ATCC 18192, *Thamnostylum piriforme* ATCC 8992, *Thielavia terricola* ATCC 13807, and *Verticillium theobromae* ATCC 12474.

Additional organisms that may be expected to show activity for the 11α hydroxylation include *Cephalosporium aphidicola* (*Phytochemistry* (1996), 42(2), 411-415), *Cochliobolus lunatus* (*J. Biotechnol.* (1995), 42(2), 145-150), *Tieghemella orchidis* (*Khim.-Farm.Zh.* (1986), 20(7), 871- 876), *Tieghemella hyalospora* (*Khim.-Farm.Zh.* (1986), 20(7), 871-876), *Monosporium olivaceum* (*Acta Microbiol. Pol.*, Ser. B. (1973), 5(2), 103-110), *Aspergillus ustus* (*Acta Microbiol. Pol.*, Ser. B. (1973), 5(2), 103-110), *Fusarium graminearum* (*Acta Microbiol. Pol.*, Ser. B. (1973), 5(2), 103-110), *Verticillium glaucum* (*Acta Microbiol. Pol.*, Ser. B. (1973), 5(2), 103-110), and *Rhizopus nigricans* (*J. Steroid Biochem.* (1987), 28(2), 197-201).

Figure 1 sets forth the nucleotide and protein sequence of *Aspergillus ochraceus* 11 alpha hydroxylase (SEQ ID NO: 1, SEQ ID NO: 2, respectively). Figure 2 sets forth the nucleotide and protein sequence of human oxidoreductase (SEQ ID NO: 3, SEQ ID NO: 4, respectively). Figure 3 sets forth the nucleotide and protein sequence of *Aspergillus ochraceus* oxidoreductase (SEQ ID NO: 5, SEQ ID NO: 6, respectively).

Figure 4 sets forth an amino acid homology alignment of *A. ochraceus* 11 alpha hydroxylase cloned in pMON45624 and aligned with related enzymes found in GenBank using BLAST. Figure 5 is a phylogenetic tree showing the this relationship graphically. Figure 6 shows the percent homology between *Aspergillus ochraceus* steroid 11 alpha hydroxylase and the top 10 enzymes found in GenBank using BLAST, calculated using Clustal W and Boxshade.

Figure 7 sets forth the amino acid homology of *Aspergillus ochraceus* and human oxidoreductase to NADPH cytochrome P450 reductases from *A. niger*, mouse, and *S. cerevisiae* (yeast). Figure 8 sets forth the amino acid alignment for *A. ochraceus*, *A. niger*, and *S. cerevisiae* oxidoreductases. Figure 9 is a phylogenetic tree showing the relatedness of *Aspergillus ochraceus* and human oxidoreductase to reductases from *A. niger*, yeast, and mouse. Figure 10 shows the percent homology between *Aspergillus ochraceus* steroid 11 alpha hydroxylase and the oxidoreductases from *A. niger*, yeast, and mouse, calculated using Clustal W and Boxshade.

Figure 11 - Alignment of human oxidoreductase with top 4 hits from SwissProt. Figure 12 sets forth a phylogenetic tree displaying the genetic relatedness of human oxidoreductase, to these hits. Figure 13 shows the percent identity between human oxidoreductase and top 4 hits from SwissProt.

Figure 14 sets forth an immunoblot illustrating expression of *Aspergillus ochraceus* P450 11 alpha hydroxylase in baculovirus-infected insect cells harvested at 25 and 48 hours post infection. The nitrocellulose membrane was probed with a 1:1 mixture of antibodies prepared from two rabbits immunized with a conjugated synthetic peptide, 11aOH peptide 2 (SEQ ID NO 24).

Figure 15 sets forth an immunoblot illustrating expression of *Aspergillus ochraceus* P450 oxidoreductase in baculovirus-infected insect cells harvested at 25 and 48 hours post infection. The nitrocellulose membrane was probed with a 1:1 mixture of antibodies prepared two rabbits immunized with a conjugated synthetic peptide, oxr peptide 1 (SEQ ID NO 26).

Figure 16 sets forth an HPLC tracing illustrating the conversion of androstenedione (AD) to its 11 alpha hydroxy counterpart after incubating AD with subcellular fractions prepared from baculovirus-infected insect cells expressing *Aspergillus ochraceus* 11 alpha hydroxylase and human oxidoreductase.

15 *Cloning techniques*

Genetic engineering techniques now standard in the art (U.S. Patent 4,935,233 and Sambrook et al., "Molecular Cloning A Laboratory Manual", Cold Spring Harbor Laboratory, 1989) may be used in the construction of the DNA sequences of the present invention. One such method is cassette mutagenesis (Wells et al., *Gene* 34:315-323, 1985) in which a portion of the coding sequence in a plasmid is replaced with synthetic oligonucleotides that encode the desired amino acid substitutions in a portion of the gene between two restriction sites.

Pairs of complementary synthetic oligonucleotides encoding the desired gene can be made and annealed to each other. The DNA sequence of the oligonucleotide would encode sequence for amino acids of desired gene with the exception of those substituted and/or deleted from the sequence.

Plasmid DNA can be treated with the chosen restriction endonucleases then ligated to the annealed oligonucleotides. The ligated mixtures can be used to transform competent *E. coli* cells which will confer resistance to an appropriate antibiotic. Single colonies can be picked and the plasmid DNA examined by restriction analysis or by DNA sequencing to identify plasmids with the desired genes.

Cloning of DNA sequences encoding novel proteins and fusion proteins may be accomplished by the use of intermediate vectors. Linkers and adapters can be used to join DNA sequences, and to replace lost sequences, where a restriction site is internal to the region of interest. DNA encoding a single polypeptide or a fusion protein (comprising a first polypeptide, a peptide linker, and a second polypeptide) is inserted into a suitable expression vector which is then transformed or transfected into appropriate bacterial, fungal, insect, or mammalian host cells. The transformed organism or host cell line is grown and the recombinant protein isolated by standard techniques. Recombinant fusion proteins have all or a portion 10 of a first protein joined by a linker region to a all or a portion of second protein.

Hybridization

Nucleic acid molecules and fragment nucleic acid molecules encoding 11 alpha hydroxylases or oxidoreductases can specifically hybridize with other nucleic acid molecules. Two nucleic acid molecules are said to be capable of specifically hybridizing to one another if the two molecules are capable of forming an anti-parallel, double-stranded nucleic acid structure. A nucleic acid molecule is said to be the "complement" of another nucleic acid molecule, if they exhibit complete complementarity. Molecules exhibit "complete complementarity" when every nucleotide of one of the molecules is complementary to a nucleotide of the other. 15 Two molecules are "minimally complementary" if they can hybridize to one another with sufficient stability to permit them to remain annealed to one another under at least conventional "low-stringency" conditions. Similarly, the molecules are "complementary" if they can hybridize to one another with sufficient stability to permit them to remain annealed to one another under conventional "high- 20 stringency" conditions. Conventional stringency conditions are described by Sambrook, *et al.*, *Molecular Cloning, A Laboratory Manual*, 2nd Ed., Cold Spring Harbor Press, Cold Spring Harbor, New York (1989), and by Haymes, *et al.* *Nucleic Acid Hybridization, A Practical Approach*, IRL Press, Washington, DC, 1985). Departures from complete complementarity are therefore permissible, as long as 25 such departures do not completely preclude the capacity of the molecules to form a double-stranded structure.

Appropriate stringency conditions which promote DNA hybridization are well known to those skilled in the art, or can be found in *Current Protocols in Molecular Biology*, John Wiley & Sons, N.Y., 6.3.1-6.3.6, (1989). Basic conditions 30 would include, for example, 6X sodium saline citrate (SSC) at about 45°C, followed by a wash of 2X SSC at 50°C. Stringency can be varied, for example, by altering

- the salt concentration in the wash step from about 2X SSC at 50°C (moderately low stringency) to about 0.2X SSC at 50°C (high stringency). Stringency can also be altered by changing the temperature in the wash step, from room temperature, about 22°C (low stringency conditions), to about 65°C (high stringency conditions).
- 5 Both temperature and salt may be varied, or either the temperature or the salt concentration may be held constant while the other variable is changed.

Expression vectors

Another aspect of the present invention includes plasmid DNA vectors for use in the expression of these novel hydroxylases and oxidoreductases. These 10 vectors contain the novel DNA sequences described above which code for the novel polypeptides of the invention. Appropriate vectors which can transform microorganisms or cell lines capable of expressing the hydroxylases and 15 oxidoreductases include expression vectors comprising nucleotide sequences coding for the hydroxylases and oxidoreductases joined to transcriptional and translational regulatory sequences which are selected according to the host cells used.

Vectors incorporating modified sequences as described above are included 20 in the present invention and are useful in the production of the hydroxylases and oxidoreductases. The vector employed in the method also contains selected regulatory sequences in operative association with the DNA coding sequences of the invention and which are capable of directing the replication and expression thereof in selected host cells.

Methods for producing the hydroxylases and oxidoreductases is another 25 aspect of the present invention. The method of the present invention involves culturing suitable cells or cell lines, which has been transformed with a vector containing a DNA sequence encoding novel hydroxylases and oxidoreductases. Suitable cells or cell lines may be bacterial cells. For example, various strains of *E. coli* are well-known as host cells in the field of biotechnology. Examples of such strains include *E. coli* strains DH5 alpha, DH10B and MON105 (Obukowicz et al., 30 *Applied Environmental Microbiology* 58: 1511-1523, 1992). Also included in the present invention is the expression of the hydroxylases and oxidoreductases utilizing a chromosomal expression vector for *E. coli* based on the bacteriophage Mu (Weinberg et al., *Gene* 126: 25-33, 1993). Various other strains of bacteria, including the Enteric bacteria (e.g., *Salmonella* sp.) and *B. subtilis*, may also be 35 employed in this method.

When expressed in the *E. coli* cytoplasm, the gene encoding the proteins of the present invention may also be constructed such that at the 5' end of the gene codons are added to encode Met²-Ala¹, Met²-Ser¹, Met²-Cys¹, or Met¹ at the N-terminus of the protein. The N termini of proteins made in the cytoplasm of *E. coli* are affected by post-translational processing by methionine aminopeptidase (Ben Bassat et al., *J. Bacteriol.* 169:751-757, 1987), and possibly by other peptidases, so that upon expression the methionine is cleaved off the N-terminus. The proteins of the present invention may include polypeptides having Met¹, Ala¹, Ser¹, Cys¹, Met²-Ala¹, Met²-Ser¹, or Met²-Cys¹ at the N-terminus. These mutant proteins may also be expressed in *E. coli* by fusing a secretion signal peptide to the N-terminus. This signal peptide is cleaved from the polypeptide as part of the secretion process.

Yeast

Many strains of yeast cells known to those skilled in the art are also available as host cells for expression of the polypeptides of the present invention. Under another embodiment, the protein or fragment thereof of the present invention is expressed in a yeast cell, preferably *Saccharomyces cerevisiae*. The proteins or fragments thereof of the present invention can be expressed in *S. cerevisiae* by fusing it to the N-terminus of the URA3, CYC1 or ARG3 genes (Guarente and Ptashne, *Proc. Natl. Acad. Sci. (U.S.A.)* 78:2199-2203 (1981); Rose et al., *Proc. Natl. Acad. Sci. (U.S.A.)* 78:2460-2464 (1981); and Crabeel et al., *EMBO J.* 2:205-212 (1983)). Alternatively, proteins or fragments thereof of the present invention can be fused to either the PGK or TRP1 genes (Tuite et al., *EMBO J.* 1:603-608 (1982); and Dobson et al., *Nucleic Acids Res.* 11:2287-2302 (1983)). More preferably, the protein or fragment thereof of the present invention is expressed as a mature protein (Hitzeman et al., *Nature* 293:717-722 (1981); Valenzuela et al., *Nature* 298:347-350 (1982); and Deryck et al., *Nucleic Acids Res.* 11:1819-1837 (1983)).

Native and engineered yeast promoters suitable for use in the present invention have been reviewed by Romanos et al., *Yeast* 8:423-488 (1992). Most preferably, the protein or fragment thereof of the present invention is secreted by the yeast cell (Blobel and Dobberstein, *J. Cell Biol.* 67:835-851 (1975); Kurjan and Herskowitz, *Cell* 30:933-943 (1982); Bostian et al., *Cell* 36:741-751 (1984); Rothman and Orci, *Nature* 355:409-415 (1992); Julius et al., *Cell* 32:839-852 (1983); and Julius et al., *Cell* 36:309-318 (1984)).

Mammalian

General methods for expression of foreign genes in mammalian cells have been reviewed (Kaufman, R. J., 1987, "Genetic Engineering, Principles and Methods", Vol. 9, J. K. Setlow, editor, Plenum Press, New York; Colosimo et al., 5 *Biotechniques* 29: 314-331, 2000). Recombinant proteins are generally targeted to their natural locations within the host cell (e.g., cytoplasm, nucleus, or various membrane compartments), or are secreted, if a signal peptide is present. An expression vector is constructed in which a strong promoter capable of functioning in mammalian cells drives transcription of a eukaryotic secretion signal peptide coding region, which is translationally joined to the coding region for the desired protein. For example, plasmids such as pcDNA I/Neo, pRc/RSV, and pRc/CMV (obtained from Invitrogen Corp., San Diego, California) can be used. The 10 eukaryotic secretion signal peptide coding region can be from the gene itself or it can be from another secreted mammalian protein (Bayne, M. L. et al., *Proc. Natl. Acad. Sci. USA* 84: 2638-2642, 1987). After construction of the vector containing the gene, the vector DNA is transfected into mammalian cells such as the COS7, HeLa, BHK, Chinese hamster ovary (CHO), or mouse L lines. The cells can be cultured, for example, in DMEM media (JRH Scientific). The polypeptide secreted 15 into the media can be recovered by standard biochemical approaches following transient expression for 24 - 72 hours after transfection of the cells or after establishment of stable cell lines following selection for antibiotic resistance. The selection of suitable mammalian host cells and methods for transformation, culture, amplification, screening and product production and purification are known in the art. See, e.g., Gething and Sambrook, *Nature*, 293:620-625, 1981, or 20 alternatively, Kaufman et al, *Mol. Cell. Biol.*, 5(7):1750-1759, 1985) or Howley et al., and U.S. Pat. No. 4,419,446. Other suitable mammalian cell lines are the monkey COS-1 cell line and the CV-1 cell line.

Mammalian cells can also be used to express the nucleic acid molecules of the present invention. The nucleic acid molecules of the present invention can be 30 cloned into a suitable retroviral vector (see, e.g., Dunbar et al., *Blood* 85:3048-3057 (1995); Baum et al., *J. Hematother.* 5: 323-329 (1996); Bregni et al., *Blood* 80:1418-1422 (1992); Boris-Lawrie and Temin, *Curr. Opin. Genet. Dev.* 3:102-109 (1993); Boris-Lawrie and Temin, *Annal. New York Acad. Sci.* 716:59-71 (1994); Miller, *Current Top. Microbiol. Immunol.* 158:1-24 (1992)), adenovirus vector (Berkner, 35 *BioTechniques* 6:616-629 (1988); Berkner, *Current Top. Microbiol. Immunol.* 158:39-66 (1992); Brody and Crystal, *Annal. New York Acad. Sci.* 716:90-103 (1994); Baldwin et al., *Gene Ther.* 4:1142-1149 (1997)), RSV, MuSV, SSV, MuLV

(Baum *et al.*, *J. Hematother.* 5: 323-329 (1996)), AAV (Chen *et al.*, *Gene Ther.* 5:50-58 (1998); Hallek *et al.*, *Cytokines Mol. Ther.* 2: 69-79 (1996)), AEV, AMV, or CMV (Griffiths *et al.*, *Biochem. J.* 241: 313-324 (1987)).

Transformation and transfection

5 In another aspect, the invention provides a transformed cell having a nucleic acid molecule which comprises an exogenous promoter region which functions in a cell to cause the production of an mRNA molecule which is linked to a structural nucleic acid molecule, wherein the structural nucleic acid molecule encodes an 11 alpha hydroxylase or oxidoreductase gene or fragment thereof. This
10 nucleic acid molecule is linked to a 3' non-translated sequence that functions in a cell to cause termination of transcription and addition of polyadenylated ribonucleotides to a 3' end of the mRNA molecule.

15 Methods and compositions for transforming eukaryotic cells, bacteria and other microorganisms are known in the art (see, for example, Sambrook *et al.*, *Molecular Cloning: A Laboratory Manual*, Second Edition, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., (1989); Colosimo *et al.*, *Biotechniques* 29: 314-331, 2000).

20 Technology for introduction of DNA into cells is well known to those of skill in the art. Four general methods for delivering a gene into cells have been described: (1) chemical methods (Graham and van der Eb, *Virology* 54:536-539 (1973)); (2) physical methods such as microinjection (Capecchi, *Cell* 22:479-488 (1980)), electroporation (Wong and Neumann, *Biochem. Biophys. Res. Commun.* 107:584-587 (1982); Fromm *et al.*, *Proc. Natl. Acad. Sci. (U.S.A.)* 82:5824-5828 (1985); U.S. Patent No. 5,384,253); and the gene gun (Johnston and Tang, *Methods Cell Biol.* 43:353-365 (1994); (3) viral vectors (Clapp, *Clin. Perinatol.* 20:155-168 (1993); Lu *et al.*, *J. Exp. Med.* 178:2089-2096 (1993); Eglitis and Anderson, *Biotechniques*, 6:608-614 (1988)); and (4) receptor-mediated mechanisms (Curiel *et al.*, *Hum. Gen. Ther.* 3:147-154 (1992), Wagner *et al.*, *Proc. Natl. Acad. Sci. (U.S.A.)* 89:6099-6103 (1992)). Other methods well known in the art can also be used.
25

30 Transformation can be achieved using methods based on calcium phosphate precipitation, polyethylene glycol treatment, electroporation, and combinations of these treatments (see for example Potrykus *et al.*, *Mol. Gen. Genet.* 205:193-200 (1986); Lorz *et al.*, *Mol. Gen. Genet.* 199:178 (1985); Fromm *et al.*, *Nature* 319:791 (1986); Uchimiya *et al.*, *Mol. Gen. Genet.* 204:204 (1986); Marcotte *et al.*, *Nature* 335:454-457 (1988)).
35

Assays for gene expression based on the transient expression of cloned nucleic acid constructs have been developed by introducing the nucleic acid molecules into cells by polyethylene glycol treatment, electroporation, or particle bombardment (Marcotte *et al.*, *Nature* 335: 454-457 (1988); McCarty *et al.*, *Cell* 66: 5 895-905 (1991); Hattori *et al.*, *Genes Dev.* 6: 609-618 (1992); Goff *et al.*, *EMBO J.* 9: 2517-2522 (1990)). Transient expression systems may be used to functionally dissect the regulatory and structural features of expression cassettes comprising operably-linked genetic elements.

Insect Cell Expression

10 Insect cells may be used as host cells to express recombinant proteins of the present invention (See, e.g., Luckow, V.A., *Protein Eng.* J. L. Cleland., Wiley-Liss, New York, NY: 183-218, 1996, and references cited therein). General methods for expression of foreign genes in insect cells using baculovirus vectors have been described (O'Reilly, D.R., L.K. Miller *et al.* *Baculovirus Expression Vectors: A 15 Laboratory Manual*. New York, W.H. Freeman and Company, 1992; and King, L.A. and R.D. Possee, *The Baculovirus Expression System: A Laboratory Guide*, London, Chapman & Hall).

20 A baculovirus expression vector can be constructed by inserting the desired gene (e.g., 11 alpha hydroxylase or oxidoreductase) into a baculovirus transfer vector which can recombine into the baculovirus genome by homologous recombination. Many transfer vectors use a strong baculovirus promoter (such as the polyhedrin promoter) to drive transcription of the desired gene. Some vectors permit the expression of fusion proteins or direct the secretion of proteins from the 25 cell by fusing a eukaryotic secretion signal peptide coding region to the coding region of the desired gene. The plasmid pVL1393 (obtained from Invitrogen Corp., San Diego, California) can be used, for example, to direct transcription of nonfused foreign genes in baculovirus-infected insect cells. The baculovirus transfer vector containing the desired gene is transfected into *Spodoptera frugiperda* (Sf9) insect cells along with circular or linearized genomic baculovirus DNA, and recombinant 30 baculoviruses purified and amplified after one or more plaque assays.

Recombinant baculoviruses can also be created using the baculovirus shuttle vector system (Luckow, V.A. et al., *J. Virol.* 67(8): 4566-4579, 1993; U.S. Patent 5,348,886) now marketed as the Bac-To-Bac™ Expression System (Life Technologies, Inc., Rockville, MD). The desired genes are inserted downstream 35 from the polyhedrin promoter in mini-Tn7 cassettes that are transposed *in vivo*

into a baculovirus shuttle vector genome propagated in *E. coli*. Composite viral DNAs are isolated from *E. coli* and transfected into Sf9 cells and stocks of recombinant baculoviruses are rapidly prepared without the need for multiple rounds of tedious plaque purification common to methods that rely on homologous recombination.

Recombinant baculoviruses can also be created using the Gateway Recombinational Cloning System (Life Technologies) of shuttling genes from vector to vector using modified genetic elements (attachment sites) and modified proteins (e.g., int, IHF, xis) that are involved in the site-specific integration and excision of bacteriophage lambda.

Pure recombinant baculoviruses carrying the 11 alpha hydroxylase or oxidoreductase gene are used to infect cells cultured, for example, in Excell 401 serum-free medium (JRH Biosciences, Lenexa, Kansas) or Sf900-II (Life Technologies). Hydroxylases or oxidoreductases that are localized to membranes can be prepared using standard protocols that fractionate and enrich for enzymes in mitochondrial or microsomal fractions (Engel and White, *Dev Biol.* 140: 196-208, 1990). Hydroxylases or oxidoreductases that are secreted or leak into the medium can also be recovered by standard biochemical approaches.

Simultaneous expression of two or more recombinant proteins in baculovirus-infected insect cells can be carried out by two general approaches. The simplest approach is to coinfect insect cells with titered stocks of recombinant baculoviruses harboring a single heterologous gene under the control of a strong baculovirus promoter, such as the polyhedrin or the p10 promoter. These promoters are highly transcribed during the late stages of infection when most host cell protein synthesis has been shut down. Earlier baculovirus promoters or other insect or eukaryotic cell promoters can also be used to direct synthesis at other times, which generally result in lower expression levels. Varying the ratio of two or more recombinant viruses used in a coinfection or selecting viruses that use different promoters to drive expression of the recombinant protein will permit one skilled in the art to select conditions suitable for optimal expression of the desired recombinant proteins.

Construction of dual- or multiple-expression vectors will also permit the expression of two or more recombinant proteins in baculovirus-infected insect cells. Generally, these vectors permit the introduction two or more gene cassettes into a single locus in the baculovirus genome. The structures of a variety of dual

expression vectors have been described (O'Reilly, D. R., L. K. Miller et al. *Baculovirus Expression Vectors: A Laboratory Manual*. New York, W.H. Freeman and Company, 1992; and King, L. A. and R. D. Possee, *The Baculovirus Expression System: A Laboratory Guide*, London, Chapman & Hall).

5 **Materials and Methods**

General methods

General methods of cloning, expressing, and characterizing proteins are found in T. Maniatis, et al., *Molecular Cloning, A Laboratory Manual*, Cold Spring Harbor Laboratory, 1982, and references cited therein, incorporated herein by reference; and in J. Sambrook, et al., *Molecular Cloning, A Laboratory Manual*, 2nd edition, Cold Spring Harbor Laboratory, 1989, and references cited therein, incorporated herein by reference. General features and maps of a wide variety of cloning and expression vectors have been also been published (Gacesa, P. and Ramji, D.P., *Vectors: Essential Data*, John Wiley & Sons, 1994). General methods for the cloning and expression of genes in mammalian cells are also found in Colosimo et al., *Biotechniques* 29: 314-331, 2000. General and specific conditions and procedures for the construction, manipulation and isolation of polyclonal and monoclonal antibodies are well known in the art (See, for example, Harlow and Lane, *Antibodies: A Laboratory Manual*, Cold Spring Harbor Press, Cold Spring Harbor, New York, 1988).

Unless noted otherwise, all specialty chemicals were obtained from Sigma (St. Louis, MO). Restriction endonucleases and T4 DNA ligase were obtained from Life Technologies (Rockville, MD), New England Biolabs (Beverly, MA), Roche Molecular Biochemicals (Indianapolis, IN), or Promega (Madison, WI). All parts are by weight and temperatures are in degrees centigrade (°C), unless otherwise indicated.

Strains, plasmids, and sequence cross listings

The bacterial strains used in these studies are listed in Table 1. Plasmids used or constructed for this study are listed in Table 2. Brief descriptions of sequences of relevant oligonucleotides, genes, or proteins are listed in Table 3.

Table 1: Strains

Designation	Description or Genotype	Reference/Source
DH5α TM	F, <i>phi</i> 80 <i>dlacZdeltaM15</i> , delta(<i>lacZYA-argF</i>)U169, <i>deoR</i> , <i>recA1</i> , <i>endA1</i> , <i>hsdR17</i> (rk',mk'), <i>phoA</i> , <i>supE44</i> , <i>lambda-</i> , <i>thi-1</i> , <i>gyrA96</i> , <i>relA1</i>	Life Technologies, Rockville, Maryland
DH10B TM	F, <i>mcrA</i> D(<i>mrr-hsdRMS-mcrBC</i>) <i>phi</i> 80 <i>dlacZDM15</i> <i>DlacX74</i> <i>endA1</i> <i>recA1</i> <i>deoR</i> D(<i>ara, leu</i>)7697 <i>araD139</i> <i>galU</i> <i>galK</i> <i>nupG</i> <i>rpsL</i>	Life Technologies, Rockville, Maryland
DH10Bac TM	DH10B harboring the baculovirus shuttle vector bMON14272 (Kan ^R) and the helper plasmid pMON7124 (Tet ^R)	Life Technologies, Rockville, Maryland; See also Luckow et al., <i>J. Virol.</i> 67: 4566-4579 (1993)

Table 2: Plasmids

Plasmid	SEQ ID	Mark er NO.	Description	Source
pFastBac1		Amp ^R	Baculovirus donor plasmid containing multiple cloning site downstream of an AcNPV polyhedrin promoter within a mini-Tn7 transposable element capable of being transposed to a baculovirus shuttle vector	Life Technologies Inc. (Rockville, MD); See also Luckow et al., J. Virol. 67: 4566-4579 (1993)
pBluescript II SK		Amp ^R	Multifunctional phagemid cloning vector derived from pUC19.	Stratagene, La Jolla, CA
pCRII-TOPO		Amp ^R Kan ^R	Multifunctional cloning vector for direct cloning of polymerase chain reaction products using the T overhang	Invitrogen, Carlsbad, CA
pSport1		Amp ^R	Multifunctional cloning vector for cloning and in vitro transcription from either strand using SP6 or T7 promoters	Life Technologies, Rockville, MD
pGEM-T		Amp ^R	A derivative of pGEM-5Zf(+) with single 5' T overhangs at the insertion site to improve the efficiency of PCR product ligation	Promega, Madison, WI
pMON45624	#1	Amp ^R Gent ^R	pFastBac1 EcoRI/XbaI + PCR fragment EcoRI/XbaI encoding <i>Aspergillus ochraceus</i> 11 alpha hydroxylase	This work

pMON45603		Amp ^R	pBluescriptII SK <i>Bam</i> HI/ <i>Hinc</i> II + <i>Bam</i> HI/ <i>Hinc</i> II 5' segment of human oxidoreductase	This work
pMON45604		Amp ^R	pBluescriptII SK <i>Hinc</i> II/ <i>Kpn</i> I + <i>Hinc</i> II/ <i>Kpn</i> I 3' segment of human oxidoreductase	This work
pMON45605	#3	Amp ^R Gent ^R	pFastBac1 <i>Bam</i> HI/ <i>Kpn</i> I + <i>Bam</i> HI/ <i>Kpn</i> I complete coding region of human oxidoreductase cDNA.	This work
pMON45630		Amp ^R Kan ^R	pCRII-TOPO <i>Sal</i> I/ <i>Bam</i> HI + <i>Sal</i> I/ <i>Bam</i> HI 5' segment of <i>A.</i> <i>ochraceus</i> oxidoreductase cDNA	This work
pMON45631		Amp ^R Kan ^R	pCRII-TOPO <i>Bam</i> HI/ <i>Xho</i> I + <i>Bam</i> HI/ <i>Xho</i> I 3' segment of <i>A.</i> <i>ochraceus</i> oxidoreductase cDNA which lacked the intron.	This work
pMON45632	#5	Amp ^R Gent ^R	pFastBac1 <i>Sal</i> I/ <i>Xho</i> I + containing assembled coding region of <i>Aspergillus</i> <i>ochraceus</i> oxidoreductase	This work

Table 3: Table of Sequences

SEQ ID NO	Description	Length/Sequence	Type
(SEQ ID NO: 01)	Nucleotide sequence of <i>Aspergillus ochraceus</i> 11alphaOH gene from pMON45624	1776	DNA
(SEQ ID NO: 02)	<i>Aspergillus ochraceus</i> 11alphaOH protein sequence from pMON45624	514	Protein
(SEQ ID NO: 03)	Nucleotide sequence of human oxidoreductase gene from pMON45605	2031	DNA
(SEQ ID NO: 04)	Human oxidoreductase protein sequence from pMON45605	677	Protein
(SEQ ID NO: 05)	Nucleotide sequence of <i>Aspergillus ochraceus</i> oxidoreductase gene from pMON45632	2322	DNA
(SEQ ID NO: 06)	<i>Aspergillus ochraceus</i> oxidoreductase protein sequence from pMON45632	705	Protein
(SEQ ID NO: 07)	Primer H. oxred 1A	gatcggtatccaaatATGG GAGACTCCCACGTGGAC AC	DNA
(SEQ ID NO: 08)	Primer H. oxred 1B	CAGCTGGTTGACGAGAG CAGAG	DNA
(SEQ ID NO: 09)	Primer H. oxred 2A	CTCTGCTCTCGTCAACC AGCTG	DNA
(SEQ ID NO: 10)	Primer H. oxred 2B	gatcggtacaccttaGCTC CACACGTCCAGGGAGTA G	DNA
(SEQ ID NO: 11)	Primer A.oxred-for1	GACGGIGCIGGTACAAT GGA	DNA
(SEQ ID NO: 12)	Primer A.oxred-rev1	TTAIGACCAIAACATCIT CCTGGTAGC	DNA
(SEQ ID NO: 13)	Primer pSport-for1	CAAGCTCTAAATACGACT CACTATAGGGA	DNA
(SEQ ID NO: 14)	Primer A.oxred-rev2	CAGGAACCGATCGACCT CGGAA	DNA
(SEQ ID NO: 15)	Primer A.oxred-rev3	GTCACCCCTCACCAAGCAG AGCCAATG	DNA
(SEQ ID NO: 16)	Primer A.oxred-rev4	CCACATTGCGAACATA GCCTTGTTAGTG	DNA
(SEQ ID NO: 17)	Primer pSport-for2	GCCAAGCTCTAAATACGA CTCACTATAGGGAAAGC	DNA
(SEQ ID NO: 18)	Primer A.oxred-for2	gtcgacATGGCGCAAAT CGATACTCTC	DNA
(SEQ ID NO: 19)	Primer A.oxred-rev5	ctcgagttAGGACCAGA CATCGTCCCTGGTAG	DNA
(SEQ ID NO: 20)	Primer A.oxred-for3	GGATCCCTCGCGACCTG TGATCAT	DNA
(SEQ ID NO: 21)	Primer A.oxred-for4	CGAAAGATTCTTGTACA AGGATGAATGGAAGACT TTTC	DNA
(SEQ ID NO: 22)	Primer A.oxred-rev6	CTGAAAAGTCTTCCATT CATCCTTGATACAAGAAA TC	DNA
(SEQ ID NO: 23)	11aOH peptide 1	AAAYWLATLQPSDLPEL N	Protein
(SEQ ID NO: 24)	11aOH peptide 2	CRQILTPYIHKRKSLKG TTDE	Protein
(SEQ ID NO: 25)	11aOH peptide 3	HMGFGHGVHACPGRFFA	Protein

		SNEI	
(SEQ ID NO: 26)	oxr peptide 1	CTYWAVAKDPYASAGPA	Protein
	MNG		
(SEQ ID NO: 27)	CAA75565; cytochrome P450 monooxygenase [<i>Gibberella fujikuroi</i>]		Protein
(SEQ ID NO: 28)	CAB91316; probable cytochrome P450 monooxygenase (<i>lovA</i>) [<i>Neurospora crassa</i>]		Protein
(SEQ ID NO: 29)	CAB56503; cytochrome P450 [<i>Catharanthus roseus</i>]		Protein
(SEQ ID NO: 30)	AAB94588; CYP71D10p [<i>Glycine max</i>]		Protein
(SEQ ID NO: 31)	CAA75566; cytochrome P450 monooxygenase [<i>Gibberella fujikuroi</i>]		Protein
(SEQ ID NO: 32)	AAD34552; cytochrome P450 monooxygenase [<i>Aspergillus terreus</i>]		Protein
(SEQ ID NO: 33)	CAA75567; cytochrome P450 monooxygenase [<i>Gibberella fujikuroi</i>]		Protein
(SEQ ID NO: 34)	CAA76703; cytochrome P450 [<i>Gibberella fujikuroi</i>]		Protein
(SEQ ID NO: 35)	CAA57874; unnamed protein product [<i>Fusarium oxysporum</i>]		Protein
(SEQ ID NO: 36)	CAA91268; similar to cytochrome P450-cDNA EST yk423b11.3 comes from this gene; [<i>Caenorhabditis elegans</i>]		Protein
(SEQ ID NO: 37)	BAA02936 NADPH-cytochrome P450 reductase precursor [<i>Saccharomyces cerevisiae</i>]		Protein
(SEQ ID NO: 38)	CAA81550 NADPH cytochrome P450 oxidoreductase [<i>Aspergillus niger</i>]		Protein
(SEQ ID NO: 39)	BAA04496 NADPH-cytochrome P450 oxidoreductase [<i>Mus musculus</i>]		Protein
(SEQ ID NO: 40)	Universal bacteriophage M13 reverse primer	CAG GAA ACA GCT ATG AC	DNA
(SEQ ID NO: 41)	Universal bacteriophage T7 promoter primer	TAA TAC GAC TCA CTA TAG GG	DNA
(SEQ ID NO: 42)	Aspergillus ochraceus Primer 11alphaOH-for	gatcgaattcATGCCCT TCTTCACTGGGCT	DNA
(SEQ ID NO: 43)	Aspergillus ochraceus Primer 11alphaOH-rev	gatctctagattacaca gttaaactcgccatATTC GAT	DNA
(SEQ ID NO: 44)	pFastBac1 Primer Bacfwd	CTGTTTTCGTAACAGTT TTG	DNA
(SEQ ID NO: 45)	pFastBac1 Primer PolyA	CCTCTACAAATGTGGTA TG	DNA
(SEQ ID NO: 46)	Aspergillus ochraceus Primer 45624-for1	GAGATCAAGATTGCCTT	DNA
(SEQ ID NO: 47)	Aspergillus ochraceus Primer 45624-for2	CTTCGACGCTCTCAA	DNA
(SEQ ID NO: 48)	Aspergillus ochraceus Primer 45624-rev1	GCAATCTTGATCTCGTT	DNA
(SEQ ID NO: 49)	S90469 human cytochrome P450 reductase [placental, mRNA Partial, 2403 nt].	2403	DNA
(SEQ ID NO: 50)	AAB21814 human cytochrome P450 reductase, placental, partial	676	Protein
(SEQ ID NO: 51)	A60557 human NADPH-ferrihemoprotein reductase	677	Protein
(SEQ ID NO: 52)	P16435 Human NADPH-cytochrome P450 reductase	677	Protein

(SEQ ID NO: 53)	P00389 Rabbit NADPH-cytochrome P450 reductase	679	Protein
(SEQ ID NO: 54)	P00388 Rat NADPH-cytochrome P450 reductase	678	Protein
(SEQ ID NO: 55)	P37040 Mouse NADPH-cytochrome P450 reductase	678	Protein
(SEQ ID NO: 56)	P04175 Pig NADPH-cytochrome P450 reductase	678	Protein
(SEQ ID NO: 57)	Universal bacteriophage SP6 primer	gatttaggtgacactat ag	DNA
(SEQ ID NO: 58)	NotI-poly-dT adapter	5' - pGACTAGT <u>TCTAGA TCGCGA</u> <u>GCGGCCGC CC (T)₁₅</u> - 3'	DNA
(SEQ ID NO: 59)	Sall adapter, top strand	5' - TCGACCCACGCGTCGG - 3'	DNA
(SEQ ID NO: 60)	Sall adapter, bottom strand	3' - GGGTGCGCAGGCp - 5'	DNA
(SEQ ID NO: 61)	Primer oxred 1C	GTGGACCACAAGCTCGT ACTG	DNA
(SEQ ID NO: 62)	Primer oxred 2C	CATCGACCACCTGTGTG AGCTG	DNA
(SEQ ID NO: 63)	Primer oxred 2D	GTACAGGTAGTCCTCAT CCGAG	DNA
(SEQ ID NO: 64)	<i>Aspergillus niger</i> NADP CYP450 oxidoreductase Z26838	3710	DNA
(SEQ ID NO: 65)	<i>Aspergillus niger</i> NADP CYP450 oxidoreductase CAA81550	693	Protein

Specific Methods

Transformation of E. coli strains

5 *E. coli* strains such as DH5 alpha and DH10B (Life Technologies, Rockville, MD) are routinely used for transformation of ligation reactions and are the hosts used to prepare plasmid DNA for transfecting mammalian cells. *E. coli* strains, such as DH10B and MON105 (Obukowicz, et al., *Appl. and Envir. Micr.*, 58: 1511-1523, 1992) can be used for expressing the proteins of the present invention in the cytoplasm or periplasmic space.

10 DH10B and DH5alpha subcloning efficiency cells are purchased as competent cells and are ready for transformation using the manufacturer's protocol. Other *E. coli* strains are rendered competent to take up DNA using a CaCl₂ method. Typically, 20 to 50 mL of cells are grown in LB medium (1% Bacto-tryptone, 0.5% Bacto-yeast extract, 150 mM NaCl) to a density of approximately 1.0 absorbance unit at 600 nanometers (OD600) as measured by a Baush & Lomb Spectronic spectrophotometer (Rochester, NY). The cells are collected by

centrifugation and resuspended in one-fifth culture volume of CaCl_2 solution [50 mM CaCl_2 , 10 mM Tris-Cl ((10 mM 2-amino-2-(hydroxymethyl) 1,3-propanediol hydrochloride, pH 7.4] and are held at 4°C for 30 minutes. The cells are again collected by centrifugation and resuspended in one-tenth culture volume of CaCl_2 solution. Ligated DNA is added to 0.1 ml of these cells, and the samples are held at 4°C for 30-60 minutes. The samples are shifted to 42°C for 45 seconds and 1.0 ml of LB is added prior to shaking the samples at 37°C for one hour. Cells from these samples are spread on plates (LB medium plus 1.5% Bacto-agar) containing either ampicillin (100 micrograms/mL, ug/ml) when selecting for ampicillin-resistant transformants, or spectinomycin (75 ug/ml) when selecting for spectinomycin-resistant transformants. The plates are incubated overnight at 37°C. Colonies are picked and inoculated into LB plus appropriate antibiotic (100 ug/ml ampicillin or 75 ug/ml spectinomycin) and are grown at 37°C while shaking.

DNA isolation and characterization

Plasmid DNA can be isolated by a number of different methods and using commercially available kits known to those skilled in the art. Plasmid DNA is isolated using the Promega Wizard™ Miniprep kit (Madison, WI), the Qiagen QIAwell Plasmid isolation kits (Chatsworth, CA) or Qiagen Plasmid Midi or Mini kit. These kits follow the same general procedure for plasmid DNA isolation. Briefly, cells are pelleted by centrifugation (5000 x g), the plasmid DNA released with sequential NaOH/acid treatment, and cellular debris is removed by centrifugation (10000 x g). The supernatant (containing the plasmid DNA) is loaded onto a column containing a DNA-binding resin, the column is washed, and plasmid DNA eluted. After screening for the colonies with the plasmid of interest, the *E. coli* cells are inoculated into 50-100 ml of LB plus appropriate antibiotic for overnight growth at 37°C in an air incubator while shaking. The purified plasmid DNA is used for DNA sequencing, further restriction enzyme digestion, additional subcloning of DNA fragments and transfection into *E. coli*, mammalian cells, or other cell types.

DNA Sequencing protocols

Purified plasmid DNA is resuspended in dH₂O and its concentration is determined by measuring the absorbance at 260/280 nm in a Bausch and Lomb Spectronic 601 UV spectrometer. DNA samples are sequenced using ABI PRISM™ DyeDeoxy™ terminator sequencing chemistry (Applied Biosystems Division of Perkin Elmer Corporation, Lincoln City, CA) kits (Part Number 401388 or 402078)

according to the manufacturer's suggested protocol. Occasionally, 5% DMSO is added to the mixture in repeat experiments, to facilitate the sequencing of difficult templates.

Sequencing reactions are performed in a DNA thermal cycler (Perkin
5 Elmer Corporation, Norwalk, CT) following the recommended amplification conditions. Typically, DNA samples were prepared containing 500 ng of template DNA and 100 ng of primer of choice in thin-walled 0.2 mL PCR tubes that have been brought to 12 uL with Millipore milli-Q (mQ)-quality water. 2 uL of 2 mM Mg⁺ was added to each tube. Tubes were denatured for 5 minutes at 96°C in a
10 Perkin-Elmer System 9700 thermal cycler. After denaturation, the tubes were chilled to a temperature of 4°C by the thermal cycler. 6 uL of ABI Prism Big Dye Terminator Cycle Sequencing Ready Reaction Kit was added to each tube. The samples were returned to the thermal cycler and cycle-sequenced using the following program: (1) 96°C for 30 sec; (2) 50°C for 5 sec; (3) 60°C for 4 min,
15 followed by step (1) for 24 additional cycles and then held at 4°C. Cycle sequencing was complete after about 2.5 hours.

Samples are purified to remove excess dye terminators with using Centri-Sep™ spin columns (Princeton Separations, Adelphia, NJ) or purified through a
20 Millipore MAHV N45 50 Multiscreen-HV filtration plate which had been filled with 25 uL Sephadex G-50 superfine resin and 300 uL mQ water. Before loading samples onto filtration plates, the plate was prespun in a centrifuge at 750 x g for 2 min to remove excess water. The samples were loaded onto the resin and the plate spun again at 750 x g for 4 min. The purified sample was collected into a 96-well plate that was placed directly underneath the Sephadex-filled plate during the
25 spin. The liquid in the 96-well plate was dried at room temperature in a Speed Vac. After 45-60 min the DNA was dried and pelleted at the bottom of the plate. Samples were resuspended in 3 uL of a formamide/blue Dextran loading dye and were heated for 2 minutes (see p.33 of Perkin-Elmer Big Dye manual for loading
30 buffer recipe). Samples were loaded onto 48 cm well-to-read length 4.5% acrylamide gels and sequenced for 7 hr using ABI automated DNA sequencers (typically run module Seq Run 48E-1200 and dye set DT, Program BD, Set Any-Primer).

Overlapping DNA sequence fragments are analyzed and assembled into master DNA contigs using Sequencher DNA Analysis software (Gene Codes
35 Corporation, Ann Arbor, MI) or the Perkin-Elmer Data Collection and Sequence Analysis programs to assign bases to the data collected.

BLAST, ClustalW, and Boxshade homology alignment tools

A variety of programs can be used to align nucleotide or peptide sequences to each other and to facilitate homology searches in large sequence databases. BLAST (Basic Local Alignment Search Tool), which implements the statistical matching theory by Karlin and Altschul (*Proc. Natl. Acad. Sci. USA* 87: 2264-2268, 1990; *Proc. Natl. Acad. Sci. USA* 90: 5873-5877, 1993), is a widely used program for rapidly detecting ungapped nucleotide or peptide subsequences that match a given query sequence (Available from the National Center for Biotechnology Information, <http://www.ncbi.nlm.nih.gov>). BLAST uses a heuristic algorithm which seeks local 5 as opposed to global alignments and is therefore able to detect relationships among sequences which share only isolated regions of similarity (Altschul et al., *J. Mol. Biol.* 215: 403-410, 1990).

Two parameters can be varied which alter the sensitivity and quantity of BLAST search results. Parameter B (with a default value of 10) regulates the 15 number of high-scoring segment pairs (alignments) reported in the results. Parameter V (with a default value of 10) is the maximum number of database sequences (hits) for which one-line descriptions will be reported. Matches are based on high-scoring segment pairs (HSPs). Two sequences may share more than one HSP, if the HSPs are separated by gaps. The BLAST algorithm is sensitive to 20 ambiguities in the sequence and is not well-suited for sequences that contain many gaps.

The program blastp compares an amino acid query sequence against a protein sequence database. blastn compares a nucleotide query sequence against a nucleotide sequence database. blastx compares a nucleotide query sequence 25 translated in all reading frames against a protein sequence database. You could use this option to find potential translation products of an unknown nucleotide sequence. tblastn compares a protein query sequence against a nucleotide sequence database dynamically translated in all reading frames. tblastx compares the six-frame translations of a nucleotide query sequence against the six-frame 30 translations of a nucleotide sequence database (See <http://www.ncbi.nlm.nih.gov/Education/BLASTinfo/> for more information on BLAST, related programs, and pattern matching algorithms).

Nucleotides searches performed with BLAST, score = 98-557, word length 35 514 letters, were used to obtain nucleotide sequences homologous to nucleic acid molecules of the present invention. Protein searches are performed with BLASTP,

score = 50, word length = 3 to obtain amino acid sequences homologous to a reference polypeptide (e.g., SEQ ID NO: 2).

Clustal W version 1.74, which implements a different algorithm for alignment of multiple DNA or protein sequences, was also used to prepare 5 alignments and to assign percent identities between different sequences. This program improves the sensitivity of progressive multiple sequence alignment through sequence weighting, position specific gap penalties and weight matrix choice (Thompson et al., *Nucleic Acids Research*, 22(22):4673-4680, 1994). The default parameters for version 1.74 were used facilitate alignments and to assign 10 percent identities between two sequences. The input consisted of sequences in FASTA format and the output is the alignment shown in the figures. For nucleic acid sequences, the iub DNA weight matrix was used. For amino acid sequences, the blosum protein weight matrix was used (See <http://www.ncbi.nlm.nih.gov/Education/BLASTinfo/> for more information on BLAST, related programs, and 15 pattern matching algorithms.

Boxshade v 3.31 is a public domain program for creating nicely formatted printouts from muliple-aligned protein or DNA sequences. Boxshade, by itself, does not create alignments, but applies shading or coloring to files that were previously prepared by other sequence alignment programs. The inputs to 20 Boxshade are the alignments created by ClustalW and the threshold values for the residues to be colored or shaded. In most cases, except where specified, a 50% identity value was used. With this setting, if a position has greater than or equal to half of the sequences sharing an identical residue, then it is shaded. Boxshade is available by ftp from ftp. or by e-mail from Kay Hofmann (khofmann@isrec-sun1-unil.ch or Michael D. Baron (michael.baron@bbsrc.ac.uk).

Protein Purification and Characterization

Protein purification can be accomplished using any of a variety of chromatographic methods such as: ion exchange, gel filtration or hydrophobic chromatography or reversed phase HPLC. In some cases, proteins which are 30 properly folded can be affinity-purified using affinity reagents, such as monoclonal antibodies or receptor subunits attached to a suitable matrix. These and other protein purification methods are described in detail in *Methods in Enzymology*, Volume 182 "Guide to Protein Purification" edited by Murray Deutscher, Academic Press, San Diego, California, 1990.

The purified protein can be analyzed by RP-HPLC, electrospray mass spectrometry, and SDS-PAGE. Protein quantitation is done by amino acid composition, RP-HPLC, and/or Bradford protein dye-binding assays. In some cases, tryptic peptide mapping is performed in conjunction with electrospray mass spectrometry to confirm the identity of the protein.

Examples

The following examples will illustrate the invention in greater detail, although it will be understood that the invention is not limited to these specific examples. Various other examples will be apparent to the person skilled in the art after reading the present disclosure without departing from the spirit and scope of the invention. It is intended that all such other examples be included within the scope of the appended claims.

Example 1 - Preparation of *A. ochraceus* spores for RNA extraction

Aspergillus ochraceus ATCC 18500 stock culture (50 ul) was grown for 3-4 days on plates containing sporulation medium: 50 g/L molasses, 5 g/L cornsteep liquid, 5 g/L KH₂PO₄, 25 g/L NaCl, 25 g/L glucose, 20 g/L agar, and 0.4 g/L progesterone, pH 5.8. Progesterone was included in the media to induce the steroid 11 α-hydroxylase. Spores were scraped from the plates into 5 to 7 ml saline, washed in saline, collected by centrifugation, and suspended in saline containing 15% glycerol. The spores were frozen on dry ice and stored at -80°C. Approximately 0.8 g spores were incubated at 30°C in a 1 liter flask containing 400 ml 1% glucose, 50 mM KH₂PO₄, and 0.1 g canrenone, pH 7.0. This treatment prior to spore disruption has three benefits: (1) to induce the steroid 11 α-hydroxylase by incubation with canrenone; (2) to determine whether the spores were catalyzing the 11 α-hydroxylation of canrenone; (3) and to soften the spore wall. After approximately 26 hours of incubating with shaking at 30°C to provide better aeration, the spores were collected by centrifugation. Visual inspection with the aid of a microscope indicated that very few had started to germinate. The spore pellets were flash frozen in liquid nitrogen and stored at -80°C. The media was analyzed for presence of 11 alpha hydroxy canrenone by HPLC to determine whether spores used for library construction demonstrated the desired activity.

Example 2 - *A. ochraceus* spores catalyze 11 α-hydroxylation of canrenone

Approximately 160 ml of media from the spore induction was extracted three times with 70 ml ethyl acetate to collect the steroid substrate and products. The organic phase was dried over anhydrous magnesium sulfate, filtered, and evaporated to dryness. The residue was dissolved in 8 ml methanol so that the final concentration of canrenone was approximately 15 mM (assuming quantitative recovery). The media extract was diluted 10- to 15-fold into 50% methanol for HPLC analysis. Stock solutions of canrenone and 11 α-hydroxy canrenone were prepared in methanol. Standards for HPLC analysis were prepared from these stock solutions by diluting to a final concentration of 750 uM with 50% methanol. Media extract and standards were chromatographed over a C-4 reverse phase HPLC column. The media exhibited a component with the same retention time as the 11 α-hydroxy canrenone standard, as monitored at 254 nm (data not shown).

Example 3 - Growth of *A. ochraceus* Mycelia for RNA extraction

Liquid cultures of *Aspergillus ochraceus* mycelia were grown in 10 g/L peptone, 10 g/L yeast extract and 10 g/L glucose containing 20 g/L canrenone for 24 to 72 hours at 28°C in a volume of 160 ml. Ten ml samples of cells were filtered, washed with cold water, frozen, and stored at -80°C.

Example 4 - Extraction of total RNA from induced spores

Approximately 0.4 g spores were disrupted in 40 ml Trizol reagent (Life Technologies, Rockville, MD) using a Mini-Beadbeater™ model 3110 (Biospec Products, Bartlesville, OK). Briefly, spore-Trizol mixture was subjected to four 30 second pulses at low speed. Between pulses, tubes containing spores were chilled on ice. Visual inspection with the aid of a microscope indicated that the majority of the spores were disrupted by this treatment. The debris was pelleted by low-speed centrifugation and the total RNA in the supernatant was extracted following the manufacturer's recommended protocols for use with Trizol. Briefly, 2 ml chloroform was added for each 10 ml Trizol in 11 ml polypropylene centrifuge tubes. Following a 3 minute extraction of proteins, phase separation was done by centrifugation and the aqueous phase containing the RNA was transferred to a clean tube for precipitation with an equal volume of isopropanol. The precipitated RNA was recovered by centrifugation and washed with 70% ethanol. The RNA was resuspended in 10 ml water, re-extracted with chloroform and precipitated with ethanol overnight at -20°C. Total RNA (3 mg) was recovered by

centrifugation and rehydrated in 2 ml water, and precipitated on ice by adding an equal volume of cold 4 M lithium chloride. This precipitation was done to remove DNA, carbohydrates, heme, and other impurities which can carry over from guanidine methods. The RNA was recovered by a 25 minute centrifugation.

5 **Example 5 - Extraction of total RNA from induced mycelia**

Approximately 0.5 g wet weight cells were pulverized to a fine powder under liquid nitrogen with a mortar and pestle pre-chilled in dry ice. The powder was added to 10 ml Trizol Reagent (Life Technologies) and homogenized with a Kinematica polytron (Kinematica AG, Lucerne, Switzerland) at setting #4.
10 Cellular debris was removed by centrifugation prior to chloroform extraction. The aqueous phase containing nucleic acids was precipitated with isopropanol for 10 minutes at room temperature. The precipitate was collected by centrifugation and washed with 70% ethanol. The RNA was rehydrated in water and re-extracted with chloroform to remove any residual proteins. The aqueous phase was
15 precipitated at -20°C with 1/10 volume of 3 M sodium acetate and 2.5 volumes absolute ethanol. The final yield was 424 ug. Approximately 4 ug and 16 ug of total RNA were separated by electrophoresis through a 1.2% agarose gel and visualized by staining in ethidium bromide. Chromosomal DNA was present as a minor contaminant.

20 **Example 6 - Extraction of Total RNA from HepG2 cells**

Hepatocellular human liver carcinoma cells (HepG2), ATCC HB-8065, were maintained in DMEM high glucose media supplemented with Penstrep, glutamate and 10% fetal bovine serum (Life Technologies, Rockville, MD). Cells were induced overnight with 0.05% ethanol and harvested for RNA extraction by trypsinization.
25 Briefly, the cell pellet was resuspended in >10X volumes of 4 M guanidine isothiocyanate, 50 mM Tris-HCl, pH 7.5, 25 mM EDTA (solution D, Life Technologies) and then vortexed. Water and sodium acetate, pH 4.1, were added such that the final concentration of sodium acetate was 0.1 M. The RNA solution was extracted with one half volume of chloroform and placed on ice for 15 minutes.
30 The aqueous phase was re-extracted with chloroform and precipitated overnight with isopropanol. Total RNA was resuspended in solution D and re-precipitated with isopropanol, followed by two precipitations in water containing 0.3 M sodium acetate pH 5.5 and 2.5 volumes of ethanol. PolyA⁺ selection was performed twice as described below.

Example 7 - PolyA⁺ Selection of mRNA

PolyA⁺ RNA was selected from total RNA with an Eppendorf 5Prime, Inc. kit (Boulder CO). Briefly, each 1 mg of total RNA was selected twice over a column containing oligo dT cellulose. The column slurry was packed by gentle centrifugation and equilibrated with 0.5 M NaCl. RNA was allowed to bind to the dT cellulose for 15 minutes at room temperature. The columns were washed once with 0.5 M NaCl, and twice with 0.1 M NaCl. PolyA⁺ RNA was eluted in 0.5 ml 10 mM Tris-HCl, 1 mM EDTA, pH 7.5. The selection by oligo dT cellulose was performed twice. The mRNA was precipitated at -20°C with 0.3 M sodium acetate in 50% ethanol, with glycogen added as carrier.

Example 8 - cDNA Synthesis and Library Construction

The Superscript™ Plasmid System for cDNA Synthesis and Plasmid Cloning kit (Life Technologies) was used for cDNA synthesis and library construction. Superscript II reverse transcriptase catalyzed the first strand of cDNA in a 20 ul reaction for 1 hour at 42°C. The final composition was 50 mM Tris-HCl, pH 8.3, 75 mM KCl, 3 mM MgCl₂, 10 mM DTT, 50 uM each dATP, dCTP, dGTP and dTTP, 50 ug/ml oligo-dT-*NotI* primer-adaptors that were phosphorylated at their 5' end (Life Technologies) and 50,000 units/ml Superscript II reverse transcriptase.

20

oligo-dT-*NotI* primer-adapter
5' - pGACTAGT TCTAGA TGGCGA GCGGCCGC CC (T)_n - 3' (SEQ ID NO: 58)
SpeI *XbaI* *NruI* *NotI*

25

A radiolabeled tracer ([α -³²P]dCTP) was not added. The second strand of cDNA was synthesized in a reaction volume of 150 ul. The final composition of this mixture including the first strand reaction was 25 mM Tris-HCl, pH 7.5, 100 mM KCl, 5 mM (NH₄)₂SO₄, 0.15 mM B-NAD⁺, 250 uM each dATP, dCTP, dGTP and dTTP, 1.2 mM DTT, 65 units/ml *E. coli* DNA ligase, 250 units/ml *E. coli* DNA polymerase I and 13 units/ml *E. coli* Rnase H. After a 2 hour incubation at 16°C, 30 10 units of T4 DNA polymerase was added, and incubated 5 minutes at 16°C. The reaction was stopped with 10 ul 0.5 M EDTA and the cDNA was separated from cDNAs smaller than 300 base pairs, primer-adaptors and deoxynucleotides with GENECLEAN II (BIO 101 Inc. La Jolla, CA). Annealed *Sal I* adaptors (Life Technologies) that were phosphorylated at their 5' blunt end were ligated to the cDNA overnight at 16°C.

35 *SalI* adapter

5' - TCGACCCACGCGTCCG - 3' (SEQ ID NO: 59)
3' - GGGTGCGCAGGCCp - 5' (SEQ ID NO: 60)

GENECLEAN II was used to remove the adaptors. The cDNA was then digested with *NotI*. QIAquick columns (QIAGEN, Valencia, CA) were used to remove small
5 DNA fragments from the cDNA, which was ethanol precipitated.

Example 9 - Size Fractionation of cDNA

The cDNA was enriched for species approximately 1.5 kb and larger by gel electrophoresis through 0.8% Sea-Plaque agarose (FMC BioProducts, Rockland ME) in TAE buffer. The preparative gel had a lane of DNA size markers which
10 was excised from the gel after electrophoresis and stained with ethidium bromide for visualization under ultraviolet light next to a ruler so that the appropriate region of the cDNA could be recovered from the gel. GENECLEAN II was used to extract the cDNA, which was eluted in 20 ul water.

15 **Example 10 - Library Construction in Vector pSport1 and Electroporation into E. coli**

An aliquot of the size-selected cDNA was ligated overnight at 4°C with pSport1 (Life Technologies, Inc., Rockville, MD) predigested with *NotI* and *Sall* in a 20 ul reaction containing 50 mM Tris-HCl, pH 7.6, 10 mM MgCl₂, 1 mM ATP, 5% (w/v) PEG 8000, 1 mM DTT, 2.5 ug/ml pSport1, approximately 0.5 ug/ml cDNA,
20 and 50 units/ml T4 DNA ligase. The ligation mixture was precipitated by the addition of 12.5 ul 7.5 M ammonium acetate, 5 ul yeast tRNA carrier and 70 ul absolute ethanol. The ligated cDNA was recovered by centrifugation at room temperature for 20 minutes and rehydrated in 5 ul sterile water. One ul of the ligated cDNA was introduced into ElectroMAX DH10B *E. coli* (Life Technologies)
25 by electroporation. Cells were allowed to recover in 1 ml SOC medium (Life Technologies) for 1 hour at 37°C, before plating an aliquot on LB with 100 ug/ml ampicillin. The titer of the *Aspergillus ochraceus* spore library (designated LIB3025) was determined by preparing serial dilutions of the cell suspension in SOC. The equivalent of 1 ul, 0.1 ul and 0.01 ul samples of the cell suspension were
30 plated, and the resulting titer was calculated to be 1.75 x 10⁶/ml colony forming units.

Example 11 - Identification of clones encoding cytochrome P450 enzymes by DNA sequence analysis and construction of plasmid pMON45624 encoding *Aspergillus ochraceus* 11 alpha hydroxylase

Cloning of 11 alpha hydroxylase from Aspergillus ochraceus

5 Approximately 2,000 colonies were selected on LB agar plates containing 100 ug/ml ampicillin and miniprep plasmid DNA samples were prepared for sequencing. Unidirectional sequencing was performed from the 3' end of the expressed sequence tags (ESTs) beginning at the *NoI* site encompassing part of the poly dT primer used for cDNA synthesis. Two universal primers were used to
10 facilitate the sequencing:

M13 reverse: CAG GAA ACA GCT ATG AC (SEQ ID NO: 40)
T7 promoter: TAA TAC GAC TCA CTA TAG GG (SEQ ID NO: 41)

15 Most known cytochrome p450s contain a conserved heme-binding region approximately 50 amino acid residues (150 nucleotides) upstream of the stop codon (Nelson *et al*, *Pharmacogenetics* 6: 1-42, 1996). The 2,000 ESTs were screened for sequences encoding the canonical heme-binding motif (FXXGXXXCXG, where "X" is any amino acid) in the appropriate region using BLASTX and visual inspection of the sequences scored as hydroxylases for the canonical heme-binding motif.
20 Only fifteen ESTs had the heme-binding motif. One EST was unique and the other fourteen appeared to be overlapping sequences. The cDNA inserts from seven clones encoding putative cytochrome p450 enzymes were then sequenced to completion. All seven encoded the same enzyme.

Gene Amplification of Aspergillus ochraceus 11 alpha hydroxylase

25 The coding region of the 11 alpha hydroxylase was amplified by PCR using a unique clone from the *A. ochraceus* cDNA spore library (LIB3025) as a template. The primers included recognition sites for *EcoRI* (forward) and *XbaI* (reverse) for directional cloning into pFastbac1. Amplification was carried out for 32 cycles using a PCR core kit (Roche) and 50 pmol of each primer. One cycle consisted of a
30 denaturation step at 94°C for 45 seconds, an annealing step at 60°C for 45 seconds, and an elongation step at 72°C for 60 seconds.

Primer 11alphaOH-for: gatcgaaattcatGCCCTTCTTCACTGGGCT (SEQ ID NO: 42)
Primer 11alphaOH-rev: gatctctagattACACAGTTAAACTCGCCATATCGAT (SEQ ID NO: 43)

Construction of pMON45624

The amplified fragments described above were purified through a QIAquick column (Qiagen, Valencia CA) and digested with *Eco*RI and *Xba*I prior to ligation into pFastBac1 cleaved with *Eco*RI and *Xba*I. The resulting plasmid was 5 designated pMON45624 and the DNA sequence verified using primers based on the vector sequence and internal primers based on the 11 alpha hydroxylase sequence (shown below).

10 Primer Bacfwd: CTGTTTCGTAACAGTTTG (SEQ ID NO: 44)
Primer PolyA: CCTCTACAAATGTGGTATG (SEQ ID NO: 45)
Primer 45624-for1: GAGATCAAGATTGCCTT (SEQ ID NO: 46)
Primer 45624-for2: CTTCGACGCTCTCAA (SEQ ID NO: 47)
Primer 45624-rev1: GCAATCTTGATCTCGTT (SEQ ID NO: 48)

15 The nucleotide and predicted amino acid sequences of the cloned 11 alpha hydroxylase are displayed in Figure 1 as SEQ ID NO: 1 and SEQ ID NO: 2, respectively.

20 Figure 4 sets forth an amino acid homology alignment of *A. ochraceus* 11 alpha hydroxylase cloned in pMON45624 and aligned with related enzymes found in GenBank using BLAST. Figure 5 is a phylogenetic tree showing the this relationship graphically. Figure 6 shows the percent homology between *Aspergillus ochraceus* steroid 11 alpha hydroxylase and the top 10 enzymes found in GenBank using BLAST, calculated using Clustal W and Boxshade.

25 **Example 12 - Amplification of cDNA encoding human NADPH Cytochrome P450 reductase and cloning into plasmids pMON45603, pMON45604, and pMON45605**

Gene Amplification of human oxidoreductase

Approximately 1 ug polyA⁺ mRNA from HepG2 cells was heated to 65°C for 10 minutes with 100 ng random hexamers (Invitrogen, Carlsbad, CA) in an 11 ul reaction. The mixture was chilled on ice, then incubated at 42°C for 75 minutes in 30 a 20 ul reaction containing 1 ul RNase inhibitor (Promega, Madison, WI), 0.01 M DTT, 5 mM dNTPs, 50 mM Tris-HCl, pH 8.3, 75 mM KCl, 3 mM MgCl₂, and 1 ul SuperScriptII enzyme (Life Technologies). The reverse transcriptase was inactivated by heating to 95°C for 2 minutes. First strand cDNA was stored at -20°C. Forward and reverse primers were based on the nucleotide sequence of 35 accession number S90469 (human placental partial mRNA encoding cytochrome P450 reductase (SEQ ID NO: 49)). The accession number of the corresponding

protein sequence is AAB21814 (SEQ ID NO: 50). The human oxidoreductase was cloned in two pieces which were assembled in pFastBac1 (Life Technologies) by ligation at an internal *Hinc*II site. The primers included restriction sites for directional subcloning into pFastBac1.

5

Primer H. oxred 1A: gatcggtatccaatATGGGAGACTCCCACGTGGACAC (SEQ ID NO: 07)
Primer H. oxred 1B: CAGCTGGTTGACGAGAGCAGAG (SEQ ID NO: 08)
Primer H. oxred 2A: CTCTGCTCTCGTCAACCAGCTG (SEQ ID NO: 09)
Primer H. oxred 2B: gatcggtaccttagCTCCACAGTCCAGGGAGTAG (SEQ ID NO: 10)

- 10 The second strand was synthesized using 400 uM dNTP and 167 nM of each primer set per 150 ul reaction. Amplification was performed with Deep Vent polymerase (New England Biolabs, Beverly, MA). The reaction for segment 2 (the 3' half of the oxidoreductase cDNA) was adjusted to 5% DMSO. The amplification included an initial cycle of denaturation at 94°C for 90 seconds, followed by annealing at 62°C
15 for 2 minutes and elongation at 72°C for 2 minutes. This was followed by 30 cycles, consisting of a 45 second denaturation step, a 45 second annealing step, and a 60 second elongation step. The elongation step was extended to 5 minutes for the final cycle.

Construction of pMON45603, pMON45604, pMON45605

- 20 The PCR fragments for the 5' half of the oxidoreductase cDNA were digested with *Bam*HI and *Hinc*II. The PCR fragments for the 3' half of the oxidoreductase cDNA were digested with *Hinc*II and *Kpn*I and ligated into pBluescript II (Stratagene, La Jolla, CA) for sequencing. The resulting plasmids were designated pMON45603 (5' segment) and pMON45604 (3' segment). The
25 *Bam*HI/*Hinc*II fragment from pMON45603 and the *Hinc*II/*Kpn*I fragment from pMON45604 were ligated into pFastbac1 cut with *Bam*HI and *Kpn*I, to generate pMON45605.

- Sequencing primers were based on the sequence of GenBank accession number S90469 (SEQ ID NO 49), a cDNA encoding cytochrome P450 reductase [human, placenta, mRNA Partial, 2403 nt]. The cognate protein sequence is:
30 AAB21814 (SEQ ID NO 50) cytochrome P450 reductase {EC 1.6.2.4} [human, placenta, Peptide Partial, 676 aa] [Homo sapiens]. The cDNA insert of pMON45603 was sequenced using primer oxred 1C, and the cDNA insert of pMON45604 was sequenced using primer oxred 2C and 2D. Universal T7 (SEQ ID NO: 41) and M13 reverse (SEQ ID NO: 40) primers, which annealed to vector sequences flanking the cDNA inserts were also used for sequencing.
35

Primer oxred 1C: GTGGACCACAAGCTCGTACTG (SEQ ID NO: 61)

Primer oxred 2C: CATCGACCACCTGTGTGAGCTG (SEQ ID NO: 62)
Primer oxred 2D: GTACAGGTAGTCCTCATCCGAG (SEQ ID NO: 63)

The nucleotide and predicted amino acid sequences of the cloned human oxidoreductase are displayed in Figure 2 as SEQ ID NO: 3 and SEQ ID NO: 4, 5 respectively. Figure 11 sets forth an alignment of human oxidoreductase with top 4 hits from SwissProt. Figure 12 sets forth a phylogenetic tree displaying the genetic relatedness of human oxidoreductase, to these hits. Figure 13 shows the percent identity between human oxidoreductase and top 4 hits from SwissProt.

Example 13 - Amplification of cDNA encoding NADPH cytochrome P450 reductase from *A. ochraceus* and cloning into plasmids pMON45630, 10 pMON45631, and pMON45632.

Gene Amplification of Aspergillus ochraceus oxidoreductase

An alignment of sequences from the *Aspergillus niger* cprA gene accession number Z26938 (SEQ ID NO: 65) and a partial cDNA clone 804561639F1 from 15 *Aspergillus fumigatus* (PathoSeq Database, Incyte Pharmaceuticals) was visually scanned to select regions of high homology for the design of primers for PCR. A primer set was selected which spanned the coding region of the cprA gene product from amino acids 203 to 693.

Primers were selected from the 5' most region of overlap where the amino 20 acid sequence was identical between both and the nucleic acid sequence differed by 2 positions in the 3rd codon position. For the 5' primer, the nucleic acid encoding the stop codon, last 7 amino acid residues and 2 additional bases corresponding to second and third positions in the codon of the amino acid residue 8 positions from the stop codon encodes ARG in *A. niger* and SER in *A. fumigatus* (CGC vs. AGC). 25 Inosines replaced the third base in codons when there was a discrepancy between the *A. niger* and *A. fumigatus* sequence.

Primer A.oxred-for1: GACGGIGCIGGTACAATGGAA (SEQ ID NO: 11)
Primer A.oxred-rev1: TTAIGACCAIACATCITCCCTGGTAGC (SEQ ID NO: 12)
30 (where I = Inosine)

A partial cDNA clone was amplified from approximately 5 µg of total RNA extracted from *A. ochraceus* mycelia. Before the first strand synthesis, the RNA was heated to 65°C for 10 minutes with 100 ng random hexamers (Promega Madison WI) in an 11 ul reaction mixture. The mixture was chilled on ice, then 35 incubated at 42°C for 75 minutes in a 20 ul reaction containing 1 ul RNase inhibitor (Promega), 0.01 M DTT, 5 mM dNTPs, 50 mM Tris-HCl, pH 8.3), 75 mM

KCl, 3 mM MgCl₂ and 1 ul SuperScriptII (LTI). The reverse transcriptase was inactivated by heating to 95°C for 2 minutes. The first strand cDNA was stored at -20°C. The second strand was synthesized using 5 ul of the first strand as template. The reaction included 500 nM primers, 200 uM each dNTP, and Taq polymerase and buffer as supplied in PCR core kit (Roche Molecular Biochemicals, Indianapolis, IN). Amplification was performed using 32 cycles of a 30 second denaturation step at 94°C, a 30 second annealing step at 60°C and a 60 second elongation step at 72°C. The amplified DNA products were cloned into pGEM-T (Promega, Madison, WI) and sequenced using universal T7 (SEQ ID NO: 41) and SP6 (SEQ ID NO: 57) primers.

Primer SP6 GATTTAGGTGACACTATAG (SEQ ID NO: 57)

Alignment of the sequences with the *A. niger* cprA gene revealed that the *A. ochraceus* clones had an intron in the same position as the intron in the *A. niger* gene. This indicated that the *A. ochraceus* PCR products might have been amplified from a genomic DNA contaminant of the total RNA. A reverse primer based on the *A. ochraceus* sequence was designed to amplify the approximately 600 missing bp including the initial methionine. The *A. ochraceus* cDNA library was then used as a template for PCR. The forward primer was based on the reverse complement of vector pSport1 (Life Technologies) bases 299 to 326. The other primer, A.oxred-rev2 was bases on the *A. ochraceus* sequence encoding residues 326-333.

Primer pSport-for1: CAAGCTCTAAATACGACTCACTATAGGGA (SEQ ID NO: 13)
Primer A.oxred-rev2: CAGGAACCGATCGACCTCGGAA (SEQ ID NO: 14)

The *A. ochraceus* spore library size made from gel-purified fragments >1.5 kb in size was then used as a template for amplifying the final 200 bases of coding region. Two new reverse primers were designed from the A.oxred sequence, and a new forward primer based on pSport1 (bases 295-328) was also used.

Primer A.oxred-rev3: GTCAACCCTCACCGCAGAGCCAATG (SEQ ID NO: 15)
Primer A.oxred-rev4: CCACATTGGAAACCATAGCGTTGTAGTG (SEQ ID NO: 16)
Primer pSport-for2: GCAAGCTCTAAATACGACTCACTATAGGGAAAGC (SEQ ID NO: 17)

Amplification was performed using an Elongase polymerase kit (Life Technologies, Rockville MD) for 35 cycles consisting of a denaturation step at 94°C for 30 seconds, an annealing step at 63°C for 30 seconds, and an elongation step at 68°C for 5 minutes. The PCR products were cloned directly into pCRII TOPO (Invitrogen). Twelve clones were sequenced, and the composite sequence, extended

for 232 bases upstream of the initial methionine, and included 2 in-frame stop codons (Data not shown).

Primers incorporating the complete coding region of A.oxred were designed with a 5' *Sal*I site and a 3' *Xho*I site for ligation into expression vector pFastBac1.

5

Primer A.oxred-for2: gtcgacATGGCGCAA~~T~~CGATACTCTC (SEQ ID NO: 18)
Primer A.oxred-rev5: ctcgagtt~~A~~GGACCAGACATCGTCCTGGTAG (SEQ ID NO: 19)

10 *A. ochraceus* total RNA was used as a template for PCR with these primers and the Elongase kit. Amplification consisted of 35 cycles with a 30 second denaturation step at 94°C, a 30 second annealing step at 64°C, and a 5 minute elongation step at 68°C. An aliquot of the cDNA from reaction ran as a single band of approximately 2.1 kb.

Construction of pMON45630

15 The PCR products were cloned directly into pCRII-TOPO (Invitrogen, Carlsbad, CA). All clones contained the internal intron noted earlier. One clone was designated pMON45630.

Construction of pMON45631 and pMON45632

20 A strategy based on two step PCR from an internal *Bam*HI site approximately 170 bp upstream of the 5' splice site was employed to generate clones lacking the intron.

Primer A.oxred-for3: GGATCCCTCGCGACCTGTGATCAT (SEQ ID NO: 20)
Primer A.oxred-for4: CGAAGATTCTTGTACAAGGATGAATGGAAGACTTTTC (SEQ ID NO: 21)
Primer A.oxred-rev6: CTGAAAAGCTTCCATTCA~~T~~CATCCTTGTACAAGAAATC (SEQ ID NO: 22)

25 Primers A.oxred-for4 and rev6 were complementary and flanked the intron. The first PCR reaction used an A.oxred clone linearized at the internal *Bam*HI site as template. Polymerase and buffers were supplied by the PCR core kit (Roche Molecular Biochemicals, Indianapolis, IN). Primer and dNTP concentrations were 500 nM and 200 uM, respectively. Two reactions were performed, using a combination of A.oxred-for3 with A.oxred-rev6, and A.oxred-for4 with A.oxred-rev5. Following a 2 minute initial denaturation, 28 cycles of PCR amplification were performed. One cycle included a 45 second denaturation at 94°C, a 45 second denaturation step at 62°C and a 45 second elongation step at 72°C. One ul of each reaction served as template for the second PCR amplification with primers A.oxred-for3 and A.oxred-rev5 using Elongase enzyme and buffers. Amplification
30
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consisted of 30 cycles with a 30 second denaturation step at 94°C, a 30 second annealing step at 62°C, and a 5 minute elongation step at 68°C. The PCR products were directly cloned into pCRII-TOPO. DNA sequencing demonstrated that the intron had been removed. This clone was designated pMON45631.

5 Plasmid pMON45632 was constructed in a three-way ligation by ligating the *SalI/BamHI* fragment from pMON45630 with the *BamHI/XhoI* fragment from pMON45631 and vector pFastBac1, which had been cut with *SalI* and *XhoI* and dephosphorylated to enhance the recovery of vectors with the desired inserts.

10 The nucleotide and amino acid sequences of the cloned *Aspergillus ochraceus* 11 oxidoreductase are displayed in Figure 3 as SEQ ID NO: 5 and SEQ ID NO: 6, respectively. Figure 7 sets forth the amino acid homology of *Aspergillus ochraceus* and human oxidoreductase to NADPH cytochrome P450 reductases from *A. niger*, mouse, and *S. cerevisiae*. Figure 8 sets forth the amino acid alignment for 15 *A. ochraceus*, *A. niger*, and *S. cerevisiae* oxidoreductases. Figure 9 is a phylogenetic tree showing the relatedness of *Aspergillus ochraceus* and human oxidoreductase to reductases from *A. niger*, yeast, and mouse. Figure 10 shows the percent homology between *Aspergillus ochraceus* steroid 11 alpha hydroxylase and the oxidoreductases from *A. niger*, yeast, and mouse, calculated using Clustal W and Boxshade.

20 **Example 15: Generation of polyclonal antibodies recognizing *Aspergillus ochraceus* 11 alpha hydroxylase and *Aspergillus ochraceus* NADPH cytochrome p450 reductase**

Generation of anti-11-a-hydroxylase Antibodies

25 Polyclonal antibodies against *Aspergillus ochraceus* 11 alpha hydroxylase and NADPH cytochrome p450 reductase were raised in rabbits against synthetic peptides (prepared by Sigma/Genesis, The Woodlands, TX) corresponding to several regions of the following predicted protein sequences:

30 11aOH peptide 1: AAAYWLATLQPSDLPELN (SEQ ID NO: 23)
11aOH peptide 2: CRQILTTPYIHKRKSLKGTTD (SEQ ID NO: 24)
11aOH peptide 3: HMGFGHGVHACPGRFFASNEI (SEQ ID NO: 25)
oxr peptide 1: CTYWAVAKDPYASAGPAMNG (SEQ ID NO: 26)

The 11aOH peptide 2 (SEQ ID NO: 24) corresponds to the G helix, G/H loop, and H helix region present in an alignment of the amino acid sequence of 11 alpha hydroxylase with the corresponding sequence of CYP3A4 described by Wang and

Lu, (*Drug Metab. Dispos.* 25(6), 762-767, 1997). The 11aOH peptide 3 (SEQ ID NO: 25) corresponded to the peptide fragment from the heme-binding domain.

Immunological grade peptides were monitored for purity using reverse phase high performance liquid chromatography (HPLC). Each peptide was conjugated to keyhole limpet hemacyanin (KLH) and suspended in Complete Freund's Adjuvant. The conjugated peptide was then injected subcutaneously at multiple sites into rabbits. Each conjugated peptide was injected into two rabbits. All subsequent immunizations were given in incomplete Freund's Adjuvant. In general, five subsequent injections were given at two-week intervals following the initial immunization. IgG fractions were affinity-purified using a Sepharose-Protein A column. Fractions from the two rabbits injected with each peptide were combined at a 1:1 ratio. The pooled anti-11 alpha hydroxylase (rabbits GN 1187/1188) was 0.34 mg/ml IgG. The pooled anti-oxred (rabbits GN 2023/2024) was 0.26 mg/ml IgG. The combined IgGs were each diluted 1:10, 1:100 and 1:1,000 for a pilot experiment to determine which dilution was optimal for probing Western blots. The 1:10 dilution gave the best results and was used for probing subsequent Westerns.

Example 16 - Insect Cell Infection and Heterologous Expression

Proteins were expressed in Sf9 insect cells using baculovirus shuttle vectors (Luckow et al., *J. Virol.* 67: 4566-4579, 1993). The baculovirus shuttle vector (bacmid) contains a mini-F replicon for expression in bacterial cells, a kanamycin resistance marker for selection, and *att*Tn7 (the target site for the bacterial Tn7 transposon) within the *lacZ* α sequence. Each of these elements is inserted into the polyhedrin locus of the *Autographa californica* nuclear polyhedrosis virus (AcNPV, the native baculovirus) genome. A donor plasmid (pFastBac1, Life Technologies) was used to deliver the gene to be expressed and was inserted into the bacmid via the bacterial Tn7 transposition elements. pFastBac1 contains the Tn7 left and right ends flanking the polyhedrin promoter, a polylinker cloning sequence, the SV40 polyA transcription termination sequence, and the gentamicin resistance gene for selection. Recombinant viruses were generated following transformation of the pFastBac1 plasmid, which contained a single 11 alpha hydroxylase or oxidoreductase cDNA, into DH10Bac *E. coli* cells (Life Technologies) that contained the bacmid and helper plasmid.

Transfections were performed using CellFectinTM reagent (Life Technologies) following the manufacturer's protocol for *Spodoptera frugiperda*

(Sf9) cells. Cells were seeded in 6-well tissue culture plates at 9×10^5 cells per well in SF-900 serum-free medium (Life Technologies) and allowed to attach for at least one hour. The transfection mixtures were made following the addition of 5 μ l miniprep DNA and 5 μ l Cellfectin to polystyrene tubes that contained 200 μ l SF-900 medium. The mixtures were allowed to incubate for 15-30 minutes at room temperature. Prior to transfection, 800 μ l SF-900 medium was added to each tube. The cells were washed one time with 2 ml SF-900 medium, and the DNA mixtures were added to the cells. The cultures were allowed to incubate for 5 hours at 27°C. Following the 5 hr incubation period, the transfection mixture was removed and the cultures were replenished with 3 ml per well IPL-41 medium (Life Technologies) supplemented with 10% fetal bovine serum. Following a three day incubation period, the cells were harvested, centrifuged, and the supernatant that contained recombinant virus (designated as passage 1 or P1 stock) was removed and stored at 4°C. A larger viral stock was made by infecting 100 ml fresh Sf9 cells at 5×10^5 cells per ml with 0.5 ml of the P1 medium. This larger (P2) stock was then titered using a plaque assay protocol (O'Reilly et al., 1992), and used for production of the 11 alpha hydroxylase or oxidoreductase enzymes, separately or in combination with each other.

Figure 14 sets forth an immunoblot illustrating expression of *Aspergillus ochraceus* P450 11 alpha hydroxylase in baculovirus-infected insect cells harvested at 25 and 48 hours post infection. The nitrocellulose membrane was probed with a 1:1 mixture of antibodies prepared two rabbits immunized with a conjugated synthetic peptide 11aOH peptide 2 (SEQ ID NO 24).

Figure 15 sets forth an immunoblot illustrating expression of *Aspergillus ochraceus* P450 oxidoreductase in baculovirus-infected insect cells harvested at 25 and 48 hours post infection. The nitrocellulose membrane was probed with a 1:1 mixture of antibodies prepared two rabbits immunized with a conjugated synthetic peptide oxr peptide 1 (SEQ ID NO 26).

Example 17: Co-infection baculoviruses expressing of *Aspergillus ochraceus* 11 alpha hydroxylase and human oxidoreductase

Sf9 cells were co-infected with virus particles that contained the steroid 11 alpha hydroxylase cDNA and a separate virus containing a human NADPH P450-oxidoreductase. Both viruses were added at a multiplicity of infection (MOI) ratio of 0.1 : 0.01 (11 aOH to oxr). One day after infection, 0.9 μ g/ml hemin chloride was added to the culture. The cells were harvested by centrifugation three days after

infection (unless specified differently), and the washed cell pellets were frozen until processed for sub-cellular fractions.

Example 18: Co-infection baculoviruses expressing of *Aspergillus ochraceus* 11 alpha hydroxylase and *Aspergillus ochraceus* oxidoreductase

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Sf9 cells are co-infected with virus particles that contain the steroid 11 alpha hydroxylase cDNA and a separate virus containing *A. ochraceus* NADPH P450-oxidoreductase. Both viruses are added at a multiplicity of infection (MOI) ratio of 0.1 : 0.01 (11 aOH to oxr). One day after infection, 0.9 µg/ml hemin chloride is added to the culture. The cells are harvested by centrifugation three days after infection (unless specified differently), and the washed cell pellets are frozen until needed in subsequent experiments that require processing into for sub-cellular fractions.

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Example 19: Preparation of subcellular fractions from baculovirus-infected insect cells

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One half gram of the cell pastes from infected sf9 cells and uninfected control cells were thawed and suspended in 40 ml of 0.25 M sucrose with 10 mM KHPO₄, adjusted to pH 7.4. The suspensions were homogenized using a Fisher Sonic Dismembrator, model 300 probe sonicator (Fisher Scientific, St. Louis, MO). The samples were transferred to a conical centrifuge tube (Corning Costar Corporation, Cambridge, MA) and subjected to centrifugation at 500 x g at 5°C for 15 minutes. The pellets were resuspended in the same volume of fresh buffer and viewed under a microscope to confirm complete lysis. Few or no whole cells were observed. The supernatants were then subjected to centrifugation at 10,000 x g for 30 minutes at 5°C to collect mitochondria, Golgi and other subcellular organelles. The pellets were resuspended in fresh buffer and subjected to centrifugation at 7,800 x g for 30 minutes at 5°C to collect mitochondria.

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The mitochondrial pellets were resuspended in buffer as described above and the centrifugation was repeated. The mitochondrial pellets were resuspended in 2 ml buffered sucrose solution and stored at -80°C in 100 ul aliquots.

The supernatants from the original mitochondrial fractionation were subjected to centrifugation at 200,000 x g for 1 hour at 5°C. The microsomal pellets were resuspended in 2 ml buffered sucrose solution and stored at -80°C in 100 ul aliquots.

Microsomal Incubations

Incubation mixtures consisted of Sf9 microsomes (1.0 mg of protein/mL final concentration), an NADPH-generating system and 250 uM substrate (AD) in 100 mM potassium phosphate buffer, pH 7.4 or 150 mM HEPES buffer, pH 7.4.

5 The NADPH-generating system was composed of the following at the indicated final concentrations: MgCl₂ (7.5 mM), D-glucose-6-phosphate (7.5 mM), NADP (0.80 mM), and glucose-6-phosphate dehydrogenase (1.0 units/mL). Incubations were carried out for the indicated times at 37°C in a water bath. Following incubation, reactions were terminated by the addition of 0.3 ml methanol. The

10 samples were vortexed three times for two seconds and placed on ice, or stored at -70°C for later analysis.

Example 20: HPLC assays to measure conversion of steroid substrates to their hydroxylated counterparts*High Performance Liquid Chromatography (HPLC)*

15 The HPLC method used to separate hydroxylated steroid compounds from steroid substrates, such as 11 α -hydroxyandrostenedione from androstenedione, is a modified version of the testosterone hydroxylase assay, described by Sonderfan et al., *Arch. Biochem. Biophys.* 255: 27-41, 1987). The standards for androstenedione and 11-beta-hydroxyandrostenedione were obtained from Sigma. 11-alpha-hydroxyandrostenedione (89.5% pure, with the major impurity being androstenedione) was provided by Searle Medicinal Chemistry. HPLC grade water and methanol were obtained from Burdick & Jackson.

20 The HPLC system consisted of a Model 1050 series pump, autoinjector and variable wavelength detector (Hewlett-Packard, Naperville, IL), and a Model TC-50 temperature controller and Model CH-30 column heater (both Eppendorf, Madison, WI).

Cell membrane fractions derived from insect cells transfected with recombinant baculoviruses expressing 11-hydroxylase and complementary electron transport proteins were analyzed for 11-hydroxylase activity in a reaction mixture containing 80 mM phosphate buffer, pH 7.4, 8 mM MgCl₂, and 0.9 mM NADP* in a final volume of 200 ul. In order to insure an adequate source of reducing equivalents, an NADPH regenerating system was provided by adding glucose-6-phosphate dehydrogenase (1.5 U/ml) and 8 mM glucose-6-phosphate. Steroid substrate (e.g., androstenedione) was provided at a final concentration of 0.3 mM.

Reaction mixtures were incubated at 37°C for 30 min. The reactions were terminated by the addition of 200 ul methanol and then placed on ice. Samples were pelleted by centrifugation to remove precipitated protein.

On one occasion, the incubation was carried out in a volume of 0.5 ml in siliconized polypropylene 1.5 ml microcentrifuge tubes at 37°C for 120 minutes. The enzyme, prepared from microsomal or mitochondrial fractions, was added and the substrate added at a concentration of 250 µM (e.g., 25 mM methanol stock solution of AD). The cofactor buffer was 100 mM potassium phosphate, pH 7.4, 7.5 mM MgCl₂, 7.5 mM glucose-6-phosphate, 0.80 mM NADP, and 1.0 units/mL glucose-6-phosphate dehydrogenase. HPLC samples were prepared by terminating the 0.5 ml reaction mixture by addition of 0.3 ml methanol, vortexing three times for 2 seconds and storing on ice. The tubes were spun for 5 minutes at ~20,000 x g in a microcentrifuge and the samples transferred to autosampler vials and capped.

Steroid components present in reaction mixtures and media extract were separated and analyzed by reverse phase HPLC using a 250 mm x 4 mm Vydac analytical C-4 column. Chromatograms were developed using a solvent gradient from 40% to 100% methanol over a ten minute time period and holding at 100% methanol for 5 minutes before re-equilibration to initial conditions. The column effluent was monitored for UV absorbance at both 254 and 220 nm.

Androstenedione, testosterone and monohydroxylated androstenedione metabolites were resolved on a Nova-pak C18 column, 4 micron, 3.9 x 150 mm (Waters Chromatography, Milford, MA) equipped with a 0.22 micron Rheodyne precolumn filter at 40°C and 1.0 ml mobile phase/min. A stepped gradient was utilized with water as mobile phase solvent A and methanol as solvent B. The initial concentration of solvent B was 42% for 6 min. The percentage of B was increased linearly to 45% over 4 minutes and then held for 3 minutes. The percentage of B was then increased linearly to 80% over 10 minutes and held there for an additional 2 minutes for a total run time of 25 minutes. The ultraviolet detection wavelength was 247 nm and the injection volume was 200 ul.

Both the "mitochondria" sample and the "microsomal" sample produced peaks matching the HPLC retention time of the 11α-hydroxyandrostenedione standard, while other fractions did not. These "mitochondria" and "microsomal" peaks were 3.2 and 2.3%, respectively, of the total peak area quantitated at 247 nm. The 11α-hydroxyandrostenedione standard was also spiked into a blank microsomal incubation sample at a concentration of 5.0 µg/mL. The concentration

of the "mitochondria" and "microsomal" 11 α -hydroxyandrostenedione peaks were 1.75 and 1.31 μ g/mL, after correcting for the purity of the standard (89.5%). These concentrations represent 2.3 and 1.7% of substrate converted to 11 α -hydroxyandrostenedione, using a substrate concentration of 250 μ M.

5 Figure 16 sets forth an HPLC tracing illustrating the conversion of androstenedione (AD) to its 11 alpha hydroxy counterpart after incubating AD with subcellular fractions prepared from baculovirus-infected insect cells expressing *Aspergillus ochraceus* 11 alpha hydroxylase and human oxidoreductase.

10 **Example 21: Recognition of *Aspergillus ochraceus* 11 alpha hydroxylase and *Aspergillus ochraceus* NADPH cytochrome p450 reductase by immunoblotting using polyclonal antibodies generated against synthetic peptides**

15 Proteins from Sf9 cell lysates (obtained from uninfected and recombinant baculovirus-infected cells) were loaded onto lanes of a 10% gradient acrylamide mini gel (BioRad, Hercules, CA) at equal concentrations (10 μ g per well). The proteins were separated by electrophoresis at 16 mAmps constant current for approximatley 1 hr in a Tris-glycine buffer containing 0.1% SDS (Sigma, St. Louis, MO). The proteins were transferred to nitrocellulose (Schleicher & Schuell, Keene, NH) for 40 min at 70 mAmp constant current. Primary antibodies were diluted 1:10 (from stock concentrations of 0.34 mg/ml IgG for anti-11 alpha hydroxylase (antibodies GN-1187 and GN-1188 prepared from peptide 11aOH peptide 2 CRQILTPYIHKRKSLKGTTD (SEQ ID NO: 24)), and 0.26 mg/ml IgG for anti-oxred (antibodies GN-2023 and GN-12024 prepared from oxr peptide 1 CTYWAVAKDPYASAGPAMNG (SEQ ID NO: 26)) and used to probe the nitrocellulose membrane. The antigens were detected using anti-rabbit horseradish peroxidase (HRP)-linked secondary antibody as recommended by the manufacturer (New England Biolabs, Beverly, MA). Chemilumiescence was detected using luminol and peroxide reagents (New England Biolabs, Beverly, MA) following the protocol provided by the vendor. Light emission was recorded using X-OMAT AR film (Eastman Kodak Company, Rochester, NY). Images were recorded using a Minolta Dimage V digital camera (Minolta Corporation, Ramsey, NJ).

Example 22: Characterization of the *Aspergillus ochraceus* genomic DNA encoding 11 alpha hydroxylase and oxidoreductase

The approaches described above can be used to facilitate the identification of genes encoding steroid hydroxylases and oxidoreductases within the genome of *Aspergillus ochraceus* and closely related microorganisms, including *Aspergillus niger* and *Aspergillus nidulans*. Other preferred organisms are *Rhizopus oryzae*, *Rhizopus stolonifer*, *Streptomyces fradiae*, *Bacillus megaterium*, *Pseudomonas cruciviae*, *Trichothecium roseum*, *Fusarium oxysporum f. sp. cepae*, *Rhizopus arrhizus*, and *Monosporium olivaceum*. Other preferred organisms that are known to have steroid 11 alpha hydroxylase activity are described in the detailed description of the invention, above.

Briefly, genomic DNA is prepared and shotgun cloned into low copy artificial chromosomes propagated in bacteria. A large number of clones are sequenced to ensure statistical representation of the entire genome, and the sequences of overlapping clones merged to produce the final map and sequence of the genome. Analysis of the open reading frames, will reveal regions which are homologous to the steroid hydroxylase and oxidoreductase genes of the present invention, and regions of the translated open reading frames which are homologous to these enzymes using programs designed to facilitate multiple sequence alignments of nucleotide and protein sequence data such as BLAST, CLUSTAL W, and BoxShade. Genes which encode these proteins are obtained from the artificial chromosomes and recloned into expression vectors such as pFastBac1, transformed into appropriate host cells, which are assayed for the presence of enzymes capable of carrying out the conversion of steroid substrates to their oxidized counterparts.

It is intended that the scope of the present invention be determined by reference to the appended claims. It is recognized that a number of variations can be made to this invention as it is currently described but which do not depart from the scope and spirit of the invention without compromising any of its advantages. These include isolation of homologous genes from microorganisms known to carry out 11 alpha hydroxylation of steroid substrates, preferably fungi and bacteria. This invention is also directed to any substitution of analogous components. This includes, but is not restricted to use of these techniques to isolate other P450s which are involved in steroidogenesis, including hydroxylases that act at other

positions in the core molecule, and use of these enzymes to facilitate bioconversion of steroid intermediates in modified host microorganisms.

All references, patents, or applications cited herein are incorporated by reference in their entirety, as if written herein.

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Claims

1. An isolated and purified nucleic acid, encoding *Aspergillus ochraceus* 11 alpha hydroxylase.
2. An isolated DNA, encoding *Aspergillus ochraceus* 11 alpha hydroxylase.
- 5 3. An isolated cDNA, encoding *Aspergillus ochraceus* 11 alpha hydroxylase.
4. An isolated gene, encoding *Aspergillus ochraceus* 11 alpha hydroxylase.
5. An isolated allele of the gene encoding *Aspergillus ochraceus* 11 alpha hydroxylase.
- 10 6. An isolated and purified nucleic acid, wherein said nucleic acid sequence is as set forth in SEQ ID NO: 1.
7. An isolated DNA, wherein said DNA sequence is as set forth in SEQ ID NO: 1.
8. An isolated cDNA, wherein said cDNA sequence is as set forth in SEQ ID NO: 1.
- 15 9. An isolated gene, wherein said gene sequence is as set forth in SEQ ID NO: 1.
10. An isolated allele of a gene, wherein said gene sequence is as set forth in SEQ ID NO: 1.
- 20 11. An isolated protein having the amino acid sequence of *Aspergillus ochraceus* 11 alpha hydroxylase.
12. An isolated variant of the protein having the amino acid sequence of *Aspergillus ochraceus* 11 alpha hydroxylase.
13. A fusion protein comprising the amino acid sequence of *Aspergillus ochraceus* 11 alpha hydroxylase.
- 25 14. An isolated protein wherein the amino acid sequence of said protein is as set forth in SEQ ID NO: 2.

15. An isolated variant of a protein wherein the amino acid sequence of said protein is as set forth in SEQ ID NO: 2.
16. A purified polypeptide, the amino acid sequence of which comprises SEQ ID NO: 2 with at least one conservative amino acid substitution.
- 5 17. A purified polypeptide, the amino acid sequence of which comprises a sequence at least 99% identical to SEQ ID NO: 2.
18. A purified polypeptide, the amino acid sequence of which comprises a sequence at least 95% identical to SEQ ID NO: 2.
19. A purified polypeptide, the amino acid sequence of which comprises a sequence at least 90% identical to SEQ ID NO: 2.
- 10 20. A purified polypeptide, the amino acid sequence of which comprises a sequence at least 75% identical to SEQ ID NO: 2.
21. A purified polypeptide, the amino acid sequence of which comprises a sequence at least 50% identical to SEQ ID NO: 2.
- 15 22. An isolated and purified nucleic acid, encoding *Aspergillus ochraceus* 11 alpha oxidoreductase.
23. An isolated DNA, encoding *Aspergillus ochraceus* oxidoreductase.
24. An isolated cDNA, encoding *Aspergillus ochraceus* oxidoreductase.
- 20 25. An isolated gene, encoding *Aspergillus ochraceus* oxidoreductase.
26. An isolated allele of the gene encoding *Aspergillus ochraceus* oxidoreductase.
27. An isolated and purified nucleic acid, wherein said nucleic acid sequence is as set forth in SEQ ID NO: 5.
- 25 28. An isolated DNA, wherein said DNA sequence is as set forth in SEQ ID NO: 5.
29. An isolated cDNA, wherein said cDNA sequence is as set forth in SEQ ID NO: 5.

30. An isolated gene, wherein said gene sequence is as set forth in SEQ ID NO: 5.
31. An isolated allele of a gene, wherein said gene sequence is as set forth in SEQ ID NO: 5.

5

32. An isolated protein having the amino acid sequence of *Aspergillus ochraceus* oxidoreductase.
33. An isolated variant of the protein having the amino acid sequence of *Aspergillus ochraceus* oxidoreductase.
- 10 34. A fusion protein comprising the amino acid sequence of *Aspergillus ochraceus* oxidoreductase.
35. An isolated protein wherein the amino acid sequence of said protein is as set forth in SEQ ID NO: 6.
- 15 36. An isolated variant of a protein wherein the amino acid sequence of said protein is as set forth in SEQ ID NO: 6.
37. A purified polypeptide, the amino acid sequence of which comprises SEQ ID NO: 6 with at least one conservative amino acid substitution.
38. A purified polypeptide, the amino acid sequence of which comprises a sequence at least 99% identical to SEQ ID NO: 6.
- 20 39. A purified polypeptide, the amino acid sequence of which comprises a sequence at least 95% identical to SEQ ID NO: 6.
40. A purified polypeptide, the amino acid sequence of which comprises a sequence at least 90% identical to SEQ ID NO: 6.
41. A purified polypeptide, the amino acid sequence of which comprises a sequence at least 75% identical to SEQ ID NO: 6.
- 25 42. A purified polypeptide, the amino acid sequence of which comprises a sequence at least 50% identical to SEQ ID NO: 6.

43. An isolated and purified nucleic acid encoding an enzyme that can catalyze the 11 alpha hydroxylation of 3 keto delta 4,5 steroids (3 keto delta 4 steroids); 3 keto delta 4, 5 delta 6, 7 steroids (3 keto delta 4 delta 6 steroids); 3 keto delta 6, 7 steroids (3 keto delta 6 steroids); or 3 keto delta 1, 2 delta 4, 5 steroids (3 keto delta 1 delta 4 steroids).
- 5
44. An isolated and purified nucleic acid of claim 43, wherein said enzyme does not catalyze the 15 beta hydroxylation of 3 keto delta 4,5 steroids; 3 keto delta 4, 5 delta 6, 7 steroids; or 3 keto delta 6, 7 steroids.
- 10
45. The isolated and purified nucleic acid of claim 43 or claim 44, wherein said hydroxylation is selected from the group consisting of:
- (a) canrenone to 11 alpha hydroxy canrenone;
- 15 (b) androstenedione to 11 alpha hydroxy androstenedione;
- (c) aldonia to 11 alpha hydroxy aldonia;
- (d) ADD (1,4 androstenedienedione) to 11 alpha hydroxy ADD;
- (e) mexrenone to 11 alpha hydroxy mexrenone;
- 20 (f) 6 beta mexrenone to 11 alpha hydroxy 6 beta mexrenone;
- (g) 9 alpha mexrenone to 11 alpha hydroxy 9 alpha mexrenone;
- (h) 12 beta mexrenone to 11 alpha hydroxy 12 beta mexrenone;
- 25 (i) delta 12 mexrenone to 11 alpha hydroxy delta 12 mexrenone;
- (j) testosterone to 11 alpha hydroxy testosterone;
- (k) progesterone to 11 alpha hydroxy progesterone;

(l) mexrenone 6,7-bis-lactone to 11 alpha hydroxy
mexrenone 6,7-bis-lactone; and

(m) mexrenone 7,9-bislactone to 11 alpha hydroxy
mexrenone 7,9-bislactone.

5 46. The isolated and purified nucleic acid of claim 45,
wherein said hydroxylation is selected from the
group consisting of:

(a) canrenone to 11 alpha hydroxy canrenone;

10 (b) androstanedione to 11 alpha hydroxy
androstanedione;

(c) aldona to 11 alpha hydroxy aldona; and

(d) ADD (1,4 androstanedienedione) to 11
alpha hydroxy ADD.

15 47. The isolated and purified nucleic acid of
claim 46, wherein said hydroxylation is
from canrenone to 11 alpha hydroxy
canrenone.

20 48. A method of expressing a protein that can catalyze the 11 alpha
hydroxylation of 3 keto delta 4,5 steroids; 3 keto delta 4, 5 delta 6, 7
steroids; 3 keto delta 6, 7 steroids; or 3 keto delta 1, 2 delta 4, 5 steroids
comprising;

(a) transforming or transfecting host cells with an expression cassette
comprising a promoter operably linked to a nucleic acid that
encodes said protein, and

25 (b) expressing said protein in said host cells.

49. A method of producing the protein of claim 48, further comprising
the step of recovering said protein.

50. The method of claim 48 or claim 49 wherein said protein is
Aspergillus ochraceus 11 alpha hydroxylase.

30 51. The method of claim 50, further comprising expressing an

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electron donor protein, wherein said electron donor protein can donate electrons to said protein that can catalyze the 11 alpha hydroxylation of 3 keto delta 4,5 steroids; 3 keto delta 4, 5 delta 6, 7 steroids; 3 keto delta 6, 7 steroids; or 3 keto delta 1, 2 delta 4, 5 steroids.

10

52. The method of claim 51 wherein said electron donor protein is selected from the group consisting of huiman oxidoreductase and *Aspergillus ochraceus* oxidoreductase.

15

53. The method of claim 51 wherein said electron donor protein is *Aspergillus ochraceus* oxidoreductase.

54. The method of claim 51, wherein the nucleic acid encoding said steroid 11 alpha hydroxylase and said electron donor protein are on separate expression cassettes.

20

55. The method of claim 51, wherein the nucleic acid encoding said steroid 11 alpha hydroxylase and said electron donor protein are on the same expression cassettes.

25

56. The method of claim 54 or claim 55 wherein said steroid 11 alpha hydroxylase is *Aspergillus ochraceus* 11 alpha hydroxylase and said electron donor protein is human oxidoreductase.

57. The method of claim 54 or claim 55 wherein said steroid 11 alpha hydroxylase is *Aspergillus ochraceus* 11 alpha hydroxylase and said electron donor protein is *Aspergillus ochraceus* oxidoreductase.

30

58. The method of claim 48 wherein said expression cassette is on an expression vector.

59. The method of claim 58, wherein said expression vector is a baculovirus.

60. The method of claim 59, wherein said baculovirus is a nuclear polyhedrosis virus is selected from the group consisting of *Autographa californica* nuclear polyhedrosis virus and *Bombyx mori* nuclear polyhedrosis virus.
- 5
61. The method of claim 60, wherein said nuclear polyhedrosis virus is *Autographa californica* nuclear polyhedrosis virus.
62. The method of claim 58 wherein said host cells are insect cells.
- 10 63. The method of claim 62 wherein said insect cells are selected from the group consisting of *Spodoptera frugiperda*, *Trichoplusia ni*, *Autographa californica*, and *Manduca sexta* cells.
- 15 64. The method of claim 63 wherein said insect cells are *Spodoptera frugiperda* cells.
65. The method of any of claims 48 through 64, wherein said *Aspergillus ochraceus* 11 alpha hydroxylase is SEQ ID NO: 2.
66. The method of any of claims 48 through 64, wherein said human oxidoreductase is SEQ ID NO: 4.
- 20 67. The method of any of claims 48 through 64, wherein said *Aspergillus ochraceus* oxidoreductase is SEQ ID NO: 6.
68. An isolated and purified polypeptide that can catalyze the 11 alpha hydroxylation of 3 keto delta 4,5 steroids (3 keto delta 4 steroids); 3 keto delta 4, 5 delta 6, 7 steroids (3 keto delta 4 delta 6 steroids); 3 keto delta 6, 7 steroids (3 keto delta 6 steroids); or 3 keto delta 1, 2 delta 4, 5 steroids (3 keto delta 1 delta 4 steroids).
- 25
69. An isolated and purified polypeptide claim 68, wherein said enzyme does not catalyze the 15 beta hydroxylation of 3 keto delta 4,5 steroids; 3 keto delta 4, 5 delta 6, 7 steroids; or 3 keto delta 6, 7 steroids.
- 30

70. The isolated and purified polypeptide of claim 68 or claim 69, wherein said hydroxylation is selected from the group consisting of :
- (a) canrenone to 11 alpha hydroxy canrenone;
- 5 (b) androstenedione to 11 alpha hydroxy androstenedione;
- (c) aldona to 11 alpha hydroxy aldon;a;
- (d) ADD (1,4 androstenedienedione) to 11 alpha hydroxy ADD;
- 10 (e) mexrenone to 11 alpha hydroxy mexrenone;
- (f) 6 beta mexrenone to 11 alpha hydroxy 6 beta mexrenone;
- (g) 9 alpha mexrenone to 11 alpha hydroxy 9 alpha mexrenone;
- 15 (h) 12 beta mexrenone to 11 alpha hydroxy 12 beta mexrenone;
- (i) delta 12 mexrenone to 11 alpha hydroxy delta 12 mexrenone;
- (j) testosterone to 11 alpha hydroxy testosterone;
- 20 (k) progesterone to 11 alpha hydroxy progesterone;
- (l) mexrenone 6,7-bis-lactone to 11 alpha hydroxy mexrenone 6,7-bis-lactone; and
- (m) mexrenone 7,9-bislactone to 11 alpha hydroxy mexrenone 7,9-bislactone.
- 25 71. The isolated and purified polypeptide of claim 70, wherein said hydroxylation is selected from the group consisting of:
- (a) canrenone to 11 alpha hydroxy canrenone;

(b) androstenedione to 11 alpha hydroxy androstenedione;

(c) aldona to 11 alpha hydroxy aldona; and

(d) ADD (1,4 androstenedienedione) to 11 alpha hydroxy ADD.

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72. The isolated and purified enzyme of claim 71, wherein said hydroxylation is from canrenone to 11 alpha hydroxy canrenone.

10 73. An expression cassette comprising a promoter operably linked to an isolated and purified nucleic acid encoding a polypeptide that can catalyze the 11 alpha hydroxylation of 3 keto delta 4,5 steroids (3 keto delta 4 steroids); 3 keto delta 4, 5 delta 6, 7 steroids (3 keto delta 4 delta 6 steroids); 3 keto delta 6, 7 steroids (3 keto delta 6 steroids); or 3 keto delta 1, 2 delta 4, 5 steroids (3 keto delta 1 delta 4 steroids).

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74. An expression cassette of claim 73, wherein said polypeptide does not catalyze the 15 beta hydroxylation of 3 keto delta 4,5 steroids; 3 keto delta 4, 5 delta 6, 7 steroids; or 3 keto delta 6, 7 steroids.

20 75. The expression cassette of claim 73 or claim 74, wherein said hydroxylation is selected from the group consisting of :

(a) canrenone to 11 alpha hydroxy canrenone;

(b) androstenedione to 11 alpha hydroxy androstenedione;

(c) aldona to 11 alpha hydroxy aldona;

25 (d) ADD (1,4 androstenedienedione) to 11 alpha hydroxy ADD;

(e) mexrenone to 11 alpha hydroxy mexrenone;

(f) 6 beta mexrenone to 11 alpha hydroxy 6 beta mexrenone;

- (g) 9 alpha mexrenone to 11 alpha hydroxy 9 alpha mexrenone;
 - (h) 12 beta mexrenone to 11 alpha hydroxy 12 beta mexrenone;
 - 5 (i) delta 12 mexrenone to 11 alpha hydroxy delta 12 mexrenone;
 - (j) testosterone to 11 alpha hydroxy testosterone;
 - (k) progesterone to 11 alpha hydroxy progesterone;
 - (l) mexrenone 6,7-bis-lactone to 11 alpha hydroxy mexrenone 6,7-bis-lactone; and
 - 10 (m) mexrenone 7,9-bislactone to 11 alpha hydroxy mexrenone 7,9-bislactone.
76. The expression cassette of claim 75, wherein said hydroxylation is selected from the group consisting of:
- (a) canrenone to 11 alpha hydroxy canrenone;
 - (b) androstenedione to 11 alpha hydroxy androstenedione;
 - (c) aldona to 11 alpha hydroxy aldona; and
 - 15 (d) ADD (1,4 androstenedienedione) to 11 alpha hydroxy ADD.
77. The expression cassette of claim 76, wherein said hydroxylation is from canrenone to 11 alpha hydroxy canrenone.
- 20 78. An expression cassette comprising a promoter operably linked to an isolated and purified nucleic acid encoding *Aspergillus ochraceus* oxidoreductase.
79. An expression cassette of claim 78 wherein said nucleic acid is SEQ ID NO: 06.

80. An expression cassette comprising a heterologous DNA encoding an enzyme from the metabolic pathway for the synthesis of sitosterol to spirostanolone wherein said enzyme catalyzes at least one conversion selected from the group consisting of:

(a) canrenone to 11 alpha hydroxy canrenone;

(b) androstenedione to 11 alpha hydroxy androstenedione;

(c) aldona to 11 alpha hydroxy aldonate;

(d) ADD (1,4 androstenedienedione) to 11 alpha hydroxy ADD;

(e) mexrenone to 11 alpha hydroxy mexrenone;

(f) 6 beta mexrenone to 11 alpha hydroxy 6 beta mexrenone;

(g) 9 alpha mexrenone to 11 alpha hydroxy 9 alpha mexrenone;

(h) 12 beta mexrenone to 11 alpha hydroxy 12 beta mexrenone;

(i) delta 12 mexrenone to 11 alpha hydroxy delta 12 mexrenone;

(j) testosterone to 11 alpha hydroxy testosterone;

(k) progesterone to 11 alpha hydroxy progesterone;

(l) mexrenone 6,7-bis-lactone to 11 alpha hydroxy mexrenone 6,7-bis-lactone;

(m) mexrenone 7,9-bislactone to 11 alpha hydroxy mexrenone 7,9-bislactone;

and wherein the heterologous DNA is operably linked to control sequences required to express the encoded enzymes in a recombinant host.

81. The expression cassette according to claim 80, characterized in that the heterologous DNA coding sequences are selected from the group consisting of the following genus and species: *Aspergillus ochraceus*, *Aspergillus ochraceus*, *Aspergillus niger*, *Aspergillus nidulans*, *Rhizopus oryzae*, *Rhizopus stolonifer*, *Streptomyces*

fradiae, Bacillus megaterium, Pseudomonas cruciviae,
Trichothecium roseum, Fusarium oxysporum Rhizopus arrhizus,
Absidia coerulea, Absidia glauca, Actinomucor elegans, Aspergillus
flavipes, Aspergillus fumigatus, Beauveria bassiana,
5 *Botryosphaeria obtusa, Calonectria decora, Chaetomium cochlioides,*
Corynespora cassiicola, Cunninghamella blakesleeana,
Cunninghamella echinulata, Cunninghamella elegans, Curvularia
clavata, Curvularia lunata, Cylindrocarpon radicicola, Epicoccum
humicola, Gongronella butleri, Hypomyces chrysospermus,
10 *Monosporium olivaceum, Mortierella isabellina, Mucor mucedo,*
Mucor griseocyanus, Myrothecium verrucaria, Nocardia corallina,
Paecilomyces carneus, Penicillium patulum, Pithomyces
atroolivaceus, Pithomyces cynodontis, Pycnosporium sp.,
Saccharopolyspora erythrae, Sepedonium chrysospermum,
15 *Stachylidium bicolor, Streptomyces hygroscopicus, Streptomyces*
purpurascens, Syncephalastrum racemosum, Thamnostylum
piriforme, Thielavia terricola, and Verticillium theobromae,
Cephalosporium aphidicola, Cochliobolus lunatus, Tieghemella
orchidis, Tieghemella hyalospora, Monosporium olivaceum,
20 *Aspergillus ustus, Fusarium graminearum, Verticillium glaucum,*
and Rhizopus nigricans.

82. The expression cassette according to claim 81, wherein the genus and species are selected from the group consisting of *Aspergillus ochraceus, Aspergillus ochraceus, Aspergillus niger, Aspergillus nidulans, Rhizopus oryzae, Rhizopus stolonifer, Streptomyces fradiae, Bacillus megaterium, Pseudomonas cruciviae, Trichothecium roseum, Fusarium oxysporum Rhizopus arrhizus, and Monosporium olivaceum.*

25 83. The expression cassette according to claim 82, wherein the genus species is *Aspergillus ochraceus.*

30 84. A recombinant host cell and progeny thereof comprising at least one expression cassette according to claim 80.

35 85. The recombinant host cell and progeny thereof according to claim 84, wherein the host is a microorganism.

86. The recombinant host cell and progeny thereof according to claim 85, wherein the host is a bacterium.
87. A process for making one or more enzymes from the metabolic pathway for the synthesis of sitosterol to eplerenone comprising incubating the recombinant host cell of claim 86 in a nutrient medium under conditions where the one or more enzymes encoded by the heterologous DNA are expressed and accumulate.
88. A process for the selective oxidation of a compound to an hydroxylated product, which process comprises the steps of: (a) incubating the compound to be hydroxylated in the presence the recombinant host cells of claim 86 under conditions where the compound is hydroxylated and the hydroxylated product accumulates, and (b) recovering the hydroxylated product.
89. A process for the selective hydroxylation of a compound to an hydroxylated product in vitro, which process comprises the steps of: (a) incubating the compound to be hydroxylated in the presence of the enzymes produced in the process of claim 88 under conditions where the compound is hydroxylated and the hydroxylated product accumulates, and (b) recovering the hydroxylated product.
90. A host cell harboring an expression cassette of any of claims 73 to 83.
91. A host cell of claim 90, wherein said expression cassette is integrated into the chromosome of said host cell.

92. A host cell of claim 90, wherein said expression cassette is integrated into an expression vector.
- 5 93. A method of determining the specific activity of a cloned 11 alpha hydroxylase comprising the steps of;
- (a) transforming host cells with an expression vector comprising a nucleic acid that encodes said 11 alpha hydroxylase,
- (b) expressing said 11 alpha hydroxylase in said host cells;
- (c) preparing subcellular membrane fractions from said cells,
- 10 (d) incubating said subcellular membrane fractions microsomes with a steroid substrate, and
- (e) monitoring conversion of the steroid substrate to its 11 alpha hydroxy steroid counterpart.
94. The method of claim 93 further comprising transforming host cells with an expression vector nucleic acid that encodes an oxidoreductase, and expressing said oxidoreductase in said host cells.
- 15 95. The method of claim 94 wherein said oxidoreductase is human or *Aspergillus ochraceus*.
- 20 96. The method of claim 95 wherein said oxidoreductase is human oxidoreductase.
97. The method of claim 95 wherein said oxidoreductase is *Aspergillus ochraceus* oxidoreductase.
- 25 98. A protein having SEQ ID NO: 2 and variants thereof that are at least 95% identical to SEQ ID NO: 2 and catalyze the 11 alpha hydroxylation of 3 keto delta 4,5 steroids; 3 keto delta 4, 5 delta 6, 7 steroids; 3 keto delta 6, 7 steroids; or 3 keto delta 1, 2 delta 4, 5 steroids, wherein said hydroxylation is selected from the group consisting of :
- 30

- (a) canrenone to 11 alpha hydroxy canrenone;
- (b) androstenedione to 11 alpha hydroxy androstenedione;
- (c) aldonia to 11 alpha hydroxy aldonia;
- (d) ADD (1,4 androstenedienedione) to 11 alpha hydroxy ADD;
- 5 (e) mexrenone to 11 alpha hydroxy mexrenone;
- (f) 6 beta mexrenone to 11 alpha hydroxy 6 beta mexrenone;
- (g) 9 alpha mexrenone to 11 alpha hydroxy 9 alpha mexrenone;
- (h) 12 beta mexrenone to 11 alpha hydroxy 12 beta mexrenone;
- (i) delta 12 mexrenone to 11 alpha hydroxy delta 12 mexrenone;
- 10 (j) testosterone to 11 alpha hydroxy testosterone;
- (k) progesterone to 11 alpha hydroxy progesterone;
- (l) mexrenone 6,7-bis-lactone to 11 alpha hydroxy mexrenone 6,7-bis-lactone; and
- (m) mexrenone 7,9-bislactone to 11 alpha hydroxy mexrenone 7,9-bislactone.

15
20
99. A protein of claim 98, which does not catalyze the 15 beta hydroxylation of 3 keto delta 4,5 steroids; 3 keto delta 4, 5 delta 6, 7 steroids; or 3 keto delta 6, 7 steroids.

100. A purified polypeptide, the amino acid sequence of which is selected from the group consisting of SEQ ID NO: 23, SEQ ID NO: 24, SEQ ID NO: 25.
101. Purified immunogenic polypeptide, the amino acid sequence of which comprises at least ten consecutive residues of SEQ ID NO: 2.
- 25 102. An isolated and purified antibody having a binding specificity for 11 alpha hydroxylase having an amino acid sequence as shown in SEQ ID NO: 2.

103. The antibody of claim 102 which binds to a protein region selected from the group consisting of
- (a) the N-terminal amino acids 1-10 of SEQ ID NO: 2;
- (b) the last 10 C-terminal amino acids of SEQ ID NO: 2;
- 5 (c) amino acids SEQ ID NO: 23;
- (d) amino acids SEQ ID NO: 24; and
- (e) amino acids SEQ ID NO: 25.
104. The antibody of claim 102 or claim 103, wherein said antibody is purified on a peptide column, wherein said peptide is selected from the group consisting of:
- (a) the N-terminal amino acids 1-10 of SEQ ID NO: 2;
- (b) the last 10 C-terminal amino acids of SEQ ID NO: 2;
- (c) amino acids SEQ ID NO: 23;
- (d) amino acids SEQ ID NO: 24; and
- 15 (e) amino acids SEQ ID NO: 25.
105. A purified polypeptide, the amino acid sequence of which is selected from the group consisting of SEQ ID NO: 26.
106. Purified immunogenic polypeptide, the amino acid sequence of which comprises at least ten consecutive residues of SEQ ID NO: 6.
- 20 107. An isolated and purified antibody having a binding specificity for 11 alpha hydroxylase having an amino acid sequence as shown in SEQ ID NO: 6.
108. The antibody of claim 107 which binds to a protein region selected from the group consisting of
- (a) the N-terminal amino acids 1-10 of SEQ ID NO: 6;
- (b) the last 10 C-terminal amino acids of SEQ ID NO: 6; and
- 25 (c) amino acids SEQ ID NO: 26.

109. The antibody of claim 107 or claim 108, wherein said antibody is purified on a peptide column, wherein said peptide is selected from the group consisting of:
- (a) the N-terminal amino acids 1-10 of SEQ ID NO: 6;
- 5 (b) the last 10 C-terminal amino acids of SEQ ID NO: 6; and
- (c) amino acids SEQ ID NO: 26.
110. A composition comprising the antibody of claim 102, 103, 104, 107, 108, or 109 and an effective carrier, vehicle, or auxiliary agent.
111. A composition comprising the antibody of claim 102, 103, 104, 107, 108, or 109 and a solution.
112. The antibody of claim 102, 103, 104, 107, 108, or 109 wherein said antibody is a polyclonal antibody.
113. The antibody of claim 102, 103, 104, 107, 108, or 109 wherein said antibody is a monoclonal antibody.
- 15 114. An antibody of claim 102, 103, 104, 107, 108, or 109 conjugated to an immunoaffinity matrix.
115. A method of using an immunoaffinity matrix of claim 114 to purify a polypeptide from a biological fluid or cell lysate.
116. An antibody of claim 114 wherein said immunoaffinity matrix is SEPHAROSE 4B.
- 20 117. A method of using an immunoaffinity matrix of claim 116 to purify a polypeptide from a biological fluid or cell lysate.
118. A method of using a peptide column to purify an antibody, wherein said peptide is selected from the group consisting of:
- 25 (a) the N-terminal amino acids 1-10 of SEQ ID NO: 2;
- (b) the last 10 C-terminal amino acids of SEQ ID NO: 2;
- (c) amino acids SEQ ID NO: 23;

(d) amino acids SEQ ID NO: 24; and

(e) amino acids SEQ ID NO: 25.

119 A method of using a peptide column to purify an antibody, wherein said peptide is selected from the group consisting of:

5 (a) the N-terminal amino acids 1-10 of SEQ ID NO: 6;

(b) the last 10 C-terminal amino acids of SEQ ID NO: 6; and

(c) amino acids SEQ ID NO: 26.

120. A method of detecting a first polypeptide in a biological fluid, wherein said first polypeptide is selected from the group consisting of 10 alpha hydroxylase and oxidoreductase, comprising the following steps:

(a) contacting said fluid with a second polypeptide, having a binding specificity for said first polypeptide, and

(b) assaying the presence of said second polypeptide to determine the level of said first polypeptide.

15

121. The method of claim 120, wherein said second polypeptide is an antibody.

122. The method of claim 120 or claim 121 wherein said second polypeptide is radiolabeled.

20

123. A process for producing an isolated nucleic acid comprising hybridizing SEQ ID NO: 1 to genomic DNA in 6XSSC and 65°C and isolating the nucleic acid detected with SEQ ID NO: 1.

25

124. The isolated DNA nucleic acid prepared according to the process of claim 123.

125. An isolated nucleic acid that specifically hybridizes under highly stringent conditions to the complement of the sequence set forth in SEQ ID NO: 1.

123. A process for producing an isolated nucleic acid comprising hybridizing SEQ ID NO: 5 to genomic DNA in 6XSSC and 65°C and isolating the nucleic acid detected with SEQ ID NO: 5.
124. The isolated DNA nucleic acid prepared according to the process of
5 claim 123.
125. An isolated nucleic acid that specifically hybridizes under highly stringent conditions to the complement of the sequence set forth in SEQ ID NO: 5.
126. A DNA construct which alters the expression of a steroid 11 alpha hydroxylase gene not normally expressed in a cell when said DNA
10 construct is inserted into chromosomal DNA of the cell, said DNA construct comprising:
 - a) a targeting sequence;
 - b) a regulatory sequence; and
 - c) the structural gene for a steroid 11 alpha hydroxylase.
127. A host cell harboring the DNA construct of claim 126.
15
128. Use of a host cell harboring a cloned 11 alpha hydroxylase for the manufacture of a medicament for therapeutic application to treat heart disease, inflammation, arthritis, or cancer.
129. A composition comprising from about 0.5-500 g/L molasses, 0.5-50 g/L
20 cornsteep liquid, 0.5-50 g/L KH₂PO₄, 2.5-250 g/L NaCl, 2.5-250 g/L glucose, and 0.04-4 g/L progesterone, pH 3.5-7.
130. A composition comprising from about 10-250 g/L molasses, 1-25 g/L cornsteep liquid, 1-25 g/L KH₂PO₄, 5-125 g/L NaCl, 5-125 g/L glucose, and 0.08-2 g/L progesterone, pH 4.5-6.5.
25
131. A composition comprising from about 25-100 g/L molasses, 2.5-10 g/L cornsteep liquid, 2.5-10 g/L KH₂PO₄, 12.5-50 g/L NaCl, 12.5-50 g/L glucose, and 0.2-0.8 g/L progesterone, pH 5.5-6.0.
132. A composition comprising from 50 g/L molasses, 5 g/L cornsteep liquid, 5
g/L KH₂PO₄, 25 g/L NaCl, 25 g/L glucose, 20 g/L agar, and 0.4 g/L
30 progesterone, pH 5.8.

133. A composition of any of claims 129-132 further comprising from about 4-100 g/L agar.
134. A composition of any of claims 129-132 further comprising from about 10-40 g/L agar.
- 5 135. A composition of any of claims 129-132 further comprising about 20 g/L agar.
136. Use of the composition of any of claims 129-135 to produce spores from the microorganism selected from the group consisting of *Aspergillus ochraceus*, *Aspergillus niger*, *Aspergillus nidulans*, *Rhizopus oryzae*, *Rhizopus stolonifer*, and *Trichothecium roseum*, *Fusarium oxysporum* *Rhizopus arrhizus*, *Monosporium olivaceum*, *Penicillium chrysogenum*, and *Absidia coerulea*.
- 10 136. Use of the composition of any of claims 129-135 to produce spores from *Aspergillus ochraceus*.

Figures

Figure 1 - Nucleotide and protein sequence of Aspergillus ochraceus 11 alpha hydroxylase

5	tggaaagttt tacacttatt atgccggagc cggaaaggatc tgagtgcagg ggttggggaa caacactata agacctaca ccacttgat ttggtgaatt tacacggca ttatcaaacc agccacaagc tgacagctca ttatc atg ccc ttc ttc act ggg ctt ctg gcg Met Pro Phe Phe Thr Gly Leu Leu Ala	60 120 172
10	1 att tac cat agt ctc ata ctc gac aac cca gtc caa acc ctg agc acc Ile Tyr His Ser Leu Ile Leu Asp Asn Pro Val Gln Thr Leu Ser Thr 10 15 20 25	220
15	att gtc gta ttg gcg gca gcg tac tgg ctc gca acg ctc cag ccg agc Ile Val Val Leu Ala Ala Ala Tyr Trp Leu Ala Thr Leu Gln Pro Ser 30 35 40	268
20	gac ctt cct gag ctg aat ccc gcc aaa cca ttc gag ttc acc aat cgt Asp Leu Pro Glu Leu Asn Pro Ala Lys Pro Phe Glu Phe Thr Asn Arg 45 50 55	316
25	cgt cgt gtt cat gag ttt gtt gaa aat agt aag aag agc ttg ctt gct cgg Arg Arg Val His Glu Phe Val Glu Asn Ser Lys Ser Leu Ala Arg 60 65 70	364
30	ggg agg gaa ttg cac ggg cac gag ccg tac aga ctc atg tct gaa tgg Gly Arg Glu Leu His Gly His Glu Pro Tyr Arg Leu Met Ser Glu Trp 75 80 85	412
35	gga tcc ttg att gtc ctg ccc cca gag tgc gac gag ctg ctc aac Gly Ser Leu Ile Val Leu Pro Pro Glu Cys Ala Asp Glu Leu Arg Asn 90 95 100 105	460
40	gac cca aga atg gac ttt gag acg ccc acc acc gac gac tcc cac gga Asp Pro Arg Met Asp Phe Glu Thr Pro Thr Asp Asp Ser His Gly 110 115 120	508
45	tat atc cct ggc ttc gac gct ctc aac gca gac ccg aac ctg act aaa Tyr Ile Pro Gly Phe Asp Ala Leu Asn Ala Asp Pro Asn Leu Thr Lys 125 130 135	556
50	gtg gtc acc aag tac ctc aca aaa gca ttg aac aag ctt act gct ccg Val Val Thr Lys Tyr Leu Thr Lys Ala Leu Asn Lys Leu Thr Ala Pro 140 145 150	604
55	atc tcg cat gaa gcg tcc atc gac atg aaa gcg gtg ctg ggt gac gat Ile Ser His Glu Ala Ser Ile Ala Met Lys Ala Val Leu Gly Asp Asp 155 160 165	652
60	cca gat tgg cgt gag atc tac cca gcc aga gac ttg ctc cag ctc gtc Pro Asp Trp Arg Glu Ile Tyr Pro Ala Arg Asp Leu Leu Gln Leu Val 170 175 180 185	700
65	gcc cgg atg tcg aca aga gtg ttc ctt ggc gag gaa atg tgc aat aac Ala Arg Met Ser Thr Arg Val Phe Leu Gly Glu Met Cys Asn Asn 190 195 200	748
70	cag gat tgg atc caa acc tca tca caa tac gcg gcc ctt gcc ttc ggt Gln Asp Trp Ile Gln Thr Ser Ser Gln Tyr Ala Ala Leu Ala Phe Gly 205 210 215	796
75	gtc ggt gac aag ctt aga ata tac ccg aga atg atc aga ccg ata gta Val Gly Asp Lys Leu Arg Ile Tyr Pro Arg Met Ile Arg Pro Ile Val 220 225 230	844
80	cat tgg ttc atg cca tcc tgt tgg gag ctg cgc cga tcg ctg cga cgc His Trp Phe Met Pro Ser Cys Trp Glu Leu Arg Arg Ser Leu Arg Arg 235 240 245	892
85	tgc cga cag att ctc acg ccg tac att cac aaa cgc aag tcc ctg aag Cys Arg Gln Ile Leu Thr Pro Tyr Ile His Lys Arg Lys Ser Leu Lys 250 255 260 265	940
90	ggg acc acg gac gag cag ggc aag ccc ctt atg ttt gat gat tcc atc Gly Thr Thr Asp Glu Gln Gly Lys Pro Leu Met Phe Asp Asp Ser Ile 270 275 280	988

	gag tgg ttc gag cga gag ctg ggt ccc aac cac gac gcg gtc ctg aag Glu Trp Phe Glu Arg Glu Leu Gly Pro Asn His Asp Ala Val Leu Lys 285 290 295	1036
5	cag gtc acg ctc tcc ata gtt gct atc cac acc acg agt gac cta ctc Gln Val Thr Leu Ser Ile Val Ala Ile His Thr Thr Ser Asp Leu Leu 300 305 310	1084
10	ttg cag gcc atg agc gat ctc gcg cag AAC ccg aaa gtg cta caa gca Leu Gln Ala Met Ser Asp Leu Ala Gln Asn Pro Lys Val Leu Gln Ala 315 320 325	1132
15	gtg cgc gag gag gtg gtc cga gtg ctg agc acc gag ggg ctc agc aag Val Arg Glu Glu Val Val Arg Val Leu Ser Thr Glu Gly Leu Ser Lys 330 335 340 345	1180
20	gtc tcg ctt cac agt ctc aag ctc atg gac agc gcg ttg aag gaa agc Val Ser Leu His Ser Leu Lys Leu Met Asp Ser Ala Leu Lys Glu Ser 350 355 360	1228
25	cag cgt ctc agg cct acg ctt ctc ggc tcc ttt cgt cgg cag gca acg Gln Arg Leu Arg Pro Thr Leu Leu Gly Ser Phe Arg Arg Gln Ala Thr 365 370 375	1276
	aat gac atc aag ctg aag agc ggg ttt gtc ata aag aaa ggg act aga Asn Asp Ile Lys Leu Lys Ser Gly Phe Val Ile Lys Lys Gly Thr Arg 380 385 390	1324
30	gtc gtg atc gac acc cat atg tgg aat ccc gag tat tac act gac Val Val Ile Asp Ser Thr His Met Trp Asn Pro Glu Tyr Tyr Thr Asp 395 400 405	1372
35	cct ctc cag tac gac ggg tac cgc tac ttc aac aag cgg cag aca ccc Pro Leu Gln Tyr Asp Gly Tyr Arg Tyr Phe Asn Lys Arg Gln Thr Pro 410 415 420 425	1420
40	ggc gag gac aag aac gcg ttg ctc gtc agc aca agc gcc aac cac atg Gly Glu Asp Lys Asn Ala Leu Leu Val Ser Thr Ser Ala Asn His Met 430 435 440	1468
45	gga ttc ggt cac ggc gtt cac gcc tgc cct ggc aga ttc ttc gcc tcc Gly Phe Gly His Gly Val His Ala Cys Pro Gly Arg Phe Phe Ala Ser 445 450 455	1516
	aac gag atc aag att gcc ttg tgt cat atc atc tta aat tat gag tgg Asn Glu Ile Lys Ile Ala Leu Cys His Ile Ile Leu Asn Tyr Glu Trp 460 465 470	1564
50	cgt ctt cca gac ggc ttc aag ccc cag cct ctc aac atc ggg atg act Arg Leu Pro Asp Gly Phe Lys Pro Gln Pro Leu Asn Ile Gly Met Thr 475 480 485	1612
55	tat ctg gcg gat ccc aat acc agg atg ctg atc agg cca cgc aag gcg Tyr Leu Ala Asp Pro Asn Thr Arg Met Leu Ile Arg Pro Arg Lys Ala 490 495 500 505	1660
	gag atc gat atg gcg agt tta act gtg tag gtcgaacacg aagtccgtat Glu Ile Asp Met Ala Ser Leu Thr Val *	1710
60	aaaaaaa gaagtgttat tggcagtgg gtgaagcaag tcgcagaaat gtgtacaata ttataagaat	1770 1776

***Figure 2 - Nucleotide and protein sequence of human
oxidoreductase***

5	atg gga gac tcc cac gtg gac acc agc tcc acc gtg tcc gag gcg gtg Met Gly Asp Ser His Val Asp Thr Ser Ser Thr Val Ser Glu Ala Val 1 5 10 15	48
10	gcc gaa gaa gta tct ctt ttc agc atg acg gac atg att ctg ttt tcg Ala Glu Glu Val Ser Leu Phe Ser Met Thr Asp Met Ile Leu Phe Ser 20 25 30	96
15	ctc atc gtg ggt ctc cta acc tac tgg ttc ctc ttc aga aag aaa aaa Leu Ile Val Gly Leu Leu Thr Tyr Trp Phe Leu Phe Arg Lys Lys Lys 35 40 45	144
20	gaa gaa gtc ccc gag ttc acc aaa att cag aca ttg acc tcc tct gtc Glu Glu Val Pro Glu Phe Thr Lys Ile Gln Thr Leu Thr Ser Ser Val 50 55 60	192
25	aga gag agc agc ttt gtg gaa aag atg aag aaa acg ggg agg aac atc Arg Glu Ser Ser Phe Val Glu Lys Met Lys Lys Thr Gly Arg Asn Ile 65 70 75 80	240
30	atc gtg ttc tac ggc tcc cag acg ggg act gca gag gag ttt gcc aac Ile Val Phe Tyr Gly Ser Gln Thr Gly Thr Ala Glu Glu Phe Ala Asn 85 90 95	288
35	cgc ctg tcc aag gac gcc cac cgc tac ggg atg cga ggc atg tca gcg Arg Leu Ser Lys Asp Ala His Arg Tyr Gly Met Arg Gly Met Ser Ala 100 105 110	336
40	gac cct gag gag tat gac ctg gcc gac ctg agc agc ctg cca gag atc Asp Pro Glu Glu Tyr Asp Leu Ala Asp Leu Ser Ser Leu Pro Glu Ile 115 120 125	384
45	gac aac gcc ctg gtg gtt ttc tgc atg gcc acc tac ggt gag gga gac Asp Asn Ala Leu Val Val Phe Cys Met Ala Thr Tyr Gly Glu Gly Asp 130 135 140	432
50	ccc acc gac aat gcc cag gtc ttc tac gac tgg ctg cag gag aca gac Pro Thr Asp Asn Ala Gln Asp Phe Tyr Asp Trp Leu Gln Glu Thr Asp 145 150 155 160	480
55	gtg gat ctc tct ggg gtc aag ttc gcg gtg ttt ggt ctt ggg aac aag Val Asp Leu Ser Gly Val Lys Phe Ala Val Phe Gly Leu Gly Asn Lys 165 170 175	528
60	acc tac gag cac ttc aat gcc atg ggc aag tac gtg gac aag cgg ctg Thr Tyr Glu His Phe Asn Ala Met Gly Lys Tyr Val Asp Lys Arg Leu 180 185 190	576
65	gag cag ctc ggc gcc cag cgc atc ttt gag ctg ggg ttg ggc gac gac Glu Gln Leu Gly Ala Gln Arg Ile Phe Glu Leu Gly Leu Gly Asp Asp 195 200 205	624
70	gat ggg aac ttg gag gag gac ttc atc acc tgg cga gag cag ttc tgg Asp Gly Asn Leu Glu Glu Asp Phe Ile Thr Trp Arg Glu Gln Phe Trp 210 215 220	672
75	ccg gcc gtg tgt gaa cac ttt ggg gtg gaa gcc act ggc gag gag tcc Pro Ala Val Cys Glu His Phe Gly Val Glu Ala Thr Gly Glu Glu Ser 225 230 235 240	720
80	agc att cgc cag tac gag ctt gtg gtc cac acc gac ata gat gcg gcc Ser Ile Arg Gln Tyr Glu Leu Val Val His Thr Asp Ile Asp Ala Ala 245 250 255	768
85	aag gtg tac atg ggg gag atg ggc cgg ctg aag agc tac gag aac cag Lys Val Tyr Met Gly Glu Met Gly Arg Leu Lys Ser Tyr Glu Asn Gln 260 265 270	816
90	aag ccc ccc ttt gat gcc aag aat ccg ttc ctg gct gca gtc acc acc Lys Pro Pro Phe Asp Ala Lys Asn Pro Phe Leu Ala Ala Val Thr Thr 275 280 285	864
95	aac cgg aag ctg aac cag gga acc gag cgc cac ctc atg cac ctg gaa Asn Arg Lys Leu Asn Gln Gly Thr Glu Arg His Leu Met His Leu Glu 290 295 300	912
100	tgt gac atc tcg gac tcc aaa atc agg tat gaa tct ggg gac cac gtg Leu Asp Ile Ser Asp Ser Lys Ile Arg Tyr Glu Ser Gly Asp His Val	960

	305	310	315	320	
5	gct gtg tac cca gcc aac gac tct gct ctc gtc aac cag ctg ggc aaa Ala Val Tyr Pro Ala Asn Asp Ser Ala Leu Val Asn Gln Leu Gly Lys 325 330 335				1008
10	atc ctg ggt gcc gac ctg gac gtc gtc atg tcc ctg aac aac ctg gat Ile Leu Gly Ala Asp Leu Asp Val Val Met Ser Leu Asn Asn Leu Asp 340 345 350				1056
15	gag gag tcc aac aag aag cac cca ttc ccg tgc cct acg tcc tac cgc Glu Glu Ser Asn Lys Lys His Pro Phe Pro Cys Pro Thr Ser Tyr Arg 355 360 365				1104
20	acg gcc ctc acc tac tac ctg gac atc acc aac ccg ccg cgt acc aac Thr Ala Leu Thr Tyr Tyr Leu Asp Ile Thr Asn Pro Pro Arg Thr Asn 370 375 380				1152
25	gtg ctg tac gag ctg gcg cag tac gcc tcg gag ccc tcg gag gag gag Val Leu Tyr Glu Leu Ala Gln Tyr Ala Ser Glu Pro Ser Glu Gln Glu 385 390 395 400				1200
30	ctg ctg cgc aag atg gcc tcc tcc ggc gag ggc aag gag ctg tac Leu Leu Arg Lys Met Ala Ser Ser Gly Glu Gly Lys Glu Leu Tyr 405 410 415				1248
35	ctg agc tgg gtg gtg gag gcc ccg agg cac atc ctg gcc atc ctg cag Leu Ser Trp Val Val Glu Ala Arg Arg His Ile Leu Ala Ile Leu Gln 420 425 430				1296
40	gac tgc ccg tcc ctg cgg ccc ccc atc gac cac ctg tgt gag ctg ctg Asp Cys Pro Ser Leu Arg Pro Pro Ile Asp His Leu Cys Glu Leu Leu 435 440 445				1344
45	ccg cgc ctg cag gcc cgc tac tac tcc atc gcc tca tcc tcc aag gtc Pro Arg Leu Gln Ala Arg Tyr Tyr Ser Ile Ala Ser Ser Ser Lys Val 450 455 460				1392
50	cac ccc aac tct gtg cac atc tgt ggc gtt gtg gag tac gag acc His Pro Asn Ser Val His Ile Cys Ala Val Val Val Glu Tyr Glu Thr 465 470 475 480				1440
55	aag gcc ggc cgc atc aac aag ggc gtg gcc acc eac tgg ctg ccg gcc Lys Ala Gly Arg Ile Asn Lys Gly Val Ala Thr Asn Trp Leu Arg Ala 485 490 495				1488
60	aag gag cct gcc ggg gag aac ggc ggc cgt gcg ctg gtg ccc atg ttc Lys Glu Pro Ala Gly Glu Asn Gly Gly Arg Ala Leu Val Pro Met Phe 500 505 510				1536
65	gtg cgc aag tcc cag ttc cgc ctg ccc ttc aag gcc acc acg cct gtc Val Arg Lys Ser Gln Phe Arg Leu Pro Phe Lys Ala Thr Thr Pro Val 515 520 525				1584
70	atc atg gtg ggc ccc ggc acc ggg gtg gca ccc ttc ata ggc ttc atc Ile Met Val Gly Pro Gly Thr Gly Val Ala Pro Phe Ile Gly Phe Ile 530 535 540				1632
75	cag gag cgg gcc tgg ctg cga cag cag ggc aag gag gtg ggg gag acg Gln Glu Arg Ala Trp Leu Arg Gln Gln Gly Lys Glu Val Gly Glu Thr 545 550 555 560				1680
80	ctg ctg tac tac ggc tgc cgc tgc gat gag gac tac ctg tac cgg Leu Leu Tyr Tyr Gly Cys Arg Arg Ser Asp Glu Asp Tyr Leu Tyr Arg 565 570 575				1728
85	gag gag ctg ggc cag ttc cac agg gac ggt ggc ctc acc cag ctc aac Glu Glu Leu Ala Gln Phe His Arg Asp Gly Ala Leu Thr Gln Leu Asn 580 585 590				1776
	gtg gcc ttc tcc cgg gag cag tcc cac aag gtc tac gtc cag cac ctg Val Ala Phe Ser Arg Glu Gln Ser His Lys Val Tyr Val Gln His Leu 595 600 605				1824
	cta aag caa gac cga gag cac ctg tgg aag ttg atc gaa ggc ggt gcc Leu Lys Gln Asp Arg Glu His Leu Trp Lys Leu Ile Glu Gly Gly Ala 610 615 620				1872
	cac atc tac gtc tgt ggg gat gca cgg aac atg gcc agg gat gtg cag His Ile Tyr Val Cys Gly Asp Ala Arg Asn Met Ala Arg Asp Val Gln 625 630 635 640				1920
	aac acc ttc tac gac atc gtg gct gag ctc ggg gcc atg gag cac ggc Asn Thr Phe Tyr Asp Ile Val Ala Glu Leu Gly Ala Met Glu His Ala 645 650 655				1968

cag gcg gtg gac tac atc aag aaa ctg atg acc aag ggc cgc tac tcc
Gln Ala Val Asp Tyr Ile Lys Lys Leu Met Thr Lys Gly Arg Tyr Ser
660 665 670

5 ctg gac gtg tgg agc
Leu Asp Val Trp Ser
675

Figure 3 - Nucleotide and protein sequence of Aspergillus ochraceus oxidoreductase

5	cttatttcgt ttaggaagag caccggcttc ggtgtcccttc cttaccctct tattttccct cttctgactc ccsttttgtt attgatgcgc catctcggtg aacatggg atatctttcc ctctccccctt cccgccccga ccctccctat ctctccctcc cgccaggcat ttatcgcc atcgaaatcg caattcccttc ctatcgactc ttatcgctg agcgccctca tc atg gcg Met Ala	60 120 180 238
10	1 caa ctc gat act ctc gat ttg gtc gtc gtg gcg ctc ttg gtg ggt Gln Leu Asp Thr Leu Asp Leu Val Val Leu Val Ala Leu Leu Val Gly 5 10 15	286
15	agc gtg gcc tac ttc acc aag ggc acc tac tgg gcc gtc gcc aaa gac Ser Val Ala Tyr Phe Thr Lys Gly Thr Tyr Trp Ala Val Ala Lys Asp 20 25 30	334
20	cct tat gcc tcg gct ggt ccg gcg atg aat gga ggc gcc aag gcc ggc Pro Tyr Ala Ser Ala Gly Pro Ala Met Asn Gly Gly Ala Lys Ala Gly 35 40 45 50	382
25	aag act cgc gac att gtt cag aaa atg gac gaa act ggc aaa aac tgt Lys Thr Arg Asp Ile Val Gln Lys Met Asp Glu Thr Gly Lys Asn Cys 55 60 65	430
30	gtg att ttc tac ggc tcg caa acc ggt acc gct gag gac tac gcg tcc Val Ile Phe Tyr Gly Ser Gln Thr Gly Thr Ala Glu Asp Tyr Ala Ser 70 75 80	478
35	aga ctg gcc aag gaa ggc tcc cag cga ttc ggt ctc aag acc atg gtg Arg Leu Ala Lys Glu Gly Ser Gln Arg Phe Gly Leu Lys Thr Met Val 85 90 95	526
40	gcc gat ctg gag gac tac gac tac gaa aac ctg gaa aag ttc ccc gag Ala Asp Leu Glu Asp Tyr Asp Tyr Glu Asn Leu Glu Lys Phe Pro Glu 100 105 110	574
45	gac aag gtt gtt ttc ttc gtt ctg gcc act tat ggc gag ggt gaa ccc Asp Lys Val Val Phe Phe Val Leu Ala Thr Tyr Gly Glu Gly Glu Pro 115 120 125 130	622
50	acg gat aat gcg gtt gaa ttc tac cag ttc gtc acg ggc gaa gat gct Thr Asp Asn Ala Val Glu Phe Tyr Gln Phe Val Thr Gly Glu Asp Ala 135 140 145	670
55	gct ttc gag agc ggc gct acc gcc gac gat aag cct ctg tct tct ctc Ala Phe Glu Ser Gly Ala Thr Ala Asp Asp Lys Pro Leu Ser Ser Leu 150 155 160	718
60	aag tat gtc acg ttt ggt ctg ggt aac aac acc tat gag cac tac aac Lys Tyr Val Thr Phe Gly Leu Gly Asn Asn Thr Tyr Glu His Tyr Asn 165 170 175	766
65	gct atg gtt cgc aat gtg gac gcc gct ctc aca aag ttc ggc gcc caa Ala Met Val Arg Asn Val Asp Ala Ala Leu Thr Lys Phe Gly Ala Gln 180 185 190	814
70	cgc att ggc tct gct ggt gag gac gac ggc gct ggt aca atg gaa Arg Ile Gly Ser Ala Gly Glu Asp Asp Gly Ala Gly Thr Met Glu 195 200 205 210	862
75	gag gat ttc ctg gcc tgg aag gaa ccc atg tgg gct gcc ctt tct gag Glu Asp Phe Leu Ala Trp Lys Glu Pro Met Trp Ala Ala Leu Ser Glu 215 220 225	910
80	gct atg aac ctg caa gag cgc gat ggc gtc tac gag ccg gtc ttc aat Ala Met Asn Leu Gln Glu Arg Asp Ala Val Tyr Glu Pro Val Phe Asn 230 235 240	958
85	gtc acc gag gac gag tcc ctg agc ccc gaa gat gag aac gtt tac ctc Val Thr Glu Asp Glu Ser Leu Ser Pro Glu Asp Glu Asn Val Tyr Leu 245 250 255	1006
90	ggt gag ccc act caa ggt cat ctc caa ggc gag ccc aag ggc ccg tac Gly Glu Pro Thr Gln Gly His Leu Gln Gly Glu Pro Lys Gly Pro Tyr 260 265 270	1054
95	tct gcg cac aac ccg ttc atc gct ccc atc tcc gaa tct cgt gaa ctg Ser Ala His Asn Pro Phe Ile Ala Pro Ile Ser Glu Ser Arg Glu Leu 275 280 285 290	1102

	ttc aac gtc aag gac cgc aac tgt ctg cac atg gaa atc agc atc gcc Phe Asn Val Lys Asp Arg Asn Cys Leu His Met Glu Ile Ser Ile Ala 295 300 305	1150
5	ggt agc aac ctc act tac cag act ggt gac cac atc gct gtt tgg ccc Gly Ser Asn Leu Thr Tyr Gln Thr Gly Asp His Ile Ala Val Trp Pro 310 315 320	1198
10	acc aac gcc ggt tcc gag gtc gat cgg ttc ctg cag gct ttt ggt ctc Thr Asn Ala Gly Ser Glu Val Asp Arg Phe Leu Gln Ala Phe Gly Leu 325 330 335	1246
15	gaa gga aag cgc cac tcc gtc atc aac att aag ggt atc gat gtg acc Glu Gly Lys Arg His Ser Val Ile Asn Ile Lys Gly Ile Asp Val Thr 340 345 350	1294
20	gct aag gtt ccg att ccc act cct acg acc tat gac gcc gca gtt cgc Ala Lys Val Pro Ile Pro Thr Pro Thr Tyr Asp Ala Ala Val Arg 355 360 365 370	1342
25	tac tac ctg gaa gtc tgt gcc ccc gtt tcc cgt cag ttt gtc tcg act Tyr Tyr Leu Glu Val Cys Ala Pro Val Ser Arg Gln Phe Val Ser Thr 375 380 385	1390
30	ctc gct gcc ttt gcc cct gat gaa gcg acc aag gcg gag atc gtt cgt Leu Ala Ala Phe Ala Pro Asp Glu Ala Thr Lys Ala Glu Ile Val Arg 390 395 400	1438
35	ttg ggt ggc gac aag gac tat ttc cat gag aag att acc aac cga tgc Leu Gly Gly Asp Lys Asp Tyr Phe His Glu Lys Ile Thr Asn Arg Cys 405 410 415	1486
40	ttc aac atc gct cag gct ctc cag agc atc acg tcc aag cct ttc acc Phe Asn Ile Ala Gln Ala Leu Gln Ser Ile Thr Ser Lys Pro Phe Thr 420 425 430	1534
45	gcc gtc ccg ttc tcc ctg ctt atc gaa ggt atc acc aag ctt cag ccc Ala Val Pro Phe Ser Leu Leu Ile Glu Gly Ile Thr Lys Leu Gln Pro 435 440 445 450	1582
50	cgt tac tac tcg atc tcc tcg tct tcc ctg gtt cag aag gac aag att Arg Tyr Tyr Ser Ile Ser Ser Ser Leu Val Gln Lys Asp Lys Ile 455 460 465	1630
55	agc att acc gcc gtt gtg gag tcg gtt cgc ttg cct ggt gag gaa cac Ser Ile Thr Ala Val Val Glu Ser Val Arg Leu Pro Gly Glu Glu His 470 475 480	1678
60	att gtc aag ggt gtg acc acg aac tat ctt ctc gcg ctc aag gaa aag Ile Val Lys Gly Val Thr Thr Asn Tyr Leu Leu Ala Leu Lys Glu Lys 485 490 495	1726
65	caa aac ggc gag cct tcc cct gac ccg cac ggc ttg act tac tct atc Gln Asn Gly Glu Pro Ser Pro Asp Pro His Gly Leu Thr Tyr Ser Ile 500 505 510	1774
70	act gga ccc cgt aac aag tac gat ggc atc cat gtc ccc gtt cac gtc Thr Gly Pro Arg Asn Lys Tyr Asp Gly Ile His Val Pro Val His Val 515 520 525 530	1822
75	cgc cac tcg aac ttc aaa ttg ccc tcg gat ccc tcg cga cct gtg atc Arg His Ser Asn Phe Lys Leu Pro Ser Asp Pro Ser Arg Pro Val Ile 535 540 545	1870
80	atg gtt gga ccc ggt act ggt gtt gct cct ttc cgt ggg ttt atc cag Met Val Gly Pro Gly Thr Gly Val Ala Pro Phe Arg Gly Phe Ile Gln 550 555 560	1918
85	gag cgt gct gcc ttg gcc gcg aag ggc gag aag gtc gga act acc ttg Glu Arg Ala Ala Leu Ala Ala Lys Gly Glu Lys Val Gly Thr Thr Leu 565 570 575	1966
90	ctt ttc ttc ggc tgc cgt aag tcc gac gaa gat ttc ttg tac aag gat Leu Phe Phe Gly Cys Arg Lys Ser Asp Glu Asp Phe Leu Tyr Lys Asp 580 585 590	2014
95	gaa tgg aag act ttt cag gag cag ctt ggc gac tcg ctc aag atc atc Glu Trp Lys Thr Phe Gln Glu Gln Leu Gly Asp Ser Leu Lys Ile Ile 595 600 605 610	2062
100	act gcc ttc tct cgt gaa tcg gct gag aaa gtc tac gtc cag cac agg Thr Ala Phe Ser Arg Glu Ser Ala Glu Lys Val Tyr Val Gln His Arg 615 620 625	2110
105	ctg cgt gag cat gcc gag ctg gtc agt gac ctg ctg aag cag aaa gcc	2158

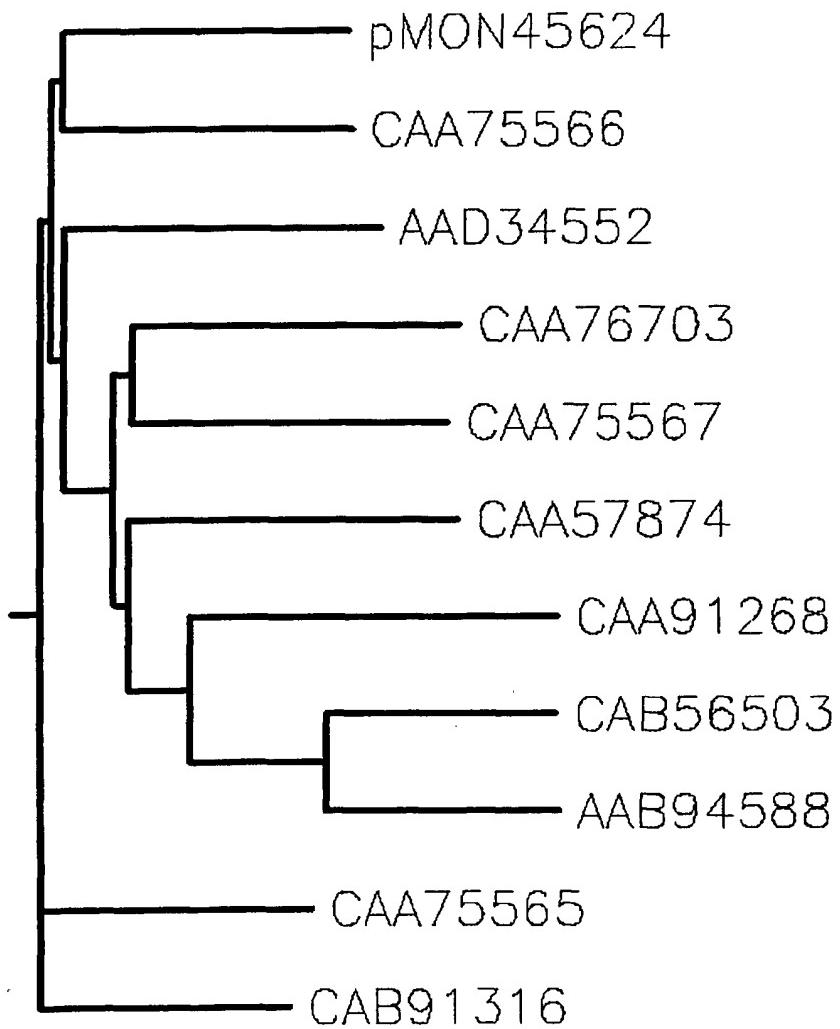
	Leu Arg Glu His Ala Glu Leu Val Ser Asp Leu Leu Lys Gln Lys Ala			
	630	635	640	
5	act ttc tat gtt tgc ggt gac gct gcc aac atg gcc cgt gaa gtc aac	2206		
	Thr Phe Tyr Val Cys Gly Asp Ala Ala Asn Met Ala Arg Glu Val Asn			
	645	650	655	
10	ctc gtg ctt ggg caa atc att gcc aag cag cgc ggt ctc cct gcc gag	2254		
	Leu Val Leu Gly Gln Ile Ile Ala Lys Gln Arg Gly Leu Pro Ala Glu			
	660	665	670	
15	aag ggc gag gag atg gtg aag cac atg cgc agc agc ggc agc tac cag	2302		
	Lys Gly Glu Glu Met Val Lys His Met Arg Ser Ser Gly Ser Tyr Gln			
	675	680	685	690
	gac gat gtc tgg tcc taa aa	2322		
	Asp Asp Val Trp Ser *			
	695			

Figure 4 - Amino acid homology alignment of A. ochraceus 11 alpha hydroxylase with the top 10 BLAST hits from GenBank

5	CAA75565 CAB91316 CAB56503 AAB94588 pMON45624 CAA75566 AAD34552 CAA75567 CAA76703 CAA57874 CAA91268	1 ---MANHSSYYHEFYKDHSHHTVLLMSEKPVILPSLILGTCAVLLCIQWLK--PQPLIM 1 -----MERLDIKSICDPSATPSYLVTAFLLAVVVMSLQGPRF-PKNIKH 1 -----LLFCFILSKTTKRGQNSQYS-NHDELP 1 -----MVMELHNHTPFSIYFITSILFIFVFFKLVQRS--DSKTS-STCKLP 1 -----MPFFTGLLAIYHSILILDNEVQTLTIVLAAAYN-LATLQ--PSDLPE 1 -----MSIFNMITSYAGSQLLPPFYIAIFVFTLVPWAIR-SWLELRK-GSVVPL 1 ---MTVDALTQPHHLLSLAWNDTQOHGSWFAPLVTTSAGLLCLLNLCSSGR--RSDLPV 1 ----- 1 MSKSNSMNNTSHETLFQQQLVLGLDRMPLMDVHWLIYVAFGAWLCSIVHVLSSSTVKP 1 ----- 1 MALLILSSLVISIFTFFIYIILRARERKLREKIGLSGPEPH	
10	CAA75567 CAA76703 CAA57874 CAA91268	1 ----- 1 MSKSNSMNNTSHETLFQQQLVLGLDRMPLMDVHWLIYVAFGAWLCSIVHVLSSSTVKP 1 ----- 1 MALLILSSLVISIFTFFIYIILRARERKLREKIGLSGPEPH	
15	CAA75565 CAB91316 CAB56503 AAB94588 pMON45624 CAA75566 AAD34552 CAA75567 CAA76703 CAA57874 CAA91268	56 VNGRKFGELSNVRAARDFITFGARQILEKSLKMSPKPFPRINGDVGEGHLPLIKYVFRN 45 LNKGPLEFSDTPINKEFVYGSRQULANFKANPKPCVNSIDFGEALVLFPRMNEETNN 28 PGIPQIPILGNAEOLSGGH-TheTURDLKKYGPMLHKK-IGEVSTIVASSQIQEEETFR 45 PGIPRTLPLIGNIQIVGSLPVHYYTKNLADKYGPMHLK-IGEVVSMILIVTS-EMQFIMK 46 LNAKPKFETTNRRRVHEFVENSNSLLARERLHGHEPYFLMSENGSLIVLPLPECDFLN 48 ANIPD-SLGFTGKQHRSVVKLSRPILAKERSLFPNEPFPLTITNGEVLLPLIDFDFDFRN 56 FNKTKTWWELTTMRARRDIDANAPSWSLESWFSQN-DKPIFIVESGYCTILSSMID-FSK 1 ----- 61 VVGYRSVVFEPITWLLRLR-----WEGGS-----IIGQSYNKFDSIFQVRKLGTDVVIIHNYIDEVK 1 ----- 43 WFLGNLKQTAERENLGMDDANWFNELHEQYGETFGIY-YGSQMNTIVSNEKDIKFWFI	
20	CAA75565 CAB91316 CAB56503 AAB94588 pMON45624 CAA75566 AAD34552 CAA75567 CAA76703 CAA57874 CAA91268	116 NEKISFTMAA--FKWFYAHLPGEIG--FREGTNESHIMKLARHQLT--HOLTLATGAS 105 DDRISFTTRWTF--YKAPEGHLPGEIG--FGEASRESHINQIMRDLT--HYLNKWTETPA 86 THDILFADRPSNLESKIVSYDSDMVSPYGNWQDFRISMMELSQHSQSFRSIRE 104 THDLNFSDRP-DFVLSRIVSYNGSGIVFSOHGDYWRQFRKCTVETLTAAVRQSFIRSIRE 106 DPMDFETPT--DTSDFGYIPEHADA--LN-ADPNLTW-VTKYL--KALNKITAPIS 107 DPRUSFSKAA--MQDNHAGIPDFET--VALVGREDOLQFARKQLC--KHLSAVIEPSS 115 MKPLACMYKFLG--TDFHSHLPEDOG--FKEVTRDAHLYFVVMNQFO--TQAPKVVKPA 121 LS---QDKTRSVEPFINDFAQYT--RGMVFLQSDLQMNVIQQRLE--PKLVSUTKVK 1 ----- 102 KNFSNEDSRD--VPSIYEANQLTASLLMNSYSSGWKHTRSAIAPIFS-TGKOKANOETIN	
25	CAA75565 CAB91316 CAB56503 AAB94588 pMON45624 CAA75566 AAD34552 CAA75567 CAA76703 CAA57874 CAA91268	116 NEKISFTMAA--FKWFYAHLPGEIG--FREGTNESHIMKLARHQLT--HOLTLATGAS 105 DDRISFTTRWTF--YKAPEGHLPGEIG--FGEASRESHINQIMRDLT--HYLNKWTETPA 86 THDILFADRPSNLESKIVSYDSDMVSPYGNWQDFRISMMELSQHSQSFRSIRE 104 THDLNFSDRP-DFVLSRIVSYNGSGIVFSOHGDYWRQFRKCTVETLTAAVRQSFIRSIRE 106 DPMDFETPT--DTSDFGYIPEHADA--LN-ADPNLTW-VTKYL--KALNKITAPIS 107 DPRUSFSKAA--MQDNHAGIPDFET--VALVGREDOLQFARKQLC--KHLSAVIEPSS 115 MKPLACMYKFLG--TDFHSHLPEDOG--FKEVTRDAHLYFVVMNQFO--TQAPKVVKPA 121 LS---QDKTRSVEPFINDFAQYT--RGMVFLQSDLQMNVIQQRLE--PKLVSUTKVK 1 ----- 102 KNFSNEDSRD--VPSIYEANQLTASLLMNSYSSGWKHTRSAIAPIFS-TGKOKANOETIN	
30	CAA75565 CAB91316 CAB56503 AAB94588 pMON45624 CAA75566 AAD34552 CAA75567 CAA76703 CAA57874 CAA91268	170 EECALVVKWVYTDSP--EHDITAKDANNKLMRITSVFLSKDNCRDPOMIRH--STIA 159 QETSMARAEANLPLKAANGE-STINERSKILPFIVARISSEFVLDCEPILCRPTEPIMKVI-QOAT 146 EEVLNPFLKSLG--SKEG-TRUNLSEKNSLILYLTTHAAFEKKNKNTPEFIRLDQLTK 163 EEAELVKKIAATASEEGGSIIFNTQSTOSYIYMTFC-IAAAFAFUKKSRYCQVEFISNNHKQLM 156 HFASTIAKAVLGDDP-DIRETYPARDLQLVFMSTFVFLGEDMCNODIOTNS-SOIA 161 RESTLAWSLNFGETT--EKAIRKPALEDTIARISSTPYLQDOLCRKAEALKI-KTAT 169 NEASGIITLFGDSN-EWHTVWPYNOCDLWVITVIFIAW-SKIAHDEEMIDUA-KHIA 17 MTKSFRWPRTS--KSSVSYDMMERTVVALESQ-AFVULPLCRDEGLQAS-IGYT 50 CAA75567 CAA76703 CAA57874 CAA91268	170 EECALVVKWVYTDSP--EHDITAKDANNKLMRITSVFLSKDNCRDPOMIRH--STIA 159 QETSMARAEANLPLKAANGE-STINERSKILPFIVARISSEFVLDCEPILCRPTEPIMKVI-QOAT 146 EEVLNPFLKSLG--SKEG-TRUNLSEKNSLILYLTTHAAFEKKNKNTPEFIRLDQLTK 163 EEAELVKKIAATASEEGGSIIFNTQSTOSYIYMTFC-IAAAFAFUKKSRYCQVEFISNNHKQLM 156 HFASTIAKAVLGDDP-DIRETYPARDLQLVFMSTFVFLGEDMCNODIOTNS-SOIA 161 RESTLAWSLNFGETT--EKAIRKPALEDTIARISSTPYLQDOLCRKAEALKI-KTAT 169 NEASGIITLFGDSN-EWHTVWPYNOCDLWVITVIFIAW-SKIAHDEEMIDUA-KHIA 17 MTKSFRWPRTS--KSSVSYDMMERTVVALESQ-AFVULPLCRDEGLQAS-IGYT 173 EEDYATKKEPMDMKNDENVEMDSSIWURLISEFISAFVLEPEH-RIOE-LTTA-ABPS 1 ----- 159 SKVDLFDIDIREKAS--SGQKWDYDDFQGTLTDVIGKCAFNIIDSNCDFRNDVFYHPVT
35	CAA75565 CAB91316 CAB56503 AAB94588 pMON45624 CAA75566 AAD34552 CAA75567 CAA76703 CAA57874 CAA91268	227 VIAFRAVEELRLINE-SWLRFVVQWEMPHCTQSPALVQEAFDILINELLER-RFEDV--AEA 218 EDGFGAAEDLRLW-EAACRFIMHWFL-SCQRAFADVRVRSILDEVLKA-RFQPA--AAN 202 AVAEPNIADMFPSL-KFLQLISTSKYKIEKIKHQDFDVIVETIEGHKKEJINKPLS--QEN 223 LLGGFSVADLYPSS-RVFOMMGATG-KLEKVHFVTDRLVLODIIIDEHK--NPNRS--SEE 213 ALAFGVGDMLRIVY-RMIRFIMHWFM-SCWELPRSLRRCQI2GTYH-RKSLR--GTT 60 CAA75566 AAD34552 CAA75567 CAA76703 CAA57874 CAA91268	227 VIAFRAVEELRLINE-SWLRFVVQWEMPHCTQSPALVQEAFDILINELLER-RFEDV--AEA 218 EDGFGAAEDLRLW-EAACRFIMHWFL-SCQRAFADVRVRSILDEVLKA-RFQPA--AAN 202 AVAEPNIADMFPSL-KFLQLISTSKYKIEKIKHQDFDVIVETIEGHKKEJINKPLS--QEN 223 LLGGFSVADLYPSS-RVFOMMGATG-KLEKVHFVTDRLVLODIIIDEHK--NPNRS--SEE 213 ALAFGVGDMLRIVY-RMIRFIMHWFM-SCWELPRSLRRCQI2GTYH-RKSLR--GTT 218 TNFYTASTLMEF--RSIRPLAHWFLECRKLPQERHDAGIYFLIFER-RFELR--RAA 226 VTMAIQARQLRLW-EVILRFLVHWFEOGAKLPQAOVRRAPQDIDFIIOE-RFAPB--DAC 70 VOCVSIRDQLFTWS-PVLRPFIIGFEL-SVRSVFRHLRFSAEIMALISQALQDPRQHRAD 232 ESLFITGFILRVVB-HILLRFLIAPLLFSYRFTLLRNWSSCPFRVIGDIIR--SQQ-- 1 ----- 217 VKITINNPTYFHSSSPGTTFESTLQIHTTGFCRNSTCPTVKCGFQDKAKFCSDYE
40	CAA75565 CAB91316 CAB56503 AAB94588 pMON45624 CAA75566 AAD34552 CAA75567 CAA76703 CAA57874 CAA91268	227 VIAFRAVEELRLINE-SWLRFVVQWEMPHCTQSPALVQEAFDILINELLER-RFEDV--AEA 218 EDGFGAAEDLRLW-EAACRFIMHWFL-SCQRAFADVRVRSILDEVLKA-RFQPA--AAN 202 AVAEPNIADMFPSL-KFLQLISTSKYKIEKIKHQDFDVIVETIEGHKKEJINKPLS--QEN 223 LLGGFSVADLYPSS-RVFOMMGATG-KLEKVHFVTDRLVLODIIIDEHK--NPNRS--SEE 213 ALAFGVGDMLRIVY-RMIRFIMHWFM-SCWELPRSLRRCQI2GTYH-RKSLR--GTT 60 CAA75566 AAD34552 CAA75567 CAA76703 CAA57874 CAA91268	227 VIAFRAVEELRLINE-SWLRFVVQWEMPHCTQSPALVQEAFDILINELLER-RFEDV--AEA 218 EDGFGAAEDLRLW-EAACRFIMHWFL-SCQRAFADVRVRSILDEVLKA-RFQPA--AAN 202 AVAEPNIADMFPSL-KFLQLISTSKYKIEKIKHQDFDVIVETIEGHKKEJINKPLS--QEN 223 LLGGFSVADLYPSS-RVFOMMGATG-KLEKVHFVTDRLVLODIIIDEHK--NPNRS--SEE 213 ALAFGVGDMLRIVY-RMIRFIMHWFM-SCWELPRSLRRCQI2GTYH-RKSLR--GTT 218 TNFYTASTLMEF--RSIRPLAHWFLECRKLPQERHDAGIYFLIFER-RFELR--RAA 226 VTMAIQARQLRLW-EVILRFLVHWFEOGAKLPQAOVRRAPQDIDFIIOE-RFAPB--DAC 70 VOCVSIRDQLFTWS-PVLRPFIIGFEL-SVRSVFRHLRFSAEIMALISQALQDPRQHRAD 232 ESLFITGFILRVVB-HILLRFLIAPLLFSYRFTLLRNWSSCPFRVIGDIIR--SQQ-- 1 ----- 217 VKITINNPTYFHSSSPGTTFESTLQIHTTGFCRNSTCPTVKCGFQDKAKFCSDYE
45	CAA75565 CAB91316 CAB56503 AAB94588 pMON45624 CAA75566 AAD34552 CAA75567 CAA76703 CAA57874 CAA91268	227 VIAFRAVEELRLINE-SWLRFVVQWEMPHCTQSPALVQEAFDILINELLER-RFEDV--AEA 218 EDGFGAAEDLRLW-EAACRFIMHWFL-SCQRAFADVRVRSILDEVLKA-RFQPA--AAN 202 AVAEPNIADMFPSL-KFLQLISTSKYKIEKIKHQDFDVIVETIEGHKKEJINKPLS--QEN 223 LLGGFSVADLYPSS-RVFOMMGATG-KLEKVHFVTDRLVLODIIIDEHK--NPNRS--SEE 213 ALAFGVGDMLRIVY-RMIRFIMHWFM-SCWELPRSLRRCQI2GTYH-RKSLR--GTT 60 CAA75566 AAD34552 CAA75567 CAA76703 CAA57874 CAA91268	227 VIAFRAVEELRLINE-SWLRFVVQWEMPHCTQSPALVQEAFDILINELLER-RFEDV--AEA 218 EDGFGAAEDLRLW-EAACRFIMHWFL-SCQRAFADVRVRSILDEVLKA-RFQPA--AAN 202 AVAEPNIADMFPSL-KFLQLISTSKYKIEKIKHQDFDVIVETIEGHKKEJINKPLS--QEN 223 LLGGFSVADLYPSS-RVFOMMGATG-KLEKVHFVTDRLVLODIIIDEHK--NPNRS--SEE 213 ALAFGVGDMLRIVY-RMIRFIMHWFM-SCWELPRSLRRCQI2GTYH-RKSLR--GTT 218 TNFYTASTLMEF--RSIRPLAHWFLECRKLPQERHDAGIYFLIFER-RFELR--RAA 226 VTMAIQARQLRLW-EVILRFLVHWFEOGAKLPQAOVRRAPQDIDFIIOE-RFAPB--DAC 70 VOCVSIRDQLFTWS-PVLRPFIIGFEL-SVRSVFRHLRFSAEIMALISQALQDPRQHRAD 232 ESLFITGFILRVVB-HILLRFLIAPLLFSYRFTLLRNWSSCPFRVIGDIIR--SQQ-- 1 ----- 217 VKITINNPTYFHSSSPGTTFESTLQIHTTGFCRNSTCPTVKCGFQDKAKFCSDYE
50	CAA75567 CAA76703 CAA57874 CAA91268	173 EEDYATKKEPMDMKNDENVEMDSSIWURLISEFISAFVLEPEH-RIOE-LTTA-ABPS 1 ----- 159 SKVDLFDIDIREKAS--SGQKWDYDDFQGTLTDVIGKCAFNIIDSNCDFRNDVFYHPVT	
55	CAA75565 CAB91316 CAB56503 AAB94588 pMON45624 CAA75566 AAD34552 CAA75567 CAA76703 CAA57874 CAA91268	227 VIAFRAVEELRLINE-SWLRFVVQWEMPHCTQSPALVQEAFDILINELLER-RFEDV--AEA 218 EDGFGAAEDLRLW-EAACRFIMHWFL-SCQRAFADVRVRSILDEVLKA-RFQPA--AAN 202 AVAEPNIADMFPSL-KFLQLISTSKYKIEKIKHQDFDVIVETIEGHKKEJINKPLS--QEN 223 LLGGFSVADLYPSS-RVFOMMGATG-KLEKVHFVTDRLVLODIIIDEHK--NPNRS--SEE 213 ALAFGVGDMLRIVY-RMIRFIMHWFM-SCWELPRSLRRCQI2GTYH-RKSLR--GTT 60 CAA75566 AAD34552 CAA75567 CAA76703 CAA57874 CAA91268	227 VIAFRAVEELRLINE-SWLRFVVQWEMPHCTQSPALVQEAFDILINELLER-RFEDV--AEA 218 EDGFGAAEDLRLW-EAACRFIMHWFL-SCQRAFADVRVRSILDEVLKA-RFQPA--AAN 202 AVAEPNIADMFPSL-KFLQLISTSKYKIEKIKHQDFDVIVETIEGHKKEJINKPLS--QEN 223 LLGGFSVADLYPSS-RVFOMMGATG-KLEKVHFVTDRLVLODIIIDEHK--NPNRS--SEE 213 ALAFGVGDMLRIVY-RMIRFIMHWFM-SCWELPRSLRRCQI2GTYH-RKSLR--GTT 218 TNFYTASTLMEF--RSIRPLAHWFLECRKLPQERHDAGIYFLIFER-RFELR--RAA 226 VTMAIQARQLRLW-EVILRFLVHWFEOGAKLPQAOVRRAPQDIDFIIOE-RFAPB--DAC 70 VOCVSIRDQLFTWS-PVLRPFIIGFEL-SVRSVFRHLRFSAEIMALISQALQDPRQHRAD 232 ESLFITGFILRVVB-HILLRFLIAPLLFSYRFTLLRNWSSCPFRVIGDIIR--SQQ-- 1 ----- 217 VKITINNPTYFHSSSPGTTFESTLQIHTTGFCRNSTCPTVKCGFQDKAKFCSDYE
60	CAA75566 AAD34552 CAA75567 CAA76703 CAA57874 CAA91268	217 VKITINNPTYFHSSSPGTTFESTLQIHTTGFCRNSTCPTVKCGFQDKAKFCSDYE	
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	CAA75565	283	ERTKEKV-TYNTDAEVLLDDLAREK--GVGYDPACACLSLSVAA--LHSTTDFFTQWIFDI
	CAB91316	274	--SGKA-EHDIAAEFERTA--K--GKYYIPAVAVLVLSLVA--IHTTSDLTCOWITNL
	CAB56503	259	G--EKKE-DLVIDVILNIQRNDFE--EFLGCKKNIKAIIFNIFSACTETSSTTIVDWAICEN
5	AAB94588	276	R--EAVE-DLVDVILKFQKES--E--FRLTUDNNKAVIQDIFIGGGETSSSVVEWSESEL
	pMON45624	269	DEOKPL-MFDTSNEFEREL---GPNHIAVINKVTLISIVA--IHTTSDLHLQAMSDI
	CAA75566	274	IAAOPLPVFHADID--SSEQEAEAGTCAASFLPVMT-LTGSLLA--IHTTYDLLOOTMIDI
	AAD34552	282	RAKIEPPRYVLSIQFEDTAK---EKWYDAAGACLANDFAS--IYGTSDLIGSLVCI
	CAA75567	129	TLLSDQTEGRGTFIS:LLRHP-EELRTPEQVQGD:MLVFAA--IHTTITMALTWKWWEI
10	CAA76703	282	--EDGN--ECLLSMRDAATGEE-KQIDNIAORMLISIAS--IHTTAMTMTHCYDLYD
	CAA57874	44	--LEDPTTMLDHSNGRNHEIA---DVVELQLEHMTIAVE-TVTFSSSTTOQIYD
	CAA91268	277	RRKGEGSDSVTFLKLLNREDDKS-KPMTKQEWIENCFFLAGYEETTAFTYCSYL
	CAA75565	338	AONPELIEPLREELIAALGKQ-----ASFNSZYNLFLDNEVLFRESOFLY--E-----
15	CAB91316	324	NONPEFIAPLREEMIQUNSEG-----WKFISLYNMFLLDSEVKKSFVK--E-----
	CAB56503	314	IKNETVUMKAQEEFKRKIFNEEPN----VDETECHOLYQFLQVKEKETFLH--E-----
	AAB94588	329	INNEURMEKAQAFVRQYDYSKEY----VDETECHOLYKLYKEIIEKETFLH--E-----
	pMON45624	321	AONAKULCAVREEVRLUSTEC----LSPVUEHSIPLMESADMEWFLP--E-----
	CAA75566	332	GRHPEYIPLFQEVQLLREER----WKFITLUFKMLLESALKESOFMK--E-----
20	AAD34552	336	VRHHHLLEPLFDEIRYIIGOG----WTPASVYKLLDLSLKFDFVK--E-----
	CAA75567	186	VPRPFYIPLFTEQDIFGPDVSPDICINFEASRLKMLDFIR:VFVFC--E-----
	CAA76703	333	CAOPFYIPLFD2EKSAGAS----AD:TA:INRFKLDFLWRSJHFN--E-----
	CAA57874	96	WAHPEYIPLREESIPLDPNGN----ETD:TVAMDKLDFLWRSFFNS:DLSMSNL
	CAA91268	336	SKVFNWQAK:YEFTIMEAKENG----LTYDSEHNMYLCEVYKHTLFY--E-----
25	CAA75565	384	IAIASMREFTTHWKLSDVIL--INKLTLIS:FOHW----DEYKRE
	CAB91316	370	--TGVASMRIVAYEKEV-LDDTFIPFVAFIA:SFHDW----NSEVVEQ
	CAB56503	361	--PVPLLVPFECREOCKKIK-YT111SKSRVILINWAIJR----DNYNIE
30	AAB94588	376	--PVPLLVPFVSRCRQIN-EYELISKIRIILIN-WAIJR----NICKYRCE
	pMON45624	367	--TLLGSFRQATNIIKLKSIVFKVIIARWILDSTMW----N:BYTE
	CAA75566	378	--GSIVTMPEVVTETITLSSLTLLKKGIFLNUIDNRLLD----D:KINDN
	AAD34552	382	--VECATMFSYALQEVIFSNQTFLPKGEELVAA:DRMS--N:EVSE
	CAA75567	238	--STFVIPSFRVMKSMTLSNIKLCORG:SHAFF:HAHMSEETPTFS:DFSS
35	CAA76703	379	--VFLLIIFNIIYHOSMFLSDITNIISCTRIAPSHAMQ----DSAHVFG
	CAA57874	152	KNYKLCESLTGHNSNLPTRTIAMKIPDTFVFKCTKWEINTCSIIHK----DHKLHEN
	CAA91268	382	--PHFSFIRELCRELITR-QFYPHATIVCLPFAVHR----N:ENADS
40	CAA75565	427	ELK----FDGYRFPNMREP--G:ESKAQLVSATPHEMGFLY:LHN:PGRFH:SEHIFI
	CAB91316	413	AK----WDCRFLRMETPGAFENVAVOLVS:APBHLGFCH:OIACPGFPA:ANETFI
	CAB56503	404	PKK----FNPDPFLES----VDFKGKNSPFLYLPFGCRPITPCITPALANEI
	AAB94588	419	TBS----FKPBPFLNS----S-IDFRGIDFPIPJA:RHC:PCITH:IPN:EL
	pMON45624	410	FLO----VDCYFVFNFQHOTP--BEDKNALLVS:SANHMGFLCHVACPGFFF:SNELFI
45	CAA75566	421	FBV----WVPRFYDVFSEA:GHDGAQVLST:GNSNHMGFLCHQHSDPGRFFF:ANEFTIV
	AAD34552	425	EAK----VLFYFYLMRQFEDP--AFAFSQALENENG:HIGF:SWHPR-A:CPGRFFASKRIM
	CAA75567	289	FENPSPRFDCFFYLNLFPSIK--E:GSQHOQAH:GPFYLIENH:KHA:CPGRFAISCFIM
	CAA76703	423	FTP--PTEDGCFVSKLPSDS--NYAOKYLPMSIDSSNMAFGY:KY:CPGPFW:SNEMPI
	CAA57874	205	PBQ----FDCPFFHKWFKAP--E:EKRYMYSSSGTIDLSWF:RHACPGFYL:SAINF
50	CAA91268	425	PEE----HPEP:PEWEE----E----KSS:SLKWIPE:V:PEV:V:MR:MEM:PF
	CAA75565	480	AISHLILKVIPLPV----EGSSM:PRKYE:MNAN:TA:K:SV:REK:EE:AI----
55	CAB91316	468	AUHVLLINNE:RIP----EGSDPKIUTTFEPFSAGVDSLK:NEY:NGO:PEIEL----
	CAB56503	449	PAQLLFFHDQS----NTEKLNMKSE-SREV:V:R:EDDLY:TPVNFS:SSSPA----
	AAB94588	464	PAQOLLMFHDV:LPNKMKEELDMTE-SNC:TLR:R:QNDL:CE:IPITE----P----
	pMON45624	463	AUCHIILNNE:RIP----DGFKPQPLNIE:TYLA:DN:TR:ML:IR:PK-A:ED:MAS:LT:V
	CAA75566	474	AUCHILWVYDMFLC----PDTE:TKPDTR:MI:AKSS:VTD:LI:K:R:ES:V:ED:LE:AI--
60	AAD34552	478	MAYLLIIRYD:K:WV----PDEPLQY:YRHSF:SVR:HT:TR:LM:R:RED:ED:TR:R:PG:SL
	CAA75567	347	II:IELL:K:YDF:R:IE----DGKPGPELMRV:TE:TR:LD:TK:AGL:ED:R:R:R:
	CAA76703	479	TAI:LL:O:FE:K:U----DGKGRPRN:IT:DS:DI:PE:R:R:R:O:R:K:S--R:DE--
	CAA57874	258	IAEELL:Y:II:ELP----DGLSRPKNIE:FE:V:LAS:LN:CAN:CA
	CAA91268	468	TE:V:KL:LD:TE:EL:OF----EGEADLI:PDCN:EV:IM:RP:N:PV:R:LU:PN:EN
65	CAA75565	(SEQ ID NO: 27)	
	CAB91316	(SEQ ID NO: 28)	
	CAB56503	(SEQ ID NO: 29)	
	AAB94588	(SEQ ID NO: 30)	
70	pMON45624	(SEQ ID NO: 02)	
	CAA75566	(SEQ ID NO: 31)	
	AAD34552	(SEQ ID NO: 32)	
	CAA75567	(SEQ ID NO: 33)	
	CAA76703	(SEQ ID NO: 34)	
	CAA57874	(SEQ ID NO: 35)	
75	CAA91268	(SEQ ID NO: 36)	

**Figure 5 - Phylogenetic tree showing the relatedness of
Aspergillus ochraceus 11 alpha hydroxylase to the top 10
BLAST hits from GenBank**



**Figure 6 – Percent homology of *Aspergillus ochraceus* 11
alpha hydroxylase to the top 10 BLAST hits from GenBank**

Accession Number	Species	% ID to 11a OH
CAB91316	<i>Neurospora crassa</i>	40
CAA76565	<i>Gibberella fujikuroi</i>	37
CAA75566	<i>Gibberella fujikuroi</i>	37
AAD34552	<i>Aspergillus terreus</i>	29
CAA75567	<i>Gibberella fujikuroi</i>	24
CAA57874	<i>Fusarium oxysporum</i>	24
CAA76703	<i>Gibberella fujikuroi</i>	23
CAB56503	<i>Catharanthus roseus</i>	14
AAB94588	<i>Glycine max</i>	14
CAA91268	<i>Caenorhabditis elegans</i>	12

Figure 7 - Amino acid homology alignment of *A. ochraceus* and human oxidoreductase to NADPH cytochrome P450 reductases from *A. niger*, mouse, and *S. cerevisiae*

5	PMON45605	1	MGDS!!WDT!ST!SEA!AEE!SLR!SM!TIN!LFS!LIV!LIV!WFL!WVK!V!F!E!T!H!L!	human	1	MGDS!WDT!ST!SEA!AEE!VS!SMT!IN!LFS!LIV!LIV!WFL!WVK!V!F!E!T!H!L!
	mouse	1	MGDS!WDT!SA!M!PEA!AEE!VS!SMT!IN!LFS!LIV!LIV!WFL!WVK!V!F!E!T!H!L!		1	MGDS!WDT!SA!M!PEA!AEE!VS!SMT!IN!LFS!LIV!LIV!WFL!WVK!V!F!E!T!H!L!
	pMON45632	1	--MAQD!L!LCLV!LVALLVGVWAYF!KG-----TYI!AAG!DPYA--SAGPANNGG	niger	1	--MAQD!L!LCLV!LVALLVGVWAYF!KG-----TYI!AAG!DPYA--SAGPANNGG
	yeast	1	-MPFG!L!N!EFT!L!GLVLAV!LYU!KRN-----SIKE!LM!SDDG----DT-		1	-MPFG!L!N!EFT!L!GLVLAV!LYU!KRN-----SIKE!LM!SDDG----DT-
10						
	PMON45605	61	TSS!V!RE!S!F!V!E!P!M!K!P!G!R!N!I!W!F!Y!G!S!G!T!G!T!A!E!F!A!U!L!S!E!-A!R!Y!M!P!C!I!A!C!E!H!E!V!	human	61	TSS!V!RE!S!F!V!E!P!M!K!P!G!R!N!I!W!F!Y!G!S!G!T!G!T!A!E!F!A!U!L!S!E!-A!R!Y!M!P!C!I!A!C!E!H!E!V!
	mouse	61	PP!P!V!E!S!F!V!E!P!M!K!P!G!R!N!I!W!F!Y!G!S!G!T!G!T!A!E!F!A!U!L!S!E!-A!R!Y!M!P!C!I!A!C!E!H!E!V!		61	PP!P!V!E!S!F!V!E!P!M!K!P!G!R!N!I!W!F!Y!G!S!G!T!G!T!A!E!F!A!U!L!S!E!-A!R!Y!M!P!C!I!A!C!E!H!E!V!
15	pMON45632	47	E!R!G!Y!T!R!D!I!Q!M!D!E!T!G!K!C!W!F!Y!G!S!G!T!G!T!A!E!F!A!U!L!S!E!-A!R!Y!M!P!C!I!A!C!E!H!E!V!	niger	47	E!R!G!Y!T!R!D!I!Q!M!D!E!T!G!K!C!W!F!Y!G!S!G!T!G!T!A!E!F!A!U!L!S!E!-A!R!Y!M!P!C!I!A!C!E!H!E!V!
	yeast	42	E!V!S!G!N!R!D!A!V!U!T!E!N!K!Y!L!U!A!S!O!T!A!E!D!Y!K!U!S!E!L!V!A!P!N!U!V!C!A!L!V!E!N!Y!		42	E!V!S!G!N!R!D!A!V!U!T!E!N!K!Y!L!U!A!S!O!T!A!E!D!Y!K!U!S!E!L!V!A!P!N!U!V!C!A!L!V!E!N!Y!
20	PMON45605	120	A!D!L!S!S!L!P!E!I!F!N!A!L!U!V!F!C!H!A!T!Y!G!E!G!D!P!T!D!A!Q!F!Y!L!W!-----D!V!D!L!S!-V!F!	human	120	A!D!L!S!S!L!P!E!I!F!N!A!L!U!V!F!C!H!A!T!Y!G!E!G!D!P!T!D!A!Q!F!Y!L!W!-----D!V!D!L!S!-V!F!
	mouse	120	A!D!L!S!S!L!P!E!I!F!N!A!L!U!V!F!C!H!A!T!Y!G!E!G!D!P!T!D!A!Q!F!Y!L!W!-----D!V!D!L!S!-V!F!		120	A!D!L!S!S!L!P!E!I!F!N!A!L!U!V!F!C!H!A!T!Y!G!E!G!D!P!T!D!A!Q!F!Y!L!W!-----D!V!D!L!S!-V!F!
	pMON45632	107	E!N!U!E!K!F!P!-D!V!F!P!L!A!T!Y!G!E!B!E!T!E!I!A!V!E!P!Q!F!G!D!D!V!A!P!S!-A!S!A!D!E!K!P!-N!Y!	niger	107	E!N!U!E!K!F!P!-D!V!F!P!L!A!T!Y!G!E!B!E!T!E!I!A!V!E!P!Q!F!G!D!D!V!A!P!S!-A!S!A!D!E!K!P!-N!Y!
25	yeast	102	E!S!L!N!D!V!-V!-I!N!S!P!I!S!Y!G!E!G!U!P!F!G!-V!N!S!E!L!F!C!N!-A!A!A!G!-----A!L!N!Y!		102	E!S!L!N!D!V!-V!-I!N!S!P!I!S!Y!G!E!G!U!P!F!G!-V!N!S!E!L!F!C!N!-A!A!A!G!-----A!L!N!Y!
30	PMON45605	169	A!V!F!G!L!G!N!K!T!Y!E!H!F!N!A!M!Q!Y!W!D!F!L!E!Q!L!J!A!P!R!F!P!L!U!S!E!-D!N!E!E!F!T!R!E!F!H!A!	human	169	A!V!F!G!L!G!N!K!T!Y!E!H!F!N!A!M!Q!Y!W!D!F!L!E!Q!L!J!A!P!R!F!P!L!U!S!E!-D!N!E!E!F!T!R!E!F!H!A!
	mouse	169	A!V!F!G!L!G!N!K!T!Y!E!H!F!N!A!M!Q!Y!W!D!F!L!E!Q!L!J!A!P!R!F!P!L!U!S!E!-D!N!E!E!F!T!R!E!F!H!A!		169	A!V!F!G!L!G!N!K!T!Y!E!H!F!N!A!M!Q!Y!W!D!F!L!E!Q!L!J!A!P!R!F!P!L!U!S!E!-D!N!E!E!F!T!R!E!F!H!A!
	pMON45632	165	V!F!G!L!C!N!I!N!Y!E!H!Y!N!A!V!R!N!A!A!L!T!E!G!A!Q!R!G!S!A!E!G!D!A!-A!T!M!H!L!F!L!A!K!P!M!A!L!	niger	164	V!F!G!L!C!N!I!N!Y!E!H!Y!N!A!V!R!N!A!A!F!K!Q!P!R!G!S!A!E!G!D!A!-A!T!M!H!L!F!L!A!K!P!M!A!L!
	yeast	148	N!P!O!L!G!I!S!Y!S!F!F!I!C!A!P!K!A!P!H!S!A!A!G!I!P!G!K!L!D!E!M!B!A!T!T!D!Y!N!A!K!E!S!I!L!E!V!L!		148	N!P!O!L!G!I!S!Y!S!F!F!I!C!A!P!K!A!P!H!S!A!A!G!I!P!G!K!L!D!E!M!B!A!T!T!D!Y!N!A!K!E!S!I!L!E!V!L!
35	PMON45605	228	C!D!H!F!G!V!E!A!T!G!E!E!S!I!R!Y!E!L!V!-H!T!-----I!C!A!S!K!V!T!N!B!M!S!P!L!F!S!-----E!N!P!P	human	228	C!D!H!F!G!V!E!A!T!G!E!E!S!I!R!Y!E!L!V!-H!T!-----I!C!A!S!K!V!T!N!B!M!S!P!L!F!S!-----E!N!P!P
	mouse	228	C!D!H!F!G!V!E!A!T!G!E!E!S!I!R!Y!E!L!V!-H!T!-----I!C!A!S!K!V!T!N!B!M!S!P!L!F!S!-----E!N!P!P		228	C!D!H!F!G!V!E!A!T!G!E!E!S!I!R!Y!E!L!V!-H!T!-----I!C!A!S!K!V!T!N!B!M!S!P!L!F!S!-----E!N!P!P
40	pMON45632	225	S!E!A!M!N!L!G!-B!R!D!A!V!P!P!V!N!T!E!C!E!S!L!S!P!E!D!E!V!Y!L!C!P!T!Q!G!L!Q!-----G!P!G	niger	224	S!F!S!M!D!L!P!-B!R!A!Y!P!P!V!C!T!F!E!N!S!L!S!P!E!D!E!V!Y!L!C!P!T!Q!G!L!Q!-----G!P!G
	yeast	208	K!E!E!L!H!L!L!-E!G!E!A!K!F!T!S!C!F!Q!Y!T!V!L!N!-E!I!T!D!S!M!S!L!G!P!S!A!H!Y!L!P!S!H!Q!L!N!R!N!A!D!G!-L!G!		208	K!E!E!L!H!L!L!-E!G!E!A!K!F!T!S!C!F!Q!Y!T!V!L!N!-E!I!T!D!S!M!S!L!G!P!S!A!H!Y!L!P!S!H!Q!L!N!R!N!A!D!G!-L!G!
45	PMON45605	275	P!F!D!A!K!U!P!F!L!-A!V!T!I!N!K!L!N!Q!G!T!E!H!I!M!H!L!E!L!D!I!S!D!E!K!Y!E!S!J!D!H!V!A!Y!P!A!N!S!A!V!L!N!L!	human	275	P!F!D!A!K!U!P!F!L!-A!V!T!I!N!K!L!N!Q!G!T!E!H!I!M!H!L!E!L!D!I!S!D!E!K!Y!E!S!J!D!H!V!A!Y!P!A!N!S!A!V!L!N!L!
	mouse	275	P!F!D!A!K!U!P!F!L!-A!V!T!I!N!K!L!N!Q!G!T!E!H!I!M!H!L!E!L!D!I!S!D!E!K!Y!E!S!J!D!H!V!A!Y!P!A!N!S!A!V!L!N!L!		275	P!F!D!A!K!U!P!F!L!-A!V!T!I!N!K!L!N!Q!G!T!E!H!I!M!H!L!E!L!D!I!S!D!E!K!Y!E!S!J!D!H!V!A!Y!P!A!N!S!A!V!L!N!L!
	pMON45632	273	P!V!S!-H!N!P!F!L!-P!I!S!E!S!E!L!F!N!V!K!D!E!N!C!L!N!M!E!I!S!I!A!G!S!N!L!T!Q!T!S!H!I!A!W!T!A!G!S!E!D!R!	niger	272	P!V!S!-H!N!P!F!L!-P!I!S!E!S!E!L!F!T!V!K!D!E!N!C!L!N!M!E!I!S!I!A!G!S!N!L!T!Q!T!S!H!I!A!W!T!A!G!S!E!D!R!
	yeast	262	P!D!E!L!S!Q!Y!I!P!T!V!K!S!E!L!F!S!S!N!D!F!N!C!L!H!S!F!F!L!S!G!E!N!K!I!S!T!D!H!L!A!W!F!S!P!L!E!K!E!Q!		262	P!D!E!L!S!Q!Y!I!P!T!V!K!S!E!L!F!S!S!N!D!F!N!C!L!H!S!F!F!L!S!G!E!N!K!I!S!T!D!H!L!A!W!F!S!P!L!E!K!E!Q!
50	PMON45605	335	G!K!L!G!A!-L!D!V!V!M!S!L!N!L!D!E!S!N!K!H!P!F!C!P!T!S!Y!F!P!T!A!L!T!Y!L!D!I!T!H!P!R!T!N!V!L!Y!E!L!A!	human	335	G!K!L!G!A!-L!D!V!V!M!S!L!N!L!D!E!S!N!K!H!P!F!C!P!T!S!Y!F!P!T!A!L!T!Y!L!D!I!T!H!P!R!T!N!V!L!Y!E!L!A!
	mouse	335	G!K!L!G!A!-L!D!V!V!M!S!L!N!L!D!E!S!N!K!H!P!F!C!P!T!S!Y!F!P!T!A!L!T!Y!L!D!I!T!H!P!R!T!N!V!L!Y!E!L!A!		335	G!K!L!G!A!-L!D!V!V!M!S!L!N!L!D!E!S!N!K!H!P!F!C!P!T!S!Y!F!P!T!A!L!T!Y!L!D!I!T!H!P!R!T!N!V!L!Y!E!L!A!
55	pMON45632	333	L!Q!A!F!C!P!E!G!K!R!H!S!V!N!I!K!G!I!-V!T!A!V!P!F!I!P!T!T!Y!D!A!V!Y!L!E!V!C!A!V!S!R!F!U!S!-A!F!	niger	332	L!Q!W!F!S!E!L!G!K!R!S!V!N!I!K!G!I!-V!T!A!V!P!F!I!P!T!T!Y!D!A!V!Y!L!E!V!C!A!V!S!R!F!U!S!-A!F!
	yeast	322	L!S!F!N!L!D!-P!E!T!F!D!K!P!L!-P!T!V!W!P!F!E!T!P!T!T!G!A!I!K!H!Y!E!I!G!V!S!R!O!F!S!S!I!F!		322	L!S!F!N!L!D!-P!E!T!F!D!K!P!L!-P!T!V!W!P!F!E!T!P!T!T!G!A!I!K!H!Y!E!I!G!V!S!R!O!F!S!S!I!F!
60	PMON45605	393	A!S!-E!P!S!E!Q!L!R!K!M!A!S!S!S!C!E!G!K!E!L!Y!L!S!V!N!V!E!A!R!R!H!I!A!L!Q!C!F!-S!L!P!P!I!D!H!L!C!E!L!P!	human	393	A!S!-E!P!S!E!Q!L!R!K!M!A!S!S!S!C!E!G!K!E!L!Y!L!S!V!N!V!E!A!R!R!H!I!A!L!Q!C!F!-S!L!P!P!I!D!H!L!C!E!L!P!
	mouse	393	A!S!-E!P!S!E!Q!L!R!K!M!A!S!S!S!C!E!G!K!E!L!Y!L!S!V!N!V!E!A!R!R!H!I!A!L!Q!C!F!-S!L!P!P!I!D!H!L!C!E!L!P!		393	A!S!-E!P!S!E!Q!L!R!K!M!A!S!S!S!C!E!G!K!E!L!Y!L!S!V!N!V!E!A!R!R!H!I!A!L!Q!C!F!-S!L!P!P!I!D!H!L!C!E!L!P!
	pMON45632	391	A!P!D!E!A!K!E!I!V!F!L!G!E!D!K!Y!F!K!E!K!T!N!R!C!F!N!I!Q!-A!Q!S!I!T!S!K!-F!T!A!V!-F!S!S!L!I!E!G!I!T!K!	niger	390	A!P!M!R!K!-P!C!L!C!V!W!A!G!-L!F!P!E!G!H!O!P!M!-Q!Q!-A!Q!S!I!T!S!K!-F!S!A!V!F!S!S!L!I!E!G!I!T!K!
	yeast	378	A!P!N!A!D!V!R!E!K!L!T!L!S!K!D!K!Q!F!E!V!E!I!T!S!K!I!F!N!I!D!-A!K!Y!L!S!G!A!K!W!D!N!V!E!Q!F!V!S!P!Q!		378	A!P!N!A!D!V!R!E!K!L!T!L!S!K!D!K!Q!F!E!V!E!I!T!S!K!I!F!N!I!D!-A!K!Y!L!S!G!A!K!W!D!N!V!E!Q!F!V!S!P!Q!
70	PMON45605	451	L!Q!A!R!Y!Y!S!I!A!S!S!S!K!V!H!N!S!V!H!I!C!A!V!V!E!Y!E!T!K!-A!G!R!I!N!K!G!W!A!T!N!J!L!R!A!K!E!-A!F!-	human	451	L!Q!A!R!Y!Y!S!I!A!S!S!S!K!V!H!N!S!V!H!I!C!A!V!V!E!Y!E!T!K!-A!G!R!I!N!K!G!W!A!T!N!J!L!R!A!K!E!-A!F!-
	mouse	451	L!Q!A!R!Y!Y!S!I!A!S!S!S!K!V!H!N!S!V!H!I!C!A!V!V!E!Y!E!T!K!-A!G!R!I!N!K!G!W!A!T!N!J!L!R!A!K!E!-A!F!-		451	L!Q!A!R!Y!Y!S!I!A!S!S!S!K!V!H!N!S!V!H!I!C!A!V!V!E!Y!E!T!K!-A!G!R!I!N!K!G!W!A!T!N!J!L!R!A!K!E!-A!F!-
	pMON45632	448	L!Q!P!R!Y!Y!S!I!S!S!S!L!V!Q!D!K!I!S!I!T!A!V!E!S!V!R!L!P!-G!S!H!M!V!K!G!-T!T!N!Y!L!L!L!K!Q!K!N!E!R!S!	niger	446	L!Q!P!R!Y!Y!S!I!S!S!S!L!V!Q!D!K!I!S!I!T!A!V!E!S!V!R!L!P!-G!S!H!M!V!K!G!-T!T!N!Y!L!L!L!K!Q!K!N!E!R!S!
	yeast	436	Y!P!T!R!Y!Y!S!I!S!S!S!L!S!E!K!O!V!H!V!T!S!T!V!E!N!F!P!N!P!E!L!P!-D!I!P!P!G!V!W!T!M!L!F!N!I!C!A!C!N!V!N!Y!		436	Y!P!T!R!Y!Y!S!I!S!S!S!L!S!E!K!O!V!H!V!T!S!T!V!E!N!F!P!N!P!E!L!P!-D!I!P!P!G!V!W!T!M!L!F!N!I!C!A!C!N!V!N!Y!

	PMON45605	503	-----NGGF-----ALVEMFVRKSQFRLFFPATTPVINVGPCTGVAPFFIGFICF
	human	503	-----NGGF-----ALVEMFVRKSQFRLFFPATTPVINVGPCTGVAPFFIGFICF
	mouse	503	-----NORF-----ALVEMFVRKSQFRLFFPATTPVINVGPCTGVAPFFIGFICF
5	pMON45632	505	PDPHG-LTYSITGPPNKYDG1HVEVHVFHSNFKLFSI-SRPVIMVGPGTVAVPFRCFIG
	niger	503	SRPSR-LDLLHHGCPNNKYDG1HVEVHVFHSNFKLFSI-SRPVIMVGPGTVAVPFRCFIG
	yeast	495	AETNLPAHYDLANGPEKLFANYKEFVH/ERSNFRLESNPSTPVIMIGPGTVAVFFPGFIRF
	PMON45605	547	R-AWLFQ---QSEFI---AGETLLYTGCPRPSDELYLTFELQFHR-CALTQDINIAFSEF-
10	human	547	R-AWLFQ---QSEFI---AGETLLYTGCPRPSDELYLTFELQFHR-CALTQDINIAFSEF-
	mouse	547	R-AWLFQ---QSEFI---AGETLLYTGCPRPSDELYLTFELQFHR-CALTQDINIAFSEF-
	pMON45632	564	R-AAIAA---KSEK---WTMFFECDFKSDDEFLYLKDWWKTTQEQLWDSLKHITAFSEF-
	niger	562	R-AAIAA---KSEK---WTMFFECDFKSDDEFLYLKDWWKTYQDQWADNLKHITAFSEF-
15	yeast	555	PVAFLSQKKGNNNVSLCKHILFWNSNTF-CFLWQDWPEVAKKUDSFEQEVVAAHSLP
	PMON45605	599	[SHKVTVQHLLPQDREELWKL]-EGGAHIIYVCGCAFIRARADVQNTF[EDIVARLFAAEKA]
	human	599	[SHKVTVQHLLPQDREELWKL]-EGGAHIIYVCGCAFIRARADVQNTF[EDIVARLFAAEKA]
	mouse	599	[AHKVTVQHLLPQDREELWKL]-EGGAHIIYVCGCAFIRARADVQNTF[EDIVARLFAAEKA]
20	pMON45632	617	SAEIVVYVNR-BEHAEILSD-L-KOK-TFVCGSMIAAPE[MLVLGQIINKQRGELPAEK]
	niger	615	GPOVIVVHRLREHS-LMSD-L-KOK-TFVCGSMIAAPE[MLVLGQIINKQRGELPAEK]
	yeast	614	NTKHEVIVIDKLPDYEDQVTEAI-NNESFTIVDIDBARGENG-STALVGFLSRGKSITTD
	PMON45605	658	AVD-IKPLNTHGRISLDVAV-
	human	658	AVD-IKPLNTHGRISLDVAV-
	mouse	659	AVD-IKPLNTHGRISLDVAV-
	pMON45632	676	GEPMVPHNRSSTSIQEDVAV-
	niger	674	GEPMVPHNRSSTSIQEDVAV-
30	yeast	673	ATELIIMMKLISGPQEDVAV-
	PMON45605	(SEQ ID NO: 03)	
	human	(SEQ ID NO: 06)	
	mouse	(SEQ ID NO: 39)	
35	pMON45632	(SEQ ID NO: 05)	
	niger	(SEQ ID NO: 38)	
	yeast	(SEQ ID NO: 37)	

Figure 8 – Amino acid homology alignment of A. ochraceus oxidoreductase to NADPH cytochrome P450 reductases from A. niger and S. cerevisiae

		1 -MSAATLDDWFLVLLVGS/ATFFETTIVAVTRMLPAIRKQ-AVAKAFHNDIEK
5	A.niger	1 -MSAATLDDWFLVLLVGS/ATFFETTIVAVTRMLPAIRKQ-AVAKAFHNDIEK
	A.ochraceus	1 -MADIDLDLV/LVAILLVGSVAYETKETIVAVAFDPYSSAFAAMGAKAVYTRDVAQH
	S.cerevisiae	1 MPFGHLINTPTVLAEGVLALVILVKRNSIKPLLMSDDGDTAVSS-----NPDAAVW
	A.niger	60 BETGKNCVIFYGSQTCGAEDIASRLAKEGSQRPGGLKTM/ADLEEVYENLIDQFPEDKIAF
10	A.ochraceus	60 DETGKNCVIFYGSQTCGAEDIASRLAKEGSQRPGGLKTM/ADLFDPYDFEELKFPEDEVV+
	S.cerevisiae	55 TEENKLYLTAASQTCGAEDYKPFSEFLVAKENLNUCVIYVNLIFFSNDV,-V-VLS
	A.niger	120 FVLATYGEGETEHAWEFTHFEDIAVAFEN-ASAKIEMPLKQYTAQFGLKQHNTWVETHW
	A.ochraceus	120 FVLATYGEGETEHAWEFTHFEDIAVAFEN-GATADEPFLSFWVTFGLKQHNTWVETHW
	S.cerevisiae	113 IFISTAGGEDEPHCAGNEDFIC-----NEAGALINRIMMCLISSTIFFG
15	A.niger	179 MVFQVDA-FQTL-BPFIGSAGSEGDDACTNEEDFLAMPFEDQWALPSD-BEDFNGT
	A.ochraceus	180 MVFNVDAALT-BFACRIGSACEGDDACTNEEDFLAMPFEDQWALPSAQNQEDVATB
	S.cerevisiae	163 AAKKAHKHLSAAAGLIL-KLCEADDEACTNEEDFLAMPFEDQWALPSILEVKEEFLHDEQSKETS
20	A.niger	239 VFCVTENESLSPEDETYYLGEPTSHL-----AEPGDTSAKHFIAIAESHEI
	A.ochraceus	240 VFNVTDEESLSPEEENVTLGEPTVCHL-----EPFGPYAHKHFIAPIASMEPEI
	S.cerevisiae	223 QFCYV---V-NEITDSMSEPEHMYPSHQLNRRNADTQLEAFDLSDQEVIALAPVKSPEI
25	A.niger	290 FTVKDRNCIHLWEISIAGSNSLSTGTGDHIAVWPTNAEAEVRFQFLVFTLWVPPDVWINTK
	A.ochraceus	291 FNVKDPNCIHLWEISIAGSNSLTYQTGDHIAVWPTWVSEVD&FLVFTLWVPPDVWINTK
	S.cerevisiae	280 FSSNDPNCIHLSEFDLSSGSMIRKISTGDMHLAVWPSPLEKLEOF-SIYNLDP--STIFDLP
	A.niger	350 IDVTAKVPIPTETTYDAAVRYKYNECAPVSRQFVATLAAFAIMRKAGQDNQWVPOG-LP
30	A.ochraceus	351 IDVTAKVPIPTETTYDAAVRYKYNECAPVSPQFVSTLAAFAIMDEATPABIVRGSIMYV
	S.cerevisiae	338 LIPFVWVTPETPQIGAAINHHLITGPVSPMLFSS: IQSANNAADVPEKTLISKHIO
	A.niger	409 PREGHCPMLQH-AALOSITE-KPFPSAVPFSSLIELCITFLQFRYTSISSSSLVQHDKISIT
	A.ochraceus	411 HEKITNRCFMIAGALOSITE-KPPTAVPFSSLIELCITFLCPRYYSISSSSLVHDFKISIT
	S.cerevisiae	398 AVELYSKYTFED-PALKYLEDGAKADNVEPQLNFSWPOVTPPYVSISSSNSEJOTMFLV
35	A.niger	468 AVVESVRLI--GASHMVKGVTITNYLLALKKONRSRISRSR-EDLLHHQDPHMYDGIIH
	A.ochraceus	470 AVVESVRLI--GEPHVKGVTTIYLLALBEKQNEPSPDQHG-CTSTGPPPMVYDGIIH
	S.cerevisiae	458 SIVENFPNSELPLAPPVGVS/TTHLRLRNQIQLARIVNVIATNLPHDQNPPLKFLANYKL
40	A.niger	525 PVHVFRHSNFKELESDDPSRFVIIWVGPGTGVAAPFRGFICERAAALAKGE-----VCPVVL
	A.ochraceus	527 PVHVFRHSNFKELPSSDPSRPVIMVGPGTGVAAPFRGFICERAAALAKGE-----VCTILL
	S.cerevisiae	518 PVHVFRSIEFLRPSNPSAPVIIWVSGTGVAAPFRGFICERAAALAKGE-----VCPVVL
45	A.niger	579 FGCRKSDEDFLYKDEWMTYQDQLGDNLKIIITAFSRP-GPQKVTVKPLFEHSELVSDLLP
	A.ochraceus	581 FGCRKSDEDFLYKDEWMTFEDQGDSLKIIITAFSRP-SAEKVVVCHPLFEHSELVSDLLP
	S.cerevisiae	578 VPSNTI-DEWQDF-PEAKKUDGSEFMWVHSELPTKVVV/DKIKDYEDC-FEMIN
	A.niger	638 QKATFYVCSDAAMMAREVNVLVLQOIIIAQGQLPAEKGEENVRHMFRGRYQEDWV
50	A.ochraceus	640 QKATFYVCSDAAMMAREVNVLVLQOIIIAQGQLPAEKGEENVRHMFRGRYQEDWV
	S.cerevisiae	637 NGCFIIVCGDAKCKMKG-STALVGLSLRGSNTTCAEFLIPMLKTSGBQEDWV
	A.ochraceus, PMON45632	(SEQ ID NO: 05)
	A.niger	(SEQ ID NO: 38)
	S.cerevisiae, yeast	(SEQ ID NO: 37)

Figure 9 - Phylogenetic tree showing the relatedness of Aspergillus ochraceus and human oxidoreductase to reductases from A. niger, yeast, and mouse.

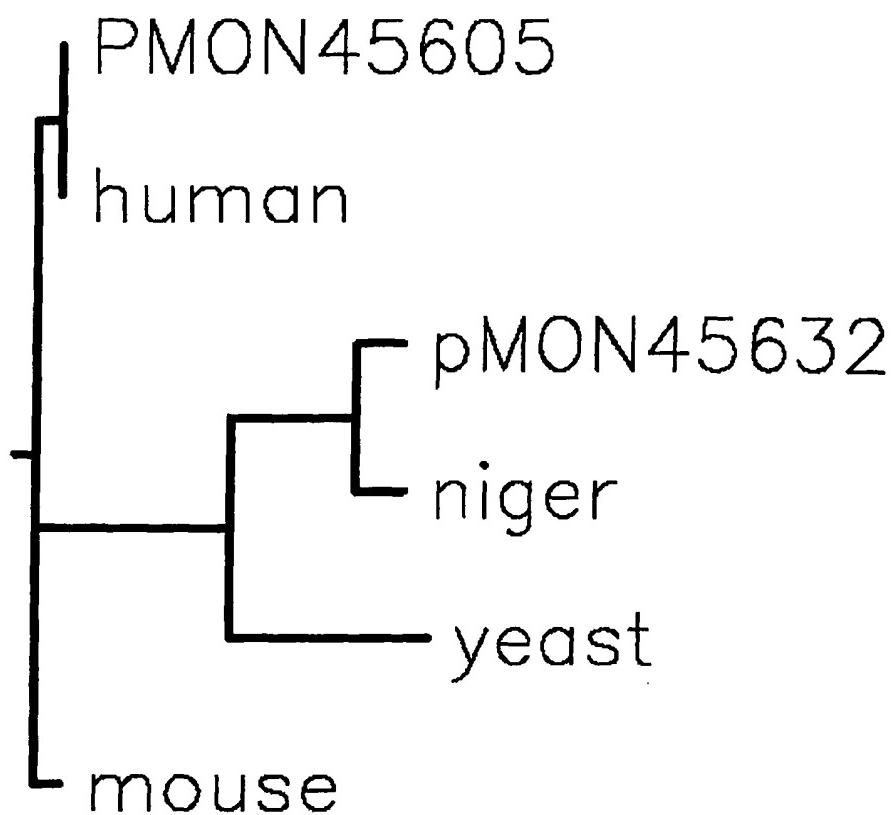


Figure 10 – Percent homology between *Aspergillus ochraceus* oxidoreductase to reductases from *A. niger*, yeast, and mouse and human.

Accession number	organism	% id to A.och oxred
CAA81550	<i>A. niger</i>	84
BAA02936	<i>S. cerevisiae</i>	37
BAA04496	mouse	34
AAB21814	human	33

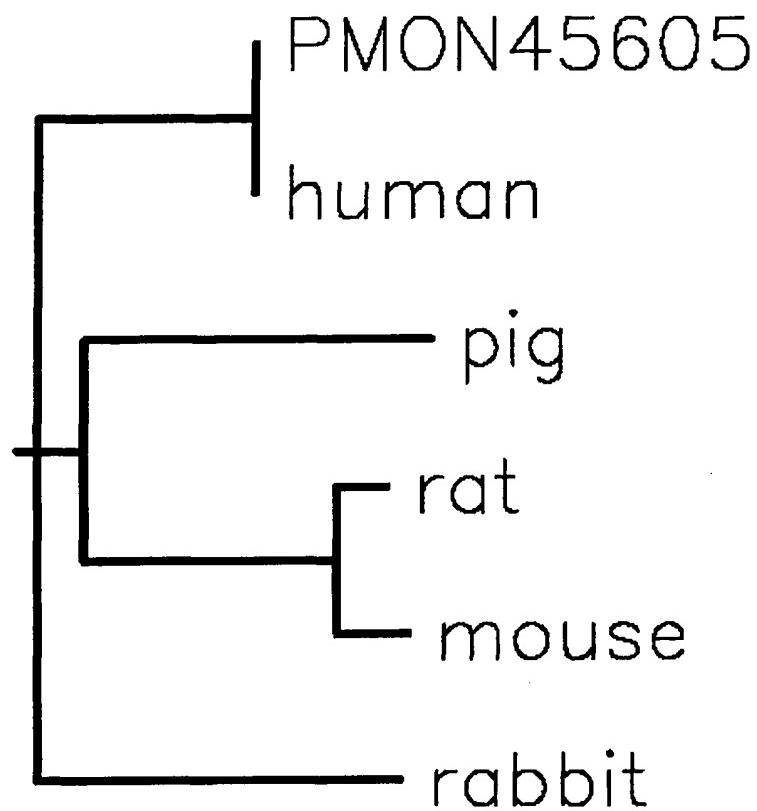
Figure 11 – Amino acid homology alignment of human oxidoreductase with the top 4 hits from SwissProt

	PMON45605	1	AGDSH/DTSTTISRAV/EEVSLFSEITLFLSLIVWNLTCILFPPVFFETPTYIPL
5	human	1	AGDSH/DTSTTISRAV/EEVSLFSEITLFLSLIVWNLTCILFPPVFFETPTYIPL
	rabbit	1	MADSPGDTCAIMPEAPOAFASVPSQTIVLFLSLIVWNLTCILFPPVFFETPTYIPL
	rat	1	MGDSH/DTSATMPEAEEVSLSSTLFLSLIVWNLTCILFPPVFFETPTYIPL
	mouse	1	MGDSH/DTSATMPEAEEVSLSSTLFLSLIVWNLTCILFPPVFFETPTYIPL
	pig	1	MGDSN/DTGTTSEMVAEEVSLFSTLFLSLIVWNLTCILFPPVFFETPTYIPL
10			
	PMON45605	61	TSS-/RESSFEKMTDPRUINWTLA/TAAFPAMKLOEDRHTYIFKQVAFPTI
	human	61	TSS-/RESSFEKMTDPRUINWTS/TAAPAMKLOEDRHTYIFKQVAFPTI
	rabbit	61	TSSE-/RESSFEKMTDPRUINWTS/TAAPAMKLOEDRHTYIFKQVAFPTI
15	rat	61	APP-/RESSFEKMTDPRUINWTS/TAAPAMKLOEDRHTYIFKQVAFPTI
	mouse	61	APP-/RESSFEKMTDPRUINWTS/TAAPAMKLOEDRHTYIFKQVAFPTI
	pig	61	TSE-/WDSSFEKMTDPRUINWTS/TAAPAMKLOEDRHTYIFKQVAFPTI
20			
	PMON45605	120	ADLSSLPEIINALWFCMATYGEGLFTDVKCDFYEWLCEFLWLSWVFAVNEHQLKTF
	human	120	ADLSSLPEIINALWFCMATYGEGLFTDVKCDFYEWLCEFLWLSWVFAVNEHQLKTF
	rabbit	121	ADLSSLPEIINALWFCATYGEGLFTDVKCDFYEWLCEFLWLSWVFAVNEHQLKTF
	rat	120	ADLSSLPEIINALWFCATYGEGLFTDVKCDFYEWLCEFLWLSWVFAVNEHQLKTF
	mouse	120	ADLSSLPEIINALWFCATYGEGLFTDVKCDFYEWLCEFLWLSWVFAVNEHQLKTF
25	pig	120	SDLSSLPEIINALWFCATYGEGLFTDVKCDFYEWLCEFLWLSWVFAVNEHQLKTF
30			
	PMON45605	180	HFIANGKYVDEKRLEVLGACOFIFELGLGDDGDLLEDFITWEEFPTPAVNEHQLKTF
	human	180	HFIANGKYVDEKRLEVLGACOFIFELGLGDDGDLLEDFITWEEFPTPAVNEHQLKTF
	rabbit	181	HFIANGKYVDEKLGLGACOFIFELGLGDDGDLLEDFITWEEFPTPAVNEHQLKTF
	rat	180	HFIANGKYVDEKLGLGACOFIFELGLGDDGDLLEDFITWEEFPTPAVNEHQLKTF
	mouse	180	HFIANGKYVDEKLGLGACOFIFELGLGDDGDLLEDFITWEEFPTPAVNEHQLKTF
	pig	180	HFIANGKYVDEKLGLGACOFIFELGLGDDGDLLEDFITWEEFPTPAVNEHQLKTF
35			
	PMON45605	240	SSIROYELVVTHTIDIAAKVAVMGEMGRPLKSYENOKPPFDKHPFLAAVTTRKLNGTER
	human	240	SSIROYELVVTHTIDIAAKVAVMGEMGRPLKSYENOKPPFDKHPFLAAVTTRKLNGTER
	rabbit	241	SSIROYELVLTHTIDIAAKVAVMGEMGRPLKSYENOKPPFDKHPFLAAVTTRKLNGTER
	rat	240	SSIROYELVVTHTIDIAAKVAVMGEMGRPLKSYENOKPPFDKHPFLAAVTTRKLNGTER
40	mouse	240	SSIROYELVVTHTIDIAAKVAVMGEMGRPLKSYENOKPPFDKHPFLAAVTTRKLNGTER
	pig	240	SSIROYELVVTHTIDIAAKVAVMGEMGRPLKSYENOKPPFDKHPFLAAVTTRKLNGTER
45			
	PMON45605	300	LMHLELDISDSKIRYESGDHVAVYPANDSALVNQLCKILGADLDVMSLNNLDEESNKK
	human	300	LMHLELDISDSKIRYESGDHVAVYPANDSALVNQLCKILGADLDVMSLNNLDEESNKK
	rabbit	301	LMHLELDISDSKIRYESGDHVAVYPANDSALVNQLGKILGADLDVMSLNNLDEESNKK
	rat	300	LMHLELDISDSKIRYESGDHVAVYPANDSALVNQCGIEIGLADLDVMSLNNLDEESNKK
	mouse	300	LMHLELDISDSKIRYESGDHVAVYPANDSALVNQCGIEIGLADLDVMSLNNLDEESNKK
	pig	300	LMHLELDISDSKIRYESGDHVAVYPANDSALVNQLGEILGADLDVMSLNNLDEESNKK
50			
	PMON45605	360	PFPCCPTSYRTALTYYLDTNPPTNVLYELAQYASEPSEQEFLRKMASSSGEGKELYLS
	human	360	PFPCCPTSYRTALTYYLDTNPPTNVLYELAQYASEPSEQEFLRKMASSSGEGKELYLS
	rabbit	361	PFPCCPTSYRTALTYYLDTNPPTNVLYELAQYAADEFNEQEFLRKMASSSGEGKELYLS
55	rat	360	PFPCCPTSYRTALTYYLDTNPPTNVLYELAQYASEPSEQEFLRKMASSSGEGKELYLS

	mouse	360	PFPCCPTTYRTALTYLDITNPPRTNVLYELAQYASEPSEOEHHLHKMASSSGEGKELYLSI
	pig	360	PFPCCPTTYRTALTYLDITNPPRTNVLYELAQYASEPSEOHOLPKMASSSGEGKELYLSI
5	PMON45605	420	VVEARRHILAILQDPSLRPPIDHLCELLPRLOARYTISASSSKVHPMSVHICAVVWEYF
	human	420	VVEARRHILAILQDPSLRPPIDHLCELLPRLOARYTISASSSKVHPMSVHICAVVWEYF
	rabbit	421	VVEARRHILAILDQDPSLRPPIDHLCELLPRLOARYTISASSSKVHPMSVHICAVVWEYF
	rat	420	VVEARRHILAILDQDPSLRPPIDHLCELLPRLOARYTISASSSKVHPMSVHICAVVWEYF
	mouse	420	VVEARRHILAILDQDPSLRPPIDHLCELLPRLOARYTISASSSKVHPMSVHICAVVWEYF
10	pig	420	VVEARRHILAILDQDPSLRPPIDHLCELLPRLOARYTISASSSKVHPMSVHICAVVWEYF
	PMON45605	480	RPAASPRINKEVAGNMLPAKEPAGENLGRALVPNPVPSIFRLPPTAIVVWVQVHATVPAI
	human	480	RPAASPRINKEVAGNMLPAKEPAGENLGRALVPNPVPSIFRLPPTAIVVWVQVHATVPAI
15	rabbit	481	RPAASPRINKEVAGNMLPAKEPAGENLGRALVPNPVPSIFRLPPTAIVVWVQVHATVPAI
	rat	480	AESDPRVYVATVNLPAKEPAGENLGRALVPNPVPSIFRLPPTAIVVWVQVHATVPAI
	mouse	480	AESDPRVYVATVNLPAKEPAGENLGRALVPNPVPSIFRLPPTAIVVWVQVHATVPAI
	pig	480	TESDPRVYVATVNLPAKEPAGENLGRALVPNPVPSIFRLPPTAIVVWVQVHATVPAI

	PMON45605	540	FIGFIQERAVLFQDGKEVGETLLYFGCRSDEDLYPEELAQFHRDGA TQINVAFSPE	
	human	540	FIGFIQERAVLFQDGKEVGETLLYGC CRSDEDLYPEELAQFHRDGATQINVAFSPE	
	rabbit	541	FIGFIQEP AVLFQDGKEVGETLLYGCPFRAADDYLPEELAQFHRDGATQINVAFSPE	
	rat	540	FMGFIQERAVLFQDGKEVGETLLYGC CRSDEDLYPEELAQFHRDGATQINVAFSPE	
5	mouse	540	FMGFIQEP AVLFQDGKEVGETLLYGCRSLEDLYPEELAQFHRDGATQINVAFSPE	
	pig	540	FIGFIQERAVTQDGKEVGETLLYGC RESDEDLYPEELAQFHRDGATQINVAFSPE	
	PMON45605	600	SHKVVVAVHLLPQCREHLWKL -EGGAHITYCQDARIRAEWDWVPPVIVIAELAHEHAY	
10	human	600	SHKVVVAVHLLPQCREHLWKL -EGGAHITYCQDARIRAEWDWVPPVIVIAELAHEHAY	
	rabbit	601	SOKVVVAVHLLPQCREHLWKL RLINEGGAAHITYCQDARIRAEWDWVPPVIVIAELAHEHAY	
	rat	600	SHKVVVAVHLLPQCREHLWKL HEGGAAHITYCQDARIRAEWDWVPPVIVIAELAHEHAY	
	mouse	600	SHKVVVAVHLLPQCREHLWKL HEGGAAHITYCQDARIRAEWDWVPPVIVIAELAHEHAY	
	pig	600	POVTVVAVHLLPFPKREHLYL TDPAHVVTCQDARIRAEWDWVPPVIVIAELAHEHAY	
15	PMON45605	659	VDVYVPLM QGTVSLDWIE (SEQ ID NO: 03)	
	human	659	VDVYVPLM QGTVSLDWIE (SEQ ID NO: 52)	
	rabbit	661	VDVYVPLM QGTVSLDWIE (SEQ ID NO: 53)	
20	rat	660	VDVYVPLM QGTVSLDWIE (SEQ ID NO: 54)	
	mouse	660	VDVYVPLM QGTVSLDWIE (SEQ ID NO: 55)	
	pig	660	VDVYVPLM QGTVSLDWIE (SEQ ID NO: 56)	

Figure 12 – Phylogenetic tree showing the relatedness of human oxidoreductase (P16435) with top 4 hits from SwissProt



**Figure 13 – Percent homology between human oxidoreductase
and top 4 hits from SwissProt**

Accession number	Species	% id to human oxred
P00388	rat	92
P00389	rabbit	92
P37040	mouse	92
P04175	pig	91

Figure 14 - Expression of Aspergillus ochraceus 11 alpha hydroxylase in transfected Sf9 insect cells

Expression of 11- α -Hydroxylase in Transfected Sf9 Cells

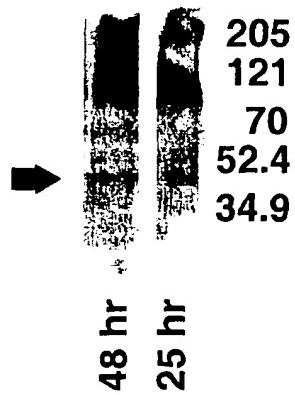


Figure 15 - Expression of Aspergillus ochraceus P450 oxidoreductase in transfected Sf9 insect cells

Expression of Fungal P-450 Oxidoreductase in Transfected Sf9 Cells

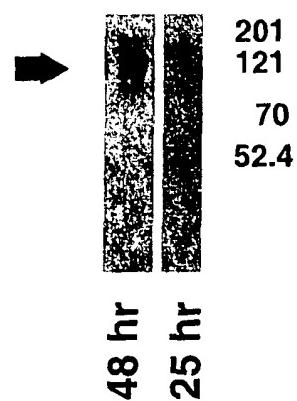
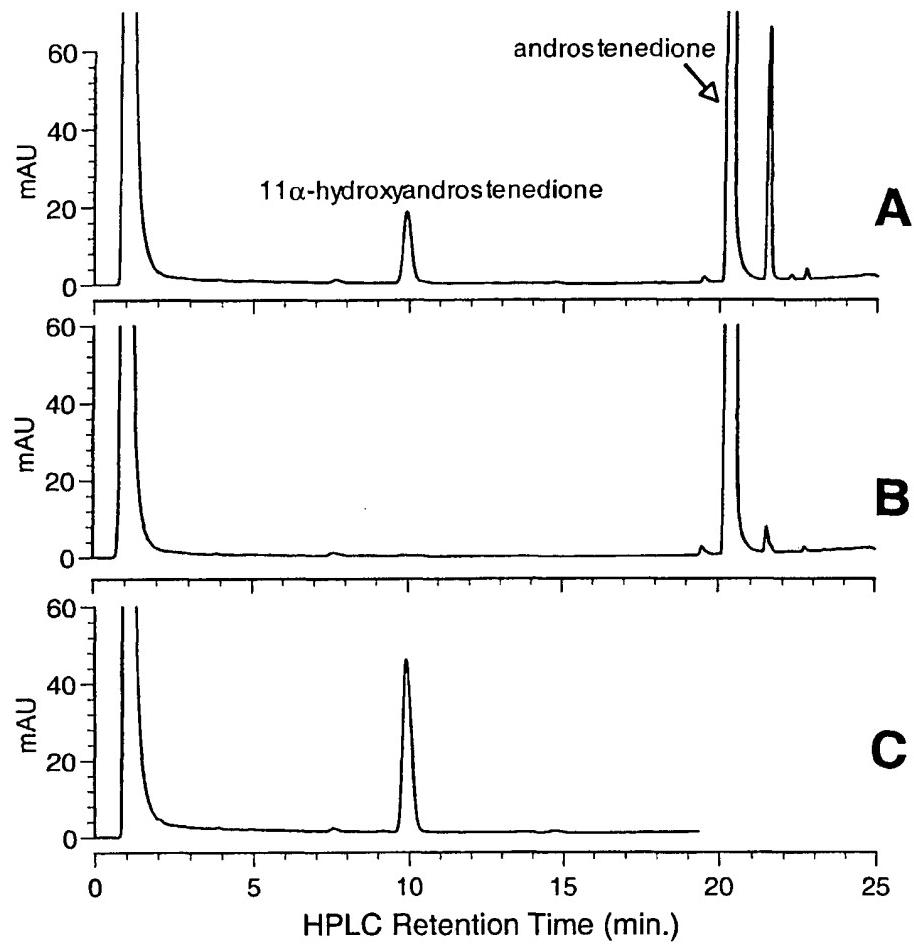


Figure 16 - Conversion of androstenedione to 11 alpha hydroxy androstenedione monitored by HPLC



Sequence Listing

SEQUENCE LISTING

	gac ctt cct gag ctg aat ccc gcc aaa cca ttc gag ttc acc aat cgt	316	
	Asp Leu Pro Glu Leu Asn Pro Ala Lys Pro Phe Glu Phe Thr Asn Arg		
45	45	50	
	55		
5			
	cgt cgt gtt cat gag ttt gtt gaa aat agt aag acg ttg ctt gct cgg	364	
	Arg Arg Val His Glu Phe Val Glu Asn Ser Lys Ser Leu Leu Ala Arg		
	60	65	70
10			
	ggg agg gaa ttg cac ggg cac gag ccg tac aga ctc atg tct gaa tgg	412	
	Gly Arg Glu Leu His Gly His Glu Pro Tyr Arg Leu Met Ser Glu Trp		
	75	80	85
15			
	gga tcc ttg att gtc ctg ccc cca gag tgc gcc gac gag ctg cgc aac	460	
	Gly Ser Leu Ile Val Leu Pro Pro Glu Cys Ala Asp Glu Leu Arg Asn		
	90	95	100
	105		
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	gac cca aga atg gac ttt gag acg ccc acc acc gac gac tcc cac gga	508	
	Asp Pro Arg Met Asp Phe Glu Thr Pro Thr Thr Asp Asp Ser His Gly		
	110	115	120
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	tat atc cct ggc ttc gac gct ctc aac gca gac ccg aac ctg act aaa	556	
	Tyr Ile Pro Gly Phe Asp Ala Leu Asn Ala Asp Pro Asn Leu Thr Lys		
	125	130	135
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	gtg gtc acc aag tac ctc aca aaa gca ttg aac aag ctt act gct ccg	604	
	Val Val Thr Lys Tyr Leu Thr Lys Ala Leu Asn Lys Leu Thr Ala Pro		
	140	145	150
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	atc tcg cat gaa gcg tcc atc gcc atg aaa gcg gtg ctg ggt gac gat	652	
	Ile Ser His Glu Ala Ser Ile Ala Met Lys Ala Val Leu Gly Asp Asp		
	155	160	165
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	cca gat tgg cgt gag atc tac cca gcc aga gac ttg ctc cag ctc gtc	700	
	Pro Asp Trp Arg Glu Ile Tyr Pro Ala Arg Asp Leu Leu Gln Leu Val		
	170	175	180
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	gcc cgg atg tcg aca aga gtg ttc ctt ggc gag gaa atg tgc aat aac	748	
	Ala Arg Met Ser Thr Arg Val Phe Leu Gly Glu Met Cys Asn Asn		
	190	195	200
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	cag gat tgg atc caa acc tca tca caa tac gcg gcc ctt gcc ttc ggt	796	
	Gln Asp Trp Ile Gln Thr Ser Ser Gln Tyr Ala Ala Leu Ala Phe Gly		
	205	210	215
55			
	gtc ggt gac aag ctt aga ata tac ccg aga atg atc aga ccg ata gta	844	
	Val Gly Asp Lys Leu Arg Ile Tyr Pro Arg Met Ile Arg Pro Ile Val		
	220	225	230
60			
	cat tgg ttc atg cca tcc tgt tgg gag ctg cgc cga tcg ctg cga cgc	892	
	His Trp Phe Met Pro Ser Cys Trp Glu Leu Arg Arg Ser Leu Arg Arg		
	235	240	245
65			
	tgc cga cag att ctc acg ccg tac att cac aaa cgc aag tcc ctg aag	940	
	Cys Arg Gln Ile Leu Thr Pro Tyr Ile His Lys Arg Lys Ser Leu Lys		
	250	255	260
	265		

	ggg acc acg gac gag cag ggc aag ccc ctt atg ttt gat gat tcc atc Gly Thr Thr Asp Glu Gln Gly Lys Pro Leu Met Phe Asp Asp Ser Ile	270	275	280	988
5	gag tgg ttc gag cga gag ctg ggt ccc aac cac gac gcg gtc ctg aag Glu Trp Phe Glu Arg Glu Leu Gly Pro Asn His Asp Ala Val Leu Lys	285	290	295	1036
10	cag gtc acg ctc tcc ata gtt gct atc cac acc acg agt gac cta ctc Gln Val Thr Leu Ser Ile Val Ala Ile His Thr Thr Ser Asp Leu Leu	300	305	310	1084
15	ttg cag gcc atg agc gat ctc gcg cag aac ccg aaa gtg cta caa gca Leu Gln Ala Met Ser Asp Leu Ala Gln Asn Pro Lys Val Leu Gln Ala	315	320	325	1132
20	gtg cgc gag gag gtg gtc cga gtg ctg agc acc gag ggg ctc agc aag Val Arg Glu Glu Val Val Arg Val Leu Ser Thr Glu Gly Leu Ser Lys	330	335	340	1180
25	gtc tcg ctt cac agt ctc aag ctc atg gac agc gcg ttg aag gaa agc Val Ser Leu His Ser Leu Lys Leu Met Asp Ser Ala Leu Lys Glu Ser	350	355	360	1228
30	cag cgt ctc agg cct acg ctt ctc ggc tcc ttt cgt cgg cag gca acg Gln Arg Leu Arg Pro Thr Leu Leu Gly Ser Phe Arg Arg Gln Ala Thr	365	370	375	1276
35	aat gac atc aag ctg aag agc ggg ttt gtc ata aag aaa ggg act aga Asn Asp Ile Lys Leu Lys Ser Gly Phe Val Ile Lys Lys Gly Thr Arg	380	385	390	1324
40	gtc gtg atc gac agc acc cat atg tgg aat ccc gag tat tac act gac Val Val Ile Asp Ser Thr His Met Trp Asn Pro Glu Tyr Tyr Thr Asp	395	400	405	1372
45	cct ctc cag tac gac ggg tac cgc tac ttc aac aag cgg cag aca ccc Pro Leu Gln Tyr Asp Gly Tyr Arg Tyr Phe Asn Lys Arg Gln Thr Pro	410	415	420	1420
50	ggc gag gac aag aac gcg ttg ctc gtc agc aca agc gcc aac cac atg Gly Glu Asp Lys Asn Ala Leu Leu Val Ser Thr Ser Ala Asn His Met	430	435	440	1468
55	gga ttc ggt cac ggc gtt cac gcc tgt cct ggc aga ttc ttc gcc tcc Gly Phe Gly His Gly Val His Ala Cys Pro Gly Arg Phe Phe Ala Ser	445	450	455	1516
60	aac gag atc aag att gcc ttg tgt cat atc atc tta aat tat gag tgg Asn Glu Ile Lys Ile Ala Leu Cys His Ile Ile Leu Asn Tyr Glu Trp	460	465	470	1564
65	cgt ctt cca gac ggc ttc aag ccc cag cct ctc aac atc ggg atg act Arg Leu Pro Asp Gly Phe Lys Pro Gln Pro Leu Asn Ile Gly Met Thr	475	480	485	1612
70	tat ctg gcg gat ccc aat acc agg atg ctg atc agg cca cgc aag gcg				1660

Tyr Leu Ala Asp Pro Asn Thr Arg Met Leu Ile Arg Pro Arg Lys Ala
 490 495 500 505
 gag atc gat atg gcg agt tta act gtg tag gtgcAACACg aagtccTgat 1710
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 Ala Lys Pro Phe Glu Phe Thr Asn Arg Arg Arg Val His Glu Phe Val
 50 55 60
 25 Glu Asn Ser Lys Ser Leu Leu Ala Arg Gly Arg Glu Leu His Gly His
 65 70 75 80
 Glu Pro Tyr Arg Leu Met Ser Glu Trp Gly Ser Leu Ile Val Leu Pro
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 Pro Glu Cys Ala Asp Glu Leu Arg Asn Asp Pro Arg Met Asp Phe Glu
 30 100 105 110
 Thr Pro Thr Thr Asp Asp Ser His Gly Tyr Ile Pro Gly Phe Asp Ala
 115 120 125
 Leu Asn Ala Asp Pro Asn Leu Thr Lys Val Val Thr Lys Tyr Leu Thr
 130 135 140
 35 Lys Ala Leu Asn Lys Leu Thr Ala Pro Ile Ser His Glu Ala Ser Ile
 145 150 155 160
 Ala Met Lys Ala Val Leu Gly Asp Asp Pro Asp Trp Arg Glu Ile Tyr
 165 170 175
 Pro Ala Arg Asp Leu Leu Gln Leu Val Ala Arg Met Ser Thr Arg Val
 40 180 185 190
 Phe Leu Gly Glu Glu Met Cys Asn Asn Gln Asp Trp Ile Gln Thr Ser
 195 200 205
 Ser Gln Tyr Ala Ala Leu Ala Phe Gly Val Gly Asp Lys Leu Arg Ile
 210 215 220
 45 Tyr Pro Arg Met Ile Arg Pro Ile Val His Trp Phe Met Pro Ser Cys
 225 230 235 240
 Trp Glu Leu Arg Arg Ser Leu Arg Arg Cys Arg Gln Ile Leu Thr Pro
 245 250 255
 Tyr Ile His Lys Arg Lys Ser Leu Lys Gly Thr Thr Asp Glu Gln Gly
 50 260 265 270
 Lys Pro Leu Met Phe Asp Asp Ser Ile Glu Trp Phe Glu Arg Glu Leu
 275 280 285
 Gly Pro Asn His Asp Ala Val Leu Lys Gln Val Thr Leu Ser Ile Val
 290 295 300
 55 Ala Ile His Thr Thr Ser Asp Leu Leu Gln Ala Met Ser Asp Leu
 305 310 315 320
 Ala Gln Asn Pro Lys Val Leu Gln Ala Val Arg Glu Val Val Arg

	325	330	335	
5	Val Leu Ser Thr Glu Gly Leu Ser Lys Val Ser Leu His Ser Leu Lys 340	345	350	
	Leu Met Asp Ser Ala Leu Lys Glu Ser Gln Arg Leu Arg Pro Thr Leu 355	360	365	
	Leu Gly Ser Phe Arg Arg Gln Ala Thr Asn Asp Ile Lys Leu Lys Ser 370	375	380	
	Gly Phe Val Ile Lys Lys Gly Thr Arg Val Val Ile Asp Ser Thr His 385	390	395	
10	Met Trp Asn Pro Glu Tyr Tyr Thr Asp Pro Leu Gln Tyr Asp Gly Tyr 405	410	415	
	Arg Tyr Phe Asn Lys Arg Gln Thr Pro Gly Glu Asp Lys Asn Ala Leu 420	425	430	
	Leu Val Ser Thr Ser Ala Asn His Met Gly Phe Gly His Gly Val His 435	440	445	
15	Ala Cys Pro Gly Arg Phe Phe Ala Ser Asn Glu Ile Lys Ile Ala Leu 450	455	460	
	Cys His Ile Ile Leu Asn Tyr Glu Trp Arg Leu Pro Asp Gly Phe Lys 465	470	475	
20	Pro Gln Pro Leu Asn Ile Gly Met Thr Tyr Leu Ala Asp Pro Asn Thr 485	490	495	
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	gcc gaa gaa gta tct ctt ttc agc atg acg gac atg att ctg ttt tcg			
	Ala Glu Glu Val Ser Leu Phe Ser Met Thr Asp Met Ile Leu Phe Ser			
	20	25	30	
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	Leu Ile Val Gly Leu Leu Thr Tyr Trp Phe Leu Phe Arg Lys Lys Lys			
	35	40	45	
50	gaa gaa gtc ccc gag ttc acc aaa att cag aca ttg acc tcc tct gtc			
	Glu Glu Val Pro Glu Phe Thr Lys Ile Gln Thr Leu Thr Ser Ser Val			
	50	55	60	
55	aga gag agc agc ttt gtg gaa aag atg aag aaa acg ggg agg aac atc			
	Arg Glu Ser Ser Phe Val Glu Lys Met Lys Lys Thr Gly Arg Asn Ile			
	65	70	75	80

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10	gac cct gag gag tat gac ctg gcc gac ctg agc agc ctg cca gag atc Asp Pro Glu Glu Tyr Asp Leu Ala Asp Leu Ser Ser Leu Pro Glu Ile 115 120 125	384
15	gac aac gcc ctg gtt ttc tgc atg gcc acc tac ggt gag gga gac Asp Asn Ala Leu Val Val Phe Cys Met Ala Thr Tyr Gly Glu Gly Asp 130 135 140	432
20	ccc acc gac aat gcc cag gac ttc tac gac tgg ctg cag gag aca gac Pro Thr Asp Asn Ala Gln Asp Phe Tyr Asp Trp Leu Gln Glu Thr Asp 145 150 155 160	480
25	gtg gat ctc tct ggg gtc aag ttc gcg gtg ttt ggt ctt ggg aac aag Val Asp Leu Ser Gly Val Lys Phe Ala Val Phe Gly Leu Gly Asn Lys 165 170 175	528
30	acc tac gag cac ttc aat gcc atg ggc aag tac gtg gac aag cgg ctg Thr Tyr Glu His Phe Asn Ala Met Gly Lys Tyr Val Asp Lys Arg Leu 180 185 190	576
35	gag cag ctc ggc gcc cag cgc atc ttt gag ctg ggg ttg ggc gac gac Glu Gln Leu Gly Ala Gln Arg Ile Phe Glu Leu Gly Leu Gly Asp Asp 195 200 205	624
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45	ccg gcc gtg tgt gaa cac ttt ggg gtg gaa gcc act ggc gag gag tcc Pro Ala Val Cys Glu His Phe Gly Val Glu Ala Thr Gly Glu Glu Ser 225 230 235 240	720
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55	aag gtg tac atg ggg gag atg ggc cgg ctg aag agc tac gag aac cag Lys Val Tyr Met Gly Glu Met Gly Arg Leu Lys Ser Tyr Glu Asn Gln 260 265 270	816
	aag ccc ccc ttt gat gcc aag aat ccg ttc ctg gct gca gtc acc acc Lys Pro Pro Phe Asp Ala Lys Asn Pro Phe Leu Ala Ala Val Thr Thr 275 280 285	864
	aac cgg aag ctg aac cag gga acc gag cgc cac ctc atg cac ctg gaa Asn Arg Lys Leu Asn Gln Gly Thr Glu Arg His Leu Met His Leu Glu 290 295 300	912
	ttg gac atc tcg gac tcc aaa atc agg tat gaa tct ggg gac cac gtg	960

	Leu Asp Ile Ser Asp Ser Lys Ile Arg Tyr Glu Ser Gly Asp His Val		
305	310	315	320
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	325	330	335
10	atc ctg ggt gcc gac ctg gac gtc gtc atg tcc ctg aac aac ctg gat Ile Leu Gly Ala Asp Leu Asp Val Val Met Ser Leu Asn Asn Leu Asp		1056
	340	345	350
15	gag gag tcc aac aag aag cac cca ttc ccg tgc cct acg tcc tac cgc Glu Glu Ser Asn Lys Lys His Pro Phe Pro Cys Pro Thr Ser Tyr Arg		1104
	355	360	365
	acg gcc ctc acc tac tac ctg gac atc acc aac ccg ccg cgt acc aac Thr Ala Leu Thr Tyr Tyr Leu Asp Ile Thr Asn Pro Pro Arg Thr Asn		1152
	370	375	380
20	gtg ctg tac gag ctg gcg cag tac gcc tcg gag ccc tcg gag cag gag Val Leu Tyr Glu Leu Ala Gln Tyr Ala Ser Glu Pro Ser Glu Gln Glu		1200
	385	390	395
25	ctg ctg cgc aag atg gcc tcc tcc ggc gag ggc aag gag ctg tac Leu Leu Arg Lys Met Ala Ser Ser Ser Gly Glu Gly Lys Glu Leu Tyr		1248
	405	410	415
30	ctg agc tgg gtg gag gcc ccg agg cac atc ctg gcc atc ctg cag Leu Ser Trp Val Val Glu Ala Arg Arg His Ile Leu Ala Ile Leu Gln		1296
	420	425	430
35	gac tgc ccg tcc ctg ccg ccc atc gac cac ctg tgt gag ctg ctg Asp Cys Pro Ser Leu Arg Pro Pro Ile Asp His Leu Cys Glu Leu Leu		1344
	435	440	445
	ccg cgc ctg cag gcc cgc tac tac tcc atc gcc tca tcc tcc aag gtc Pro Arg Leu Gln Ala Arg Tyr Tyr Ser Ile Ala Ser Ser Ser Lys Val		1392
	450	455	460
40	cac ccc aac tct gtg cac atc tgt gcg gtg gtt gtg gag tac gag acc His Pro Asn Ser Val His Ile Cys Ala Val Val Val Glu Tyr Glu Thr		1440
	465	470	475
45	aag gcc ggc cgc atc aac aag ggc gtg gcc acc aac tgg ctg ccg gcc Lys Ala Gly Arg Ile Asn Lys Gly Val Ala Thr Asn Trp Leu Arg Ala		1488
	485	490	495
50	aag gag cct gcc ggg gag aac ggc ggc cgt gcg ctg gtg ccc atg ttc Lys Glu Pro Ala Gly Glu Asn Gly Gly Arg Ala Leu Val Pro Met Phe		1536
	500	505	510
55	gtg cgc aag tcc cag ttc cgc ctg ccc ttc aag gcc acc acg cct gtc Val Arg Lys Ser Gln Phe Arg Leu Pro Phe Lys Ala Thr Thr Pro Val		1584
	515	520	525
	atc atg gtg ggc ccc ggc acc ggg gtg gca ccc ttc ata ggc ttc atc Ile Met Val Gly Pro Gly Thr Gly Val Ala Pro Phe Ile Gly Phe Ile		1632

	530	535	540	
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5	545	550	555	560
	ctg ctg tac tac ggc tgc cgc cgc tcg gat gag gac tac ctg tac cgg Leu Leu Tyr Tyr Gly Cys Arg Arg Ser Asp Glu Asp Tyr Leu Tyr Arg			1728
	565	570	575	
10	gag gag ctg gcg cag ttc cac agg gac ggt gcg ctc acc cag ctc aac Glu Glu Leu Ala Gln Phe His Arg Asp Gly Ala Leu Thr Gln Leu Asn			1776
	580	585	590	
15	gtg gcc ttc tcc cgg gag cag tcc cac aag gtc tac gtc cag cac ctg Val Ala Phe Ser Arg Glu Gln Ser His Lys Val Tyr Val Gln His Leu			1824
	595	600	605	
20	cta aag caa gac cga gag cac ctg tgg aag ttg atc gaa ggc ggt gcc Leu Lys Gln Asp Arg Glu His Leu Trp Lys Leu Ile Glu Gly Gly Ala			1872
	610	615	620	
25	cac atc tac gtc tgt ggg gat gca cgg aac atg gcc agg gat gtg cag His Ile Tyr Val Cys Gly Asp Ala Arg Asn Met Ala Arg Asp Val Gln			1920
	625	630	635	640
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	645	650	655	
30	cag gcg gtg gac tac atc aag aaa ctg atg acc aag ggc cgc tac tcc Gln Ala Val Asp Tyr Ile Lys Lys Leu Met Thr Lys Gly Arg Tyr Ser			2016
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	Ile Val Phe Tyr Gly Ser Gln Thr Gly Thr Ala Glu Glu Phe Ala Asn 85 90 95			

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 Val Asp Leu Ser Gly Val Lys Phe Ala Val Phe Gly Leu Gly Asn Lys
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 195 200 205
 15 Asp Gly Asn Leu Glu Glu Asp Phe Ile Thr Trp Arg Glu Gln Phe Trp
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 225 230 235 240
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 260 265 270
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 275 280 285
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 Glu Glu Ser Asn Lys Lys His Pro Phe Pro Cys Pro Thr Ser Tyr Arg
 355 360 365
 35 Thr Ala Leu Thr Tyr Tyr Leu Asp Ile Thr Asn Pro Pro Arg Thr Asn
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 Val Leu Tyr Glu Leu Ala Gln Tyr Ala Ser Glu Pro Ser Glu Gln Glu
 385 390 395 400
 Leu Leu Arg Lys Met Ala Ser Ser Ser Gly Glu Gly Lys Glu Leu Tyr
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	Glu Glu Leu Ala Gln Phe His Arg Asp Gly Ala Leu Thr Gln Leu Asn			
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	Val Ala Phe Ser Arg Glu Gln Ser His Lys Val Tyr Val Gln His Leu			
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	Leu Lys Gln Asp Arg Glu His Leu Trp Lys Leu Ile Glu Gly Gly Ala			
	610	615	620	
10	His Ile Tyr Val Cys Gly Asp Ala Arg Asn Met Ala Arg Asp Val Gln			
	625	630	635	640
	Asn Thr Phe Tyr Asp Ile Val Ala Glu Leu Gly Ala Met Glu His Ala			
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	20	25	30	
45	cct tat gcc tcg gct ggt ccg gcg atg aat gga ggc gcc aag gcc ggc 382 Pro Tyr Ala Ser Ala Gly Pro Ala Met Asn Gly Gly Ala Lys Ala Gly			
	35	40	45	50
50	aag act cgc gac att gtt cag aaa atg gac gaa act ggc aaa aac tgt 430 Lys Thr Arg Asp Ile Val Gln Lys Met Asp Glu Thr Gly Lys Asn Cys			
	55	60	65	
55	gtg att ttc tac ggc tcg caa acc ggt acc gct gag gac tac gcg tcc 478 Val Ile Phe Tyr Gly Ser Gln Thr Gly Thr Ala Glu Asp Tyr Ala Ser			
	70	75	80	
	aga ctg gcc aag gaa ggc tcc cag cga ttc ggt ctc aag acc atg gtg 526			

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	85	90	95	
5	gcc gat ctg gag gac tac gac tac gaa aac ctg gaa aag ttc ccc gag Ala Asp Leu Glu Asp Tyr Asp Tyr Glu Asn Leu Glu Lys Phe Pro Glu		574	
	100	105	110	
10	gac aag gtt gtt ttc ttc gtt ctg gcc act tat ggc gag ggt gaa ccc Asp Lys Val Val Phe Phe Val Leu Ala Thr Tyr Gly Glu Gly Glu Pro		622	
	115	120	125	130
15	acg gat aat gcg gtt gaa ttc tac cag ttc gtc acg ggc gaa gat gct Thr Asp Asn Ala Val Glu Phe Tyr Gln Phe Val Thr Gly Glu Asp Ala		670	
	135	140	145	
	gct ttc gag agc ggc gct acc gcc gac gat aag cct ctg tct tct ctc Ala Phe Glu Ser Gly Ala Thr Ala Asp Asp Lys Pro Leu Ser Ser Leu		718	
20	150	155	160	
	aag tat gtc acg ttt ggt ctg ggt aac aac acc tat gag cac tac aac Lys Tyr Val Thr Phe Gly Leu Gly Asn Asn Thr Tyr Glu His Tyr Asn		766	
	165	170	175	
25	gct atg gtt cgc aat gtg gac gcc gct ctc aca aag ttc ggc gcc caa Ala Met Val Arg Asn Val Asp Ala Ala Leu Thr Lys Phe Gly Ala Gln		814	
	180	185	190	
30	cgc att ggc tct gct ggt gag ggt gac gac ggc gct ggt aca atg gaa Arg Ile Gly Ser Ala Gly Glu Gly Asp Asp Gly Ala Gly Thr Met Glu		862	
	195	200	205	210
	gag gat ttc ctg gcc tgg aag gaa ccc atg tgg gct gcc ctt tct gag Glu Asp Phe Leu Ala Trp Lys Glu Pro Met Trp Ala Ala Leu Ser Glu		910	
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	gcg atg aac ctg caa gag cgc gat gcg gtc tac gag ccg gtc ttc aat Ala Met Asn Leu Gln Glu Arg Asp Ala Val Tyr Glu Pro Val Phe Asn		958	
	230	235	240	
40	gtc acc gag gag tcc ctg agc ccc gaa gat gag aac gtt tac ctc Val Thr Glu Asp Glu Ser Leu Ser Pro Glu Asp Glu Asn Val Tyr Leu		1006	
	245	250	255	
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	260	265	270	
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	340	345	350	
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	355	360	365	370
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	ttc aac atc gct gct ctc cag agc atc acg tcc aag cct ttc acc Phe Asn Ile Ala Gln Ala Leu Gln Ser Ile Thr Ser Lys Pro Phe Thr			
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40	agc att acc gcc gtt gtg gag tcg gtt cgc ttg cct ggt gag gaa cac Ser Ile Thr Ala Val Val Glu Ser Val Arg Leu Pro Gly Glu Glu His			
	470	475	480	
				1726
45	att gtc aag ggt gtg acc acg aac tat ctt ctc gcg ctc aag gaa aag Ile Val Lys Gly Val Thr Thr Asn Tyr Leu Leu Ala Leu Lys Glu Lys			
	485	490	495	
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	Tyr Leu Gly Glu Pro Thr Gln Gly His Leu Gln Gly Glu Pro Lys Gly			
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	Pro Tyr Ser Ala His Asn Pro Phe Ile Ala Pro Ile Ser Glu Ser Arg			
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	290	295	300	
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	Trp Pro Thr Asn Ala Gly Ser Glu Val Asp Arg Phe Leu Gln Ala Phe			
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	Gly Leu Glu Gly Lys Arg His Ser Val Ile Asn Ile Lys Gly Ile Asp			
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	Val Thr Ala Lys Val Pro Ile Pro Thr Pro Thr Thr Tyr Asp Ala Ala			
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	Val Arg Tyr Tyr Leu Glu Val Cys Ala Pro Val Ser Arg Gln Phe Val			
	370	375	380	
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	Val Arg Leu Gly Gly Asp Lys Asp Tyr Phe His Glu Lys Ile Thr Asn			
	405	410	415	
	Arg Cys Phe Asn Ile Ala Gln Ala Leu Gln Ser Ile Thr Ser Lys Pro			
45	420	425	430	
	Phe Thr Ala Val Pro Phe Ser Leu Leu Ile Glu Gly Ile Thr Lys Leu			
	435	440	445	
	Gln Pro Arg Tyr Tyr Ser Ile Ser Ser Ser Leu Val Gln Lys Asp			
	450	455	460	
50	Lys Ile Ser Ile Thr Ala Val Val Glu Ser Val Arg Leu Pro Gly Glu			
	465	470	475	480
	Glu His Ile Val Lys Gly Val Thr Thr Asn Tyr Leu Leu Ala Leu Lys			
	485	490	495	
	Glu Lys Gln Asn Gly Glu Pro Ser Pro Asp Pro His Gly Leu Thr Tyr			
55	500	505	510	
	Ser Ile Thr Gly Pro Arg Asn Lys Tyr Asp Gly Ile His Val Pro Val			
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	Arg Val Leu Gln Asp Ile Ile Asp Glu His Lys Asn Arg Asn Arg Ser		
	260	265	270
25	Ser Glu Glu Arg Glu Ala Val Glu Asp Leu Val Asp Val Leu Leu Lys		
	275	280	285
	Phe Gln Lys Glu Ser Glu Phe Arg Leu Thr Asp Asp Asn Ile Lys Ala		
	290	295	300
	Val Ile Gln Asp Ile Phe Ile Gly Gly Glu Thr Ser Ser Ser Val		
30	305	310	315
	Val Glu Trp Gly Met Ser Glu Leu Ile Arg Asn Pro Arg Val Met Glu		
	325	330	335
	Glu Ala Gln Ala Glu Val Arg Arg Val Tyr Asp Ser Lys Gly Tyr Val		
	340	345	350
35	Asp Glu Thr Glu Leu His Gln Leu Ile Tyr Leu Lys Ser Ile Ile Lys		
	355	360	365
	Glu Thr Met Arg Leu His Pro Pro Val Pro Leu Leu Val Pro Arg Val		
	370	375	380
	Ser Arg Glu Arg Cys Gln Ile Asn Gly Tyr Glu Ile Pro Ser Lys Thr		
40	385	390	395
	Arg Ile Ile Ile Asn Ala Trp Ala Ile Gly Arg Asn Pro Lys Tyr Trp		
	405	410	415
	Gly Glu Thr Glu Ser Phe Lys Pro Glu Arg Phe Leu Asn Ser Ser Ile		
	420	425	430
45	Asp Phe Arg Gly Thr Asp Phe Glu Phe Ile Pro Phe Gly Ala Gly Arg		
	435	440	445
	Arg Ile Cys Pro Gly Ile Thr Phe Ala Ile Pro Asn Ile Glu Leu Pro		
	450	455	460
	Leu Ala Gln Leu Leu Tyr His Phe Asp Trp Lys Leu Pro Asn Lys Met		
50	465	470	475
	Lys Asn Glu Glu Leu Asp Met Thr Glu Ser Asn Gly Ile Thr Leu Arg		
	485	490	495
	Arg Gln Asn Asp Leu Cys Leu Ile Pro Ile Thr Arg Leu Pro		
	500	505	510
55	<210> 31		
	<211> 524		

<212> PRT
<213> Gibberella fujikuroi CAA75566

<400> 31

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	Pro Phe Tyr Ile Ala Ile Phe Val Phe Thr Leu Val Pro Trp Ala Ile			
	20	25	30	
10	Arg Phe Ser Trp Leu Glu Leu Arg Lys Gly Ser Val Val Pro Leu Ala			
	35	40	45	
	Asn Pro Pro Asp Ser Leu Phe Gly Thr Gly Lys Thr Arg Arg Ser Phe			
	50	55	60	
	Val Lys Leu Ser Arg Glu Ile Leu Ala Lys Ala Arg Ser Leu Phe Pro			
	65	70	75	80
15	Asn Glu Pro Phe Arg Leu Ile Thr Asp Trp Gly Glu Val Leu Ile Leu			
	85	90	95	
	Pro Pro Asp Phe Ala Asp Glu Ile Arg Asn Asp Pro Arg Leu Ser Phe			
	100	105	110	
	Ser Lys Ala Ala Met Gln Asp Asn His Ala Gly Ile Pro Gly Phe Glu			
20	115	120	125	
	Thr Val Ala Leu Val Gly Arg Glu Asp Gln Leu Ile Gln Lys Val Ala			
	130	135	140	
	Arg Lys Gln Leu Thr Lys His Leu Ser Ala Val Ile Glu Pro Leu Ser			
	145	150	155	160
25	Arg Glu Ser Thr Leu Ala Val Ser Leu Asn Phe Gly Glu Thr Thr Glu			
	165	170	175	
	Trp Arg Ala Ile Arg Leu Lys Pro Ala Ile Leu Asp Ile Ile Ala Arg			
	180	185	190	
30	Ile Ser Ser Arg Ile Tyr Leu Gly Asp Gln Leu Cys Arg Asn Glu Ala			
	195	200	205	
	Trp Leu Lys Ile Thr Lys Thr Tyr Thr Asn Phe Tyr Thr Ala Ser			
	210	215	220	
	Thr Asn Leu Arg Met Phe Pro Arg Ser Ile Arg Pro Leu Ala His Trp			
	225	230	235	240
35	Phe Leu Pro Glu Cys Arg Lys Leu Arg Gln Glu Arg Lys Asp Ala Ile			
	245	250	255	
	Gly Ile Ile Thr Pro Leu Ile Glu Arg Arg Glu Leu Arg Arg Ala			
	260	265	270	
	Ala Ile Ala Ala Gly Gln Pro Leu Pro Val Phe His Asp Ala Ile Asp			
40	275	280	285	
	Trp Ser Glu Gln Glu Ala Glu Ala Ala Gly Thr Gly Ala Ser Phe Asp			
	290	295	300	
	Pro Val Ile Phe Gln Leu Thr Leu Ser Leu Leu Ala Ile His Thr Thr			
	305	310	315	320
45	Tyr Asp Leu Leu Gln Gln Thr Met Ile Asp Leu Gly Arg His Pro Glu			
	325	330	335	
	Tyr Ile Glu Pro Leu Arg Gln Glu Val Val Gln Leu Leu Arg Glu Glu			
	340	345	350	
	Gly Trp Lys Lys Thr Thr Leu Phe Lys Met Lys Leu Leu Asp Ser Ala			
50	355	360	365	
	Ile Lys Glu Ser Gln Arg Met Lys Pro Gly Ser Ile Val Thr Met Arg			
	370	375	380	
	Arg Tyr Val Thr Glu Asp Ile Thr Leu Ser Ser Gly Leu Thr Leu Lys			
	385	390	395	400
55	Lys Gly Thr Arg Leu Asn Val Asp Asn Arg Arg Leu Asp Asp Pro Lys			
	405	410	415	
	Ile Tyr Asp Asn Pro Glu Val Tyr Asn Pro Tyr Arg Phe Tyr Asp Met			

	420	425	430
	Arg Ser Glu Ala Gly Lys Asp His Gly Ala Gln Leu Val Ser Thr Gly		
	435	440	445
	Ser Asn His Met Gly Phe Gly His Gly Gln His Ser Cys Pro Gly Arg		
5	450	455	460
	Phe Phe Ala Ala Asn Glu Ile Lys Val Ala Leu Cys His Ile Leu Val		
	465	470	475
	Lys Tyr Asp Trp Lys Leu Cys Pro Asp Thr Glu Thr Lys Pro Asp Thr		
	485	490	495
10	Arg Gly Met Ile Ala Lys Ser Ser Pro Val Thr Asp Ile Leu Ile Lys		
	500	505	510
	Arg Arg Glu Ser Val Glu Leu Asp Leu Glu Ala Ile		
	515	520	
15	<210> 32		
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	Trp Asn Asp Thr Gln Gln His Gly Ser Trp Phe Ala Pro Leu Val Thr		
	20	25	30
25	Thr Ser Ala Gly Leu Leu Cys Leu Leu Tyr Leu Cys Ser Ser Gly		
	35	40	45
	Arg Arg Ser Asp Leu Pro Val Phe Asn Pro Lys Thr Trp Trp Glu Leu		
	50	55	60
	Thr Thr Met Arg Ala Lys Arg Asp Phe Asp Ala Asn Ala Pro Ser Trp		
30	65	70	75
	Ile Glu Ser Trp Phe Ser Gln Asn Asp Lys Pro Ile Arg Phe Ile Val		
	85	90	95
	Asp Ser Gly Tyr Cys Thr Ile Leu Pro Ser Ser Met Ala Asp Glu Phe		
	100	105	110
35	Arg Lys Met Lys Glu Leu Cys Met Tyr Lys Phe Leu Gly Thr Asp Phe		
	115	120	125
	His Ser His Leu Pro Gly Phe Asp Gly Phe Lys Glu Val Thr Arg Asp		
	130	135	140
	Ala His Leu Ile Thr Lys Val Val Met Asn Gln Phe Gln Thr Gln Ala		
40	145	150	155
	Pro Lys Tyr Val Lys Pro Leu Ala Asn Glu Ala Ser Gly Ile Ile Thr		
	165	170	175
	Asp Ile Phe Gly Asp Ser Asn Glu Trp His Thr Val Pro Val Tyr Asn		
	180	185	190
45	Gln Cys Leu Asp Leu Val Thr Arg Thr Val Thr Phe Ile Met Val Gly		
	195	200	205
	Ser Lys Leu Ala His Asn Glu Glu Trp Leu Asp Ile Ala Lys His His		
	210	215	220
	Ala Val Thr Met Ala Ile Gln Ala Arg Gln Leu Arg Leu Trp Pro Val		
50	225	230	235
	Ile Leu Arg Pro Leu Val His Trp Leu Glu Pro Gln Gly Ala Lys Leu		
	245	250	255
	Arg Ala Gln Val Arg Arg Ala Arg Gln Leu Leu Asp Pro Ile Ile Gln		
	260	265	270
55	Glu Arg Arg Ala Glu Arg Asp Ala Cys Arg Ala Lys Gly Ile Glu Pro		
	275	280	285
	Pro Arg Tyr Val Asp Ser Ile Gln Trp Phe Glu Asp Thr Ala Lys Gly		

	290	295	300
	Lys Trp Tyr Asp Ala Ala Gly Ala Gln Leu Ala Met Asp Phe Ala Gly		
	305	310	315
	Ile Tyr Gly Thr Ser Asp Leu Leu Ile Gly Gly Leu Val Asp Ile Val		320
5	325	330	335
	Arg His Pro His Leu Leu Glu Pro Leu Arg Asp Glu Ile Arg Thr Val		
	340	345	350
	Ile Gly Gln Gly Gly Trp Thr Pro Ala Ser Leu Tyr Lys Leu Lys Leu		
	355	360	365
10	Leu Asp Ser Cys Leu Lys Glu Ser Gln Arg Val Lys Pro Val Glu Cys		
	370	375	380
	Ala Thr Met Arg Ser Tyr Ala Leu Gln Asp Val Thr Phe Ser Asn Gly		
	385	390	395
	Thr Phe Ile Pro Lys Gly Glu Leu Val Ala Val Ala Asp Arg Met		400
15	405	410	415
	Ser Asn Pro Glu Val Trp Pro Glu Pro Ala Lys Tyr Asp Pro Tyr Arg		
	420	425	430
	Tyr Met Arg Leu Arg Glu Asp Pro Ala Lys Ala Phe Ser Ala Gln Leu		
	435	440	445
20	Glu Asn Thr Asn Gly Asp His Ile Gly Phe Gly Trp His Pro Arg Ala		
	450	455	460
	Cys Pro Gly Arg Phe Phe Ala Ser Lys Glu Ile Lys Met Met Leu Ala		
	465	470	475
	Tyr Leu Leu Ile Arg Tyr Asp Trp Lys Val Val Pro Asp Glu Pro Leu		480
25	485	490	495
	Gln Tyr Tyr Arg His Ser Phe Ser Val Arg Ile His Pro Thr Thr Lys		
	500	505	510
	Leu Met Met Arg Arg Asp Glu Asp Ile Arg Leu Pro Gly Ser Leu		
	515	520	525
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	<213> Gibberella fujikuroi CAA75567		
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	<400> 33		
	Met Lys Tyr Thr Thr Cys Gln Met Asn Ile Phe Pro Ser Leu Trp Ser		
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	Met Lys Thr Ser Phe Arg Trp Pro Arg Thr Ser Lys Trp Ser Ser Val		
40	20	25	30
	Ser Leu Tyr Asp Met Met Leu Arg Thr Val Ala Leu Leu Ser Gly Arg		
	35	40	45
	Ala Phe Val Gly Leu Pro Leu Cys Arg Asp Glu Gly Trp Leu Gln Ala		
	50	55	60
45	Ser Ile Gly Tyr Thr Val Gln Cys Val Ser Ile Arg Asp Gln Leu Phe		
	65	70	75
	Thr Trp Ser Pro Val Leu Arg Pro Ile Ile Gly Pro Phe Leu Pro Ser		80
	85	90	95
	Val Arg Ser Val Arg Arg His Leu Arg Phe Ala Ala Glu Ile Met Ala		
50	100	105	110
	Pro Leu Ile Ser Gln Ala Leu Gln Asp Glu Lys Gln His Arg Ala Asp		
	115	120	125
	Thr Leu Leu Ala Asp Gln Thr Glu Gly Arg Gly Thr Phe Ile Ser Trp		
	130	135	140
55	Leu Leu Arg His Leu Pro Glu Glu Leu Arg Thr Pro Glu Gln Val Gly		
	145	150	155
	Leu Asp Gln Met Leu Val Ser Phe Ala Ala Ile His Thr Thr Met		160

	165	170	175
	Ala Leu Thr Lys Val Val Trp Glu Leu Val Lys Arg Pro Glu Tyr Ile		
	180	185	190
	Glu Pro Leu Arg Thr Glu Met Gln Asp Val Phe Gly Pro Asp Ala Val		
5	195	200	205
	Ser Pro Asp Ile Cys Ile Asn Lys Glu Ala Leu Ser Arg Leu His Lys		
	210	215	220
	Leu Asp Ser Phe Ile Arg Glu Val Gln Arg Trp Cys Pro Ser Thr Phe		
	225	230	235
			240
10	Val Thr Pro Ser Arg Arg Val Met Lys Ser Met Thr Leu Ser Asn Gly		
	245	250	255
	Ile Lys Leu Gln Arg Gly Thr Ser Ile Ala Phe Pro Ala His Ala Ile		
	260	265	270
	His Met Ser Glu Glu Thr Pro Thr Phe Ser Pro Asp Phe Ser Ser Asp		
15	275	280	285
	Phe Glu Asn Pro Ser Pro Arg Ile Phe Asp Gly Phe Arg Tyr Leu Asn		
	290	295	300
	Leu Arg Ser Ile Lys Gly Gln Gly Ser Gln His Gln Ala Ala Thr Thr		
	305	310	315
			320
20	Gly Pro Asp Tyr Leu Ile Phe Asn His Gly Lys His Ala Cys Pro Gly		
	325	330	335
	Arg Phe Phe Ala Ile Ser Glu Ile Lys Met Ile Leu Ile Glu Leu Leu		
	340	345	350
	Ala Lys Tyr Asp Phe Arg Leu Glu Asp Gly Lys Pro Gly Pro Glu Leu		
25	355	360	365
	Met Arg Val Gly Thr Glu Thr Arg Leu Asp Thr Lys Ala Gly Leu Glu		
	370	375	380
	Met Arg Arg Arg		
	385		
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	<210> 34		
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	<213> Gibberella fujikuroi CAA76703		
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	<400> 34		
	Met Ser Lys Ser Asn Ser Met Asn Ser Thr Ser His Glu Thr Leu Phe		
	1	5	10
	Gln Gln Leu Val Leu Gly Leu Asp Arg Met Pro Leu Met Asp Val His		
40	20	25	30
	Trp Leu Ile Tyr Val Ala Phe Gly Ala Trp Leu Cys Ser Tyr Val Ile		
	35	40	45
	His Val Leu Ser Ser Ser Thr Val Lys Val Pro Val Val Gly Tyr		
	50	55	60
45	Arg Ser Val Phe Glu Pro Thr Trp Leu Leu Arg Leu Arg Phe Val Trp		
	65	70	75
	Glu Gly Gly Ser Ile Ile Gly Gln Gly Tyr Asn Lys Phe Lys Asp Ser		
	85	90	95
	Ile Phe Gln Val Arg Lys Leu Gly Thr Asp Ile Val Ile Ile Pro Pro		
50	100	105	110
	Asn Tyr Ile Asp Glu Val Arg Lys Leu Ser Gln Asp Lys Thr Arg Ser		
	115	120	125
	Val Glu Pro Phe Ile Asn Asp Phe Ala Gly Gln Tyr Thr Arg Gly Met		
	130	135	140
55	Val Phe Leu Gln Ser Asp Leu Gln Asn Arg Val Ile Gln Gln Arg Leu		
	145	150	155
	Thr Pro Lys Leu Val Ser Leu Thr Lys Val Met Lys Glu Glu Leu Asp		160

	165	170	175
	Tyr Ala Leu Thr Lys Glu Met Pro Asp Met Lys Asn Asp Glu Trp Val		
	180	185	190
	Glu Val Asp Ile Ser Ser Ile Met Val Arg Leu Ile Ser Arg Ile Ser		
5	195	200	205
	Ala Arg Val Phe Leu Gly Pro Glu His Cys Arg Asn Gln Glu Trp Leu		
	210	215	220
	Thr Thr Thr Ala Glu Tyr Ser Glu Ser Leu Phe Ile Thr Gly Phe Ile		
	225	230	235
10	Leu Arg Val Val Pro His Ile Leu Arg Pro Phe Ile Ala Pro Leu Leu	240	
	245	250	255
	Pro Ser Tyr Arg Thr Leu Leu Arg Asn Val Ser Ser Gly Arg Arg Val		
	260	265	270
	Ile Gly Asp Ile Ile Arg Ser Gln Gln Gly Asp Gly Asn Glu Asp Ile		
15	275	280	285
	Leu Ser Trp Met Arg Asp Ala Ala Thr Gly Glu Glu Lys Gln Ile Asp		
	290	295	300
	Asn Ile Ala Gln Arg Met Leu Ile Leu Ser Leu Ala Ser Ile His Thr		
	305	310	315
20	Thr Ala Met Thr Met Thr His Ala Met Tyr Asp Leu Cys Ala Cys Pro	320	
	325	330	335
	Glu Tyr Ile Glu Pro Leu Arg Asp Glu Val Lys Ser Val Val Gly Ala		
	340	345	350
	Ser Gly Trp Asp Lys Thr Ala Leu Asn Arg Phe His Lys Leu Asp Ser		
25	355	360	365
	Phe Leu Lys Glu Ser Gln Arg Phe Asn Pro Val Phe Leu Leu Thr Phe		
	370	375	380
	Asn Arg Ile Tyr His Gln Ser Met Thr Leu Ser Asp Gly Thr Asn Ile		
	385	390	395
30	400	405	410
	Pro Ser Gly Thr Arg Ile Ala Val Pro Ser His Ala Met Leu Gln Asp	415	
	405	410	415
	Ser Ala His Val Pro Gly Pro Thr Pro Pro Thr Glu Phe Asp Gly Phe		
	420	425	430
	Arg Tyr Ser Lys Ile Arg Ser Asp Ser Asn Tyr Ala Gln Lys Tyr Leu		
35	435	440	445
	Phe Ser Met Thr Asp Ser Ser Asn Met Ala Phe Gly Tyr Gly Lys Tyr		
	450	455	460
	Ala Cys Pro Gly Arg Phe Tyr Ala Ser Asn Glu Met Lys Leu Thr Leu		
	465	470	475
40	480	485	490
	Ala Ile Leu Leu Gln Phe Glu Phe Lys Leu Pro Asp Gly Lys Gly	495	
	485	490	495
	Arg Pro Arg Asn Ile Thr Ile Asp Ser Asp Met Ile Pro Asp Pro Arg		
	500	505	510
	Ala Arg Leu Cys Val Arg Lys Arg Ser Leu Arg Asp Glu		
45	515	520	525
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	Ala Arg Ile Lys Asp Gln Trp Thr Lys Gly Arg Lys Arg Val Met Ala	15	
55	20	25	30
	Ser Met Arg Glu Arg Gln Glu Lys Gly Gly Asn Leu Glu Asp Pro Pro		

	35	40	45
	Thr Met Leu Asp His Leu Ser Asn Gly Arg Asn Glu His Ile Ala Asp		
	50	55	60
	Asp Val Glu Leu Gln Leu Leu His Gln Met Thr Leu Ile Ala Val Gly		
5	65	70	75
	Thr Val Thr Thr Phe Ser Ser Thr Thr Gln Ala Ile Tyr Asp Leu Val		
	85	90	95
	Ala His Pro Glu Tyr Ile Thr Ile Leu Arg Glu Glu Val Glu Ser Val		
	100	105	110
10	Pro Arg Asp Pro Asn Gly Asn Phe Thr Lys Asp Ser Thr Val Ala Met		
	115	120	125
	Asp Lys Leu Asp Ser Phe Leu Lys Glu Ser Gln Arg Phe Asn Ser Pro		
	130	135	140
	Asp Leu Ser Met Ser Asn Leu Lys Asn Tyr Lys Leu Cys Glu Ser Leu		
15	145	150	155
	Thr Gly His Ser Asn Leu Pro Thr Arg Thr Ile Ala Asp Met Lys Leu		
	165	170	175
	Pro Asp Gly Thr Phe Val Pro Lys Gly Thr Lys Leu Glu Ile Asn Thr		
	180	185	190
20	Cys Ser Ile His Lys Asp His Lys Leu Tyr Glu Asn Pro Glu Gln Phe		
	195	200	205
	Asp Gly Leu Arg Phe His Lys Trp Arg Lys Ala Pro Gly Lys Glu Lys		
	210	215	220
	Arg Tyr Met Tyr Ser Ser Gly Thr Asp Asp Leu Ser Trp Gly Phe		
25	225	230	235
	Gly Arg His Ala Cys Pro Gly Arg Tyr Leu Ser Ala Ile Asn Ile Lys		
	245	250	255
	Leu Ile Met Ala Glu Leu Leu Met Asn Tyr Asp Ile Lys Leu Pro Asp		
	260	265	270
30	Gly Leu Ser Arg Pro Lys Asn Ile Glu Phe Glu Val Leu Ala Ser Leu		
	275	280	285
	Asn Ala Cys Ala Asn Ala		
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35	<210> 36		
	<211> 510		
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	<213> Caenorhabditis elegans CAA91268		
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	Phe Ile Tyr Ile Ile Leu Ala Arg Arg Glu Arg Phe Lys Leu Arg Glu		
	20	25	30
45	Lys Ile Gly Leu Ser Gly Pro Glu Pro His Trp Phe Leu Gly Asn Leu		
	35	40	45
	Lys Gln Thr Ala Glu Arg Lys Glu Lys Leu Gly Tyr Asp Asp Ala Asn		
	50	55	60
	Arg Trp Phe Asn Glu Leu His Glu Gln Tyr Gly Glu Thr Phe Gly Ile		
50	65	70	75
	Tyr Tyr Gly Ser Gln Met Asn Ile Val Ile Ser Asn Glu Lys Asp Ile		
	85	90	95
	Lys Glu Val Phe Ile Lys Asn Phe Ser Asn Phe Ser Asp Arg Ser Val		
	100	105	110
55	Pro Ser Ile Tyr Glu Ala Asn Gln Leu Thr Ala Ser Leu Leu Met Asn		
	115	120	125
	Ser Tyr Ser Ser Gly Trp Lys His Thr Arg Ser Ala Ile Ala Pro Ile		

	130	135	140
	Phe Ser Thr Gly Lys Met	Lys Ala Met Gln Glu	Thr Ile Asn Ser Lys
	145	150	155
	Val Asp Leu Phe Leu Asp Ile	Leu Arg Glu Lys Ala Ser Ser	Gly Gln
5	165	170	175
	Lys Trp Asp Ile Tyr Asp Asp	Phe Gln Gly Leu Thr	Leu Asp Val Ile
	180	185	190
	Gly Lys Cys Ala Phe Ala Ile	Asp Ser Asn Cys Gln Arg Asp Arg	Asn
	195	200	205
10	Asp Val Phe Tyr His Pro Val	Thr Val Lys Ile Thr Ile Asn Asn	Phe
	210	215	220
	Thr Tyr Phe His Ser Ser	Pro Gly Thr Phe His Phe	Leu Glu Ser
	225	230	235
	Thr Leu Gln Ile His Thr Thr	Gly Arg Cys Arg Asn Ser	Thr Cys Arg
15	245	250	255
	Arg Thr Val Lys Cys Val	Gly Phe Arg Gln Asp Lys Ala	Lys Phe Cys
	260	265	270
	Ser Asp Tyr Glu Arg Arg	Gly Glu Gly Ser Asp Ser	Val Asp
	275	280	285
20	Leu Leu Lys Leu Leu Asn Arg	Glu Asp Asp Lys Ser Lys	Pro Met
	290	295	300
	Thr Lys Gln Glu Val Ile	Glu Asn Cys Phe Ala Phe	Leu Leu Ala Gly
	305	310	315
	Tyr Glu Thr Thr Ser Thr Ala Met	Thr Tyr Cys Ser Tyr	Leu Leu Ser
25	325	330	335
	Lys Tyr Pro Asn Val Gln Gln	Lys Leu Tyr Glu Glu Ile	Met Glu Ala
	340	345	350
	Lys Glu Asn Gly Gly Leu Thr	Tyr Asp Ser Ile His Asn Met	Lys Tyr
	355	360	365
30	Leu Asp Cys Val Tyr Lys Glu	Thr Leu Arg Phe Tyr Pro	Pro His Phe
	370	375	380
	Ser Phe Ile Arg Arg Leu Cys Arg	Glu Asp Ile Thr Ile Arg	Gly Gln
	385	390	395
	Phe Tyr Pro Lys Gly Ala Ile Val Val	Cys Leu Pro His Thr	Val His
35	405	410	415
	Arg Asn Pro Glu Asn Trp Asp	Ser Pro Glu Glu Phe His	Pro Glu Arg
	420	425	430
	Phe Glu Asn Trp Glu Glu Lys	Ser Ser Ser Leu Lys Trp Ile	Pro Phe
	435	440	445
40	Gly Val Gly Pro Arg Tyr Cys	Val Gly Met Arg Phe	Ala Glu Met Glu
	450	455	460
	Phe Lys Thr Thr Ile Val Lys	Leu Leu Asp Thr Phe	Glu Leu Lys Gln
	465	470	475
	Phe Glu Gly Glu Ala Asp Leu Ile	Pro Asp Cys Asn Gly	Val Ile Met
45	485	490	495
	Arg Pro Asn Asp Pro Val Arg	Leu His Leu Lys Pro Arg	Asn
	500	505	510
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	<211> 691		
	<212> PRT		
	<213> yeast P450 reductase		
	<400> 37		
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	1	5	10
	Val Leu Ala Val Leu Leu Tyr	Val Lys Arg Asn Ser	Ile Lys Glu Leu

	20	25	30
	Leu Met Ser Asp Asp Gly Asp Ile Thr Ala Val Ser Ser Gly Asn Arg		
	35	40	45
	Asp Ile Ala Gln Val Val Thr Glu Asn Asn Lys Asn Tyr Leu Val Leu		
5	50	55	60
	Tyr Ala Ser Gln Thr Gly Thr Ala Glu Asp Tyr Ala Lys Lys Phe Ser		
	65	70	75
	Lys Glu Leu Val Ala Lys Phe Asn Leu Asn Val Met Cys Ala Asp Val		
	85	90	95
10	Glu Asn Tyr Asp Phe Glu Ser Leu Asn Asp Val Pro Val Ile Val Ser		
	100	105	110
	Ile Phe Ile Ser Thr Tyr Gly Glu Gly Asp Phe Pro Asp Gly Ala Val		
	115	120	125
15	Asn Phe Glu Asp Phe Ile Cys Asn Ala Glu Ala Gly Ala Leu Ser Asn		
	130	135	140
	Leu Arg Tyr Asn Met Phe Gly Leu Gly Asn Ser Thr Tyr Glu Phe Phe		
	145	150	155
	Asn Gly Ala Ala Lys Lys Ala Glu Lys His Leu Ser Ala Ala Gly Ala		
	165	170	175
20	Ile Arg Leu Gly Lys Leu Gly Glu Ala Asp Asp Gly Ala Gly Thr Thr		
	180	185	190
	Asp Glu Asp Tyr Met Ala Trp Lys Asp Ser Ile Leu Glu Val Leu Lys		
	195	200	205
25	Asp Glu Leu His Leu Asp Glu Gln Glu Ala Lys Phe Thr Ser Gln Phe		
	210	215	220
	Gln Tyr Thr Val Leu Asn Glu Ile Thr Asp Ser Met Ser Leu Gly Glu		
	225	230	235
	Pro Ser Ala His Tyr Leu Pro Ser His Gln Leu Asn Arg Asn Ala Asp		
	245	250	255
30	Gly Ile Gln Leu Gly Pro Phe Asp Leu Ser Gln Pro Tyr Ile Ala Pro		
	260	265	270
	Ile Val Lys Ser Arg Glu Leu Phe Ser Ser Asn Asp Arg Asn Cys Ile		
	275	280	285
35	His Ser Glu Phe Asp Leu Ser Gly Ser Asn Ile Lys Tyr Ser Thr Gly		
	290	295	300
	Asp His Leu Ala Val Trp Pro Ser Asn Pro Leu Glu Lys Val Glu Gln		
	305	310	315
	Phe Leu Ser Ile Phe Asn Leu Asp Pro Glu Thr Ile Phe Asp Leu Lys		
	325	330	335
40	Pro Leu Asp Pro Thr Val Lys Val Pro Phe Pro Thr Pro Thr Ile		
	340	345	350
	Gly Ala Ala Ile Lys His Tyr Leu Glu Ile Thr Gly Pro Val Ser Arg		
	355	360	365
45	Gln Leu Phe Ser Ser Leu Ile Gln Phe Ala Pro Asn Ala Asp Val Lys		
	370	375	380
	Glu Lys Leu Thr Leu Leu Ser Lys Asp Lys Asp Gln Phe Ala Val Glu		
	385	390	395
	Ile Thr Ser Lys Tyr Phe Asn Ile Ala Asp Ala Leu Lys Tyr Leu Ser		
	405	410	415
50	Asp Gly Ala Lys Trp Asp Asn Val Pro Met Gln Phe Leu Val Glu Ser		
	420	425	430
	Val Pro Gln Met Thr Pro Arg Tyr Tyr Ser Ile Ser Ser Ser Leu		
	435	440	445
55	Ser Glu Lys Gln Thr Val His Val Thr Ser Ile Val Glu Asn Phe Pro		
	450	455	460
	Asn Pro Glu Leu Pro Asp Ala Pro Pro Gly Val Gly Val Thr Thr Asn		
	465	470	475
			480

Leu Leu Arg Asn Ile Gln Leu Ala Gln Asn Asn Val Asn Ile Ala Glu
 485 490 495
 Thr Asn Leu Pro Val His Tyr Asp Leu Asn Gly Pro Arg Lys Leu Phe
 500 505 510
 5 Ala Asn Tyr Lys Leu Pro Val His Val Arg Arg Ser Asn Phe Arg Leu
 515 520 525
 Pro Ser Asn Pro Ser Thr Pro Val Ile Met Ile Gly Pro Gly Thr Gly
 530 535 540
 Val Ala Pro Phe Arg Gly Phe Ile Arg Glu Arg Val Ala Phe Leu Glu
 10 545 550 555 560
 Ser Gln Lys Lys Gly Gly Asn Asn Val Ser Leu Gly Lys His Ile Leu
 565 570 575
 Phe Tyr Gly Ser Arg Asn Thr Asp Asp Phe Leu Tyr Gln Asp Glu Trp
 580 585 590
 15 Pro Glu Tyr Ala Lys Lys Leu Asp Gly Ser Phe Glu Met Val Val Ala
 595 600 605
 His Ser Arg Leu Pro Asn Thr Lys Lys Val Tyr Val Gln Asp Lys Leu
 610 615 620
 Lys Asp Tyr Glu Asp Gln Val Phe Glu Met Ile Asn Asn Gly Ala Phe
 20 625 630 635 640
 Ile Tyr Val Cys Gly Asp Ala Lys Gly Met Ala Lys Gly Val Ser Thr
 645 650 655
 Ala Leu Val Gly Ile Leu Ser Arg Gly Lys Ser Ile Thr Thr Asp Glu
 660 665 670
 25 Ala Thr Glu Leu Ile Lys Met Leu Lys Thr Ser Gly Arg Tyr Gln Glu
 675 680 685
 Asp Val Trp
 690

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 35 40 45
 Ala Gly Lys Thr Arg Asn Ile Ile Glu Lys Met Glu Glu Thr Gly Lys
 50 55 60
 Asn Cys Val Ile Phe Tyr Gly Ser Gln Thr Gly Thr Ala Glu Asp Tyr
 45 65 70 75 80
 Ala Ser Arg Leu Ala Lys Glu Gly Ser Gln Arg Phe Gly Leu Lys Thr
 85 90 95
 Met Val Ala Asp Leu Glu Glu Tyr Asp Tyr Glu Asn Leu Asp Gln Phe
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 50 Pro Glu Asp Lys Val Ala Phe Phe Val Leu Ala Thr Tyr Gly Glu Gly
 115 120 125
 Glu Pro Thr Asp Asn Ala Val Glu Phe Tyr Gln Phe Phe Thr Gly Asp
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 Asp Val Ala Phe Glu Ser Ala Ser Ala Asp Glu Lys Pro Leu Ser Lys
 55 145 150 155 160
 Leu Lys Tyr Val Ala Phe Gly Leu Gly Asn Asn Thr Tyr Glu His Tyr
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Asn Ala Met Val Arg Gln Val Asp Ala Ala Phe Gln Lys Leu Gly Pro
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 Gln Arg Ile Gly Ser Ala Gly Glu Gly Asp Asp Gly Ala Gly Thr Met
 195 200 205
 5 Glu Glu Asp Phe Leu Ala Trp Lys Glu Pro Met Trp Ala Ala Leu Ser
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 Glu Ser Met Asp Leu Glu Glu Arg Glu Ala Val Tyr Glu Pro Val Phe
 225 230 235 240
 Cys Val Thr Glu Asn Glu Ser Leu Ser Pro Glu Asp Glu Thr Val Tyr
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 260 265 270
 Tyr Ser Ala His Asn Pro Phe Ile Ala Pro Ile Ala Glu Ser Arg Glu
 275 280 285
 15 Leu Phe Thr Val Lys Asp Arg Asn Cys Leu His Met Glu Ile Ser Ile
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 Ala Gly Ser Asn Leu Ser Tyr Gln Thr Gly Asp His Ile Ala Val Trp
 305 310 315 320
 Pro Thr Asn Ala Gly Ala Glu Val Asp Arg Phe Leu Gln Val Phe Gly
 20 325 330 335
 Leu Glu Gly Lys Arg Asp Ser Val Ile Asn Ile Lys Gly Ile Asp Val
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 Thr Ala Lys Val Pro Ile Pro Thr Pro Thr Thr Tyr Asp Ala Ala Val
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 25 Arg Tyr Tyr Met Glu Val Cys Ala Pro Val Ser Arg Gln Phe Val Ala
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 Thr Leu Ala Ala Phe Ala Pro Met Arg Lys Ala Arg Gln Arg Leu Cys
 385 390 395 400
 Val Trp Val Ala Gln Gly Leu Phe Pro Arg Glu Gly His Gln Pro Met
 30 405 410 415
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 Ala Val Pro Phe Ser Leu Leu Ile Glu Gly Ile Thr Lys Leu Gln Pro
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 35 Arg Tyr Tyr Ser Ile Ser Ser Ser Leu Val Gln Lys Asp Lys Ile
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 Met Val Lys Gly Val Thr Thr Asn Tyr Leu Leu Ala Leu Lys Gln Lys
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 Gln Asn Gly Arg Ser Leu Ser Arg Pro Ser Arg Leu Asp Leu Leu His
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 His Gly Pro Arg Asn Lys Tyr Asp Gly Ile His Val Pro Val His Val
 515 520 525
 45 Arg His Ser Asn Phe Lys Leu Pro Ser Asp Pro Ser Arg Pro Ile Ile
 530 535 540
 Met Val Gly Pro Gly Thr Gly Val Ala Pro Phe Arg Gly Phe Ile Gln
 545 550 555 560
 Glu Arg Ala Ala Leu Ala Ala Lys Gly Glu Lys Val Gly Pro Thr Val
 50 565 570 575
 Leu Phe Phe Gly Cys Arg Lys Ser Asp Glu Asp Phe Leu Tyr Lys Asp
 580 585 590
 Glu Trp Lys Thr Tyr Gln Asp Gln Leu Gly Asp Asn Leu Lys Ile Ile
 595 600 605
 55 Thr Ala Phe Ser Arg Glu Gly Pro Gln Lys Val Tyr Val Gln His Arg
 610 615 620
 Leu Arg Glu His Ser Glu Leu Val Ser Asp Leu Leu Lys Gln Lys Ala

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	Leu Val Leu Gly Gln Ile Ile Ala Ala Gln Arg Gly Leu Pro Ala Glu			
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	Leu Ile Val Gly Val Leu Thr Tyr Trp Phe Ile Phe Lys Lys Lys Lys			
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	Glu Glu Ile Pro Glu Phe Ser Lys Ile Gln Thr Thr Ala Pro Pro Val			
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	Ile Val Phe Tyr Gly Ser Gln Thr Gly Thr Ala Glu Glu Phe Ala Asn			
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	Arg Leu Ser Lys Asp Ala His Arg Tyr Gly Met Arg Gly Met Ser Ala			
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	Asp Pro Glu Glu Tyr Asp Leu Ala Asp Leu Ser Ser Leu Pro Glu Ile			
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	Asp Lys Ser Leu Val Val Phe Cys Met Ala Thr Tyr Gly Glu Gly Asp			
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	145	150	155	160
	Val Asp Leu Thr Gly Val Lys Phe Ala Val Phe Gly Leu Gly Asn Lys			
	165	170	175	
	Thr Tyr Glu His Phe Asn Ala Met Gly Lys Tyr Val Asp Gln Arg Leu			
40	180	185	190	
	Glu Gln Leu Gly Ala Gln Arg Ile Phe Glu Leu Gly Leu Gly Asp Asp			
	195	200	205	
	Asp Gly Asn Leu Glu Glu Asp Phe Ile Thr Trp Arg Glu Gln Phe Trp			
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	225	230	235	240
	Ser Ile Arg Gln Tyr Glu Leu Val Val His Glu Asp Met Asp Thr Ala			
	245	250	255	
	Lys Val Tyr Thr Gly Glu Met Gly Arg Leu Lys Ser Tyr Glu Asn Gln			
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	Lys Pro Pro Phe Asp Ala Lys Asn Pro Phe Leu Ala Ala Val Thr Thr			
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	Asn Arg Lys Leu Asn Gln Gly Thr Glu Arg His Leu Met His Leu Glu			
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55	Leu Asp Ile Ser Asp Ser Lys Ile Arg Tyr Glu Ser Gly Asp His Val			
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	Ala Val Tyr Pro Ala Asn Asp Ser Thr Leu Val Asn Gln Ile Gly Glu			

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	Ile Leu Gly Ala Asp Leu Asp Val Ile Met Ser Leu Asn Asn Leu Asp		
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	Glu Glu Ser Asn Lys Lys His Pro Phe Pro Cys Pro Thr Thr Tyr Arg		
5	355	360	365
	Thr Ala Leu Thr Tyr Tyr Leu Asp Ile Thr Asn Pro Pro Arg Thr Asn		
	370	375	380
	Val Leu Tyr Glu Leu Ala Gln Tyr Ala Ser Glu Pro Ser Glu Gln Glu		
	385	390	395
			400
10	His Leu His Lys Met Ala Ser Ser Ser Gly Glu Gly Lys Glu Leu Tyr		
	405	410	415
	Leu Ser Trp Val Val Glu Ala Arg Arg His Ile Leu Ala Ile Leu Gln		
	420	425	430
	Asp Tyr Pro Ser Leu Arg Pro Pro Ile Asp His Leu Cys Glu Leu Leu		
15	435	440	445
	Pro Arg Leu Gln Ala Arg Tyr Tyr Ser Ile Ala Ser Ser Ser Lys Val		
	450	455	460
	His Pro Asn Ser Val His Ile Cys Ala Val Ala Val Glu Tyr Glu Ala		
	465	470	475
			480
20	Lys Ser Gly Arg Val Asn Lys Gly Val Ala Thr Ser Trp Leu Arg Thr		
	485	490	495
	Lys Glu Pro Ala Gly Glu Asn Gly Arg Arg Ala Leu Val Pro Met Phe		
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	Val Arg Lys Ser Gln Phe Arg Leu Pro Phe Lys Pro Thr Thr Pro Val		
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	Ile Met Val Gly Pro Gly Thr Gly Val Ala Pro Phe Met Gly Phe Ile		
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	Gln Glu Arg Ala Trp Leu Arg Glu Gln Gly Lys Glu Val Gly Glu Thr		
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			560
30	Leu Leu Tyr Tyr Gly Cys Arg Arg Ser Asp Glu Asp Tyr Leu Tyr Arg		
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	Glu Glu Leu Ala Arg Phe His Lys Asp Gly Ala Leu Thr Gln Leu Asn		
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	Val Ala Phe Ser Arg Glu Gln Ala His Lys Val Tyr Val Gln His Leu		
35	595	600	605
	Leu Lys Arg Asp Lys Glu His Leu Trp Lys Leu Ile His Glu Gly Gly		
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	Ala His Ile Tyr Val Cys Gly Asp Ala Arg Asn Met Ala Lys Asp Val		
	625	630	635
40	Gln Asn Thr Phe Tyr Asp Ile Val Ala Glu Phe Gly Pro Met Glu His		
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	Thr Gln Ala Val Asp Tyr Val Lys Lys Leu Met Thr Lys Gly Arg Tyr		
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	tcctctgtca	gagagagcag	cttlytgaa	aagatgaaga	aaacggggag	gaacatcatc
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 35 40 45
 Glu Val Pro Glu Phe Thr Lys Ile Gln Thr Leu Thr Ser Ser Val Arg
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 10 Glu Ser Ser Phe Val Glu Lys Met Lys Lys Thr Gly Arg Asn Ile Ile
 65 70 75 80
 Val Phe Tyr Gly Ser Gln Thr Gly Thr Ala Glu Glu Phe Ala Asn Arg
 85 90 95
 Leu Ser Lys Asp Ala His Arg Tyr Gly Met Arg Gly Met Ser Ala Asp
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 Pro Glu Glu Tyr Asp Leu Ala Asp Leu Ser Ser Leu Pro Glu Ile Asp
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 Asn Ala Leu Val Val Phe Cys Met Ala Thr Tyr Gly Glu Gly Asp Pro
 130 135 140
 20 Thr Asp Asn Ala Gln Asp Phe Tyr Asp Trp Leu Gln Glu Thr Asp Val
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 Asp Leu Ser Gly Val Lys Phe Ala Val Phe Gly Leu Gly Asn Lys Thr
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 Tyr Glu His Phe Asn Ala Met Gly Lys Tyr Val Asp Lys Arg Leu Glu
 25 180 185 190
 Gln Leu Gly Ala Gln Arg Ile Phe Glu Leu Gly Leu Gly Asp Asp Asp
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 Gly Asn Leu Glu Glu Asp Phe Ile Thr Trp Arg Glu Gln Phe Trp Pro
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 225 230 235 240
 Ile Arg Gln Tyr Glu Leu Val Val His Thr Asp Ile Asp Ala Ala Lys
 245 250 255
 Val Tyr Met Gly Glu Met Gly Arg Leu Lys Ser Tyr Glu Asn Gln Lys
 35 260 265 270
 Pro Pro Phe Asp Ala Lys Asn Pro Phe Leu Ala Ala Val Thr Thr Asn
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 Arg Lys Leu Asn Gln Gly Thr Glu Arg His Leu Met His Leu Glu Leu
 290 295 300
 40 Asp Ile Ser Asp Ser Lys Ile Arg Tyr Glu Ser Gly Asp His Val Ala
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 325 330 335
 Leu Gly Ala Asp Leu Asp Val Val Met Ser Leu Asn Asn Leu Asp Glu
 45 340 345 350
 Glu Ser Asn Lys Lys His Pro Phe Pro Cys Pro Thr Ser Tyr Arg Thr
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 Ala Leu Thr Tyr Tyr Leu Asp Ile Thr Asn Pro Pro Arg Thr Asn Val
 370 375 380
 50 Leu Tyr Glu Leu Ala Gln Tyr Ala Ser Glu Pro Ser Glu Gln Glu Leu
 385 390 395 400
 Leu Arg Lys Met Ala Ser Ser Ser Gly Glu Gly Lys Glu Leu Tyr Leu
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 Ser Trp Val Val Glu Ala Arg Arg His Ile Leu Ala Ile Leu Gln Asp
 55 420 425 430
 Cys Pro Ser Leu Arg Pro Pro Ile Asp His Leu Cys Glu Leu Leu Pro
 435 440 445

Arg Leu Gln Ala Arg Tyr Tyr Ser Ile Ala Ser Ser Ser Lys Val His
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 Pro Asn Ser Val His Ile Cys Ala Val Val Val Glu Tyr Glu Thr Lys
 465 470 475 480
 5 Ala Gly Arg Ile Asn Lys Gly Val Ala Thr Asn Trp Leu Arg Ala Lys
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 Glu Pro Val Gly Glu Asn Gly Gly Arg Ala Leu Val Pro Met Phe Val
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 Arg Lys Ser Gln Leu Arg Leu Pro Phe Lys Ala Thr Thr Pro Val Ile
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 Glu Arg Ala Trp Leu Arg Gln Gln Gly Lys Glu Val Gly Glu Thr Leu
 545 550 555 560
 15 Leu Tyr Tyr Gly Cys Arg Arg Ser Asp Glu Asp Tyr Leu Tyr Arg Glu
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 Glu Leu Ala Gln Phe His Arg Asp Gly Ala Leu Thr Gln Leu Asn Val
 580 585 590
 Ala Phe Ser Arg Glu Gln Ser His Lys Val Tyr Val Gln His Leu Leu
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 Lys Gln Asp Arg Glu His Leu Trp Lys Leu Ile Glu Gly Gly Ala His
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 Ile Tyr Val Cys Gly Asp Ala Arg Asn Met Ala Arg Asp Val Gln Asn
 625 630 635 640
 25 Thr Phe Tyr Asp Ile Val Ala Glu Leu Gly Ala Met Glu His Ala Gln
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 Glu Glu Val Pro Glu Phe Thr Lys Ile Gln Thr Leu Thr Ser Ser Val
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 Ile Val Phe Tyr Gly Ser Gln Thr Gly Thr Ala Glu Glu Phe Ala Asn
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 Asp Pro Glu Glu Tyr Asp Leu Ala Asp Leu Ser Ser Leu Pro Glu Ile
 115 120 125
 Asp Asn Ala Leu Val Val Phe Cys Met Ala Thr Tyr Gly Glu Gly Asp
 55 130 135 140
 Pro Thr Asp Asn Ala Gln Asp Phe Tyr Asp Trp Leu Gln Glu Thr Asp
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 Thr Tyr Glu His Phe Asn Ala Met Gly Lys Tyr Val Asp Lys Arg Leu
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 Asp Gly Asn Leu Glu Asp Phe Ile Thr Trp Arg Glu Gln Phe Trp
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 Pro Ala Val Cys Glu His Phe Gly Val Glu Ala Thr Gly Glu Glu Ser
 10 225 230 235 240
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 15 Lys Pro Pro Phe Asp Ala Lys Asn Pro Phe Leu Ala Ala Val Thr Thr
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 Asn Arg Lys Leu Asn Gln Gly Thr Glu Arg His Leu Met His Leu Glu
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 325 330 335
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 25 Glu Glu Ser Asn Lys Lys His Pro Phe Pro Cys Pro Thr Ser Tyr Arg
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 Thr Ala Leu Thr Tyr Tyr Leu Asp Ile Thr Asn Pro Pro Arg Thr Asn
 370 375 380
 Val Leu Tyr Glu Leu Ala Gln Tyr Ala Ser Glu Pro Ser Glu Gln Glu
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 405 410 415
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 35 Asp Cys Pro Ser Leu Arg Pro Pro Ile Asp His Leu Cys Glu Leu Leu
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 Pro Arg Leu Gln Ala Arg Tyr Tyr Ser Ile Ala Ser Ser Ser Lys Val
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 His Pro Asn Ser Val His Ile Cys Ala Val Val Glu Tyr Glu Thr
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 Lys Glu Pro Ala Gly Glu Asn Gly Gly Arg Ala Leu Val Pro Met Phe
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 Ile Met Val Gly Pro Gly Thr Gly Val Ala Pro Phe Ile Gly Phe Ile
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 Gln Glu Arg Ala Trp Leu Arg Gln Gln Gly Lys Glu Val Gly Glu Thr
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 Glu Glu Leu Ala Gln Phe His Arg Asp Gly Ala Leu Thr Gln Leu Asn
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 55 Val Ala Phe Ser Arg Glu Gln Ser His Lys Val Tyr Val Gln His Leu
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	Asn Thr Phe Tyr Asp Ile Val Ala Glu Leu Gly Ala Met Glu His Ala		640
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	Ile Val Phe Tyr Gly Ser Gln Thr Gly Thr Ala Glu Glu Phe Ala Asn		80
	85	90	95
	Arg Leu Ser Lys Asp Ala His Arg Tyr Gly Met Arg Gly Met Ser Ala		
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	Asp Pro Glu Glu Tyr Asp Leu Ala Asp Leu Ser Ser Leu Pro Glu Ile		
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35	Pro Thr Asp Asn Ala Gln Asp Phe Tyr Asp Trp Leu Gln Glu Thr Asp		
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	165	170	175
	Thr Tyr Glu His Phe Asn Ala Met Gly Lys Tyr Val Asp Lys Arg Leu		
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	Glu Gln Leu Gly Ala Gln Arg Ile Phe Glu Leu Gly Leu Gly Asp Asp		
	195	200	205
	Asp Gly Asn Leu Glu Glu Asp Phe Ile Thr Trp Arg Glu Gln Phe Trp		
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	Ser Ile Arg Gln Tyr Glu Leu Val Val His Thr Asp Ile Asp Ala Ala		240
	245	250	255
	Lys Val Tyr Met Gly Glu Met Gly Arg Leu Lys Ser Tyr Glu Asn Gln		
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	Lys Pro Pro Phe Asp Ala Lys Asn Pro Phe Leu Ala Ala Val Thr Thr		
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	Asn Arg Lys Leu Asn Gln Gly Thr Glu Arg His Leu Met His Leu Glu		
	290	295	300
55	Leu Asp Ile Ser Asp Ser Lys Ile Arg Tyr Glu Ser Gly Asp His Val		
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	Ala Val Tyr Pro Ala Asn Asp Ser Ala Leu Val Asn Gln Leu Gly Lys		320

	325	330	335
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	Glu Glu Ser Asn Lys Lys His Pro Phe Pro Cys Pro Thr Ser Tyr Arg		
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	Thr Ala Leu Thr Tyr Tyr Leu Asp Ile Thr Asn Pro Pro Arg Thr Asn		
	370	375	380
	Val Leu Tyr Glu Leu Ala Gln Tyr Ala Ser Glu Pro Ser Glu Gln Glu		
	385	390	395
			400
10	Leu Leu Arg Lys Met Ala Ser Ser Ser Gly Glu Gly Lys Glu Leu Tyr		
	405	410	415
	Leu Ser Trp Val Val Glu Ala Arg Arg His Ile Leu Ala Ile Leu Gln		
	420	425	430
	Asp Cys Pro Ser Leu Arg Pro Pro Ile Asp His Leu Cys Glu Leu Leu		
15	435	440	445
	Pro Arg Leu Gln Ala Arg Tyr Tyr Ser Ile Ala Ser Ser Ser Lys Val		
	450	455	460
	His Pro Asn Ser Val His Ile Cys Ala Val Val Val Glu Tyr Glu Thr		
	465	470	475
			480
20	Lys Ala Gly Arg Ile Asn Lys Gly Val Ala Thr Asn Trp Leu Arg Ala		
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	Lys Glu Pro Ala Gly Glu Asn Gly Gly Arg Ala Leu Val Pro Met Phe		
	500	505	510
	Val Arg Lys Ser Gln Phe Arg Leu Pro Phe Lys Ala Thr Thr Pro Val		
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	Ile Met Val Gly Pro Gly Thr Gly Val Ala Pro Phe Ile Gly Phe Ile		
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	Gln Glu Arg Ala Trp Leu Arg Gln Gln Gly Lys Glu Val Gly Glu Thr		
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			560
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	Glu Glu Leu Ala Gln Phe His Arg Asp Gly Ala Leu Thr Gln Leu Asn		
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	Val Ala Phe Ser Arg Glu Gln Ser His Lys Val Tyr Val Gln His Leu		
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	Leu Lys Gln Asp Arg Glu His Leu Trp Lys Leu Ile Glu Gly Gly Ala		
	610	615	620
	His Ile Tyr Val Cys Gly Asp Ala Arg Asn Met Ala Arg Asp Val Gln		
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			640
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	Asp Val Asp Leu Ser Gly Val Lys Tyr Ala Val Phe Gly Leu Gly Asn		
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	Lys Thr Tyr His Phe Asn Ala Met Gly Lys Tyr Val Asp Gln Arg		
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	Asp Asp Ala Asn Leu Glu Glu Asp Phe Ile Thr Trp Arg Gln Gln Phe		
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	Ser Ser Ile Arg Gln Tyr Glu Leu Val His Thr Asp Ile Asp Val		
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	Ala Lys Val Tyr Gln Gly Glu Met Gly Arg Leu Lys Ser Tyr Glu Asn		
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	Thr Asn Arg Lys Leu Asn Gln Gly Thr Glu Arg His Leu Met His Leu		
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	Glu Ile Leu Gly Ala Asp Leu Asp Val Val Met Ser Leu Asn Asn Leu		
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40	Asp Glu Glu Ser Asn Lys Lys His Pro Phe Pro Cys Pro Thr Ser Tyr		
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	Arg Thr Ala Leu Thr Tyr Tyr Leu Asp Ile Thr Asn Pro Pro Arg Thr		
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45	Asn Val Leu Tyr Glu Leu Ala Gln Tyr Ala Ala Asp Pro Ala Glu Gln		
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	Glu Gln Leu Arg Lys Met Ala Ser Ser Ser Gly Glu Gly Lys Glu Leu		
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	Tyr Leu Ser Trp Val Val Glu Ala Arg Arg His Ile Leu Ala Ile Leu		
	420	425	430
50	Gln Asp Tyr Pro Ser Leu Arg Pro Pro Ile Asp His Leu Cys Glu Leu		
	435	440	445
	Leu Pro Arg Leu Gln Ala Arg Tyr Tyr Ser Ile Ala Ser Ser Ser Lys		
	450	455	460
55	Val His Pro Asn Ser Val His Ile Cys Ala Val Ala Val Glu Tyr Glu		
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	Thr Lys Ala Gly Arg Leu Asn Lys Gly Val Ala Thr Ser Trp Leu Arg		
	485	490	495

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 Phe Val Arg Lys Ser Gln Phe Arg Leu Pro Phe Lys Ala Thr Thr Pro
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 5 Val Ile Met Val Gly Pro Gly Thr Gly Val Ala Pro Phe Ile Gly Phe
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 Ile Gln Glu Arg Ala Trp Leu Arg Gln Gln Gly Lys Glu Val Gly Glu
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 Thr Leu Leu Tyr Tyr Gly Cys Arg Arg Ala Ala Glu Asp Tyr Leu Tyr
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 Gly Ala His Ile Tyr Val Cys Gly Asp Ala Arg Asn Met Ala Arg Asp
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 Val Gln Asn Thr Phe Tyr Asp Ile Val Ala Glu Leu Gly Ala Met Glu
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 Glu Glu Ile Pro Glu Phe Ser Lys Ile Gln Thr Thr Ala Pro Pro Val
 50 55 60
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 Ile Val Phe Tyr Gly Ser Gln Thr Gly Thr Ala Glu Glu Phe Ala Asn
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 Arg Leu Ser Lys Asp Ala His Arg Tyr Gly Met Arg Gly Met Ser Ala
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 Asp Pro Glu Glu Tyr Asp Leu Ala Asp Leu Ser Ser Leu Pro Glu Ile
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 Asp Lys Ser Leu Val Val Phe Cys Met Ala Thr Tyr Gly Glu Gly Asp
 130 135 140
 50 Pro Thr Asp Asn Ala Gln Asp Phe Tyr Asp Trp Leu Gln Glu Thr Asp
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 Val Asp Leu Thr Gly Val Lys Phe Ala Val Phe Gly Leu Gly Asn Lys
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 Thr Tyr Glu His Phe Asn Ala Met Gly Lys Tyr Val Asp Gln Arg Leu
 55 180 185 190
 Glu Gln Leu Gly Ala Gln Arg Ile Phe Glu Leu Gly Leu Gly Asp Asp
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Asp Gly Asn Leu Glu Glu Asp Phe Ile Thr Trp Arg Glu Gln Phe Trp
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 Pro Ala Val Cys Glu Phe Phe Gly Val Glu Ala Thr Gly Glu Glu Ser
 225 230 235 240
 5 Ser Ile Arg Gln Tyr Glu Leu Val Val His Glu Asp Met Asp Val Ala
 245 250 255
 Lys Val Tyr Thr Gly Glu Met Gly Arg Leu Lys Ser Tyr Glu Asn Gln
 260 265 270
 Lys Pro Pro Phe Asp Ala Lys Asn Pro Phe Leu Ala Ala Val Thr Ala
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 Ile Leu Gly Ala Asp Leu Asp Val Ile Met Ser Leu Asn Asn Leu Asp
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 Glu Glu Ser Asn Lys Lys His Pro Phe Pro Cys Pro Thr Thr Tyr Arg
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 385 390 395 400
 25 His Leu His Lys Met Ala Ser Ser Ser Gly Glu Gly Lys Glu Leu Tyr
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 Val Ala Phe Ser Arg Glu Gln Ala His Lys Val Tyr Val Gln His Leu
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 Leu Lys Arg Asp Arg Glu His Leu Trp Lys Leu Ile His Glu Gly Gly
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 Ala His Ile Tyr Val Cys Gly Asp Ala Arg Asn Met Ala Lys Asp Val
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	Leu Asp Ile Ser Asp Ser Lys Ile Arg Tyr Glu Ser Gly Asp His Val		
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	Ala Val Tyr Pro Ala Asn Asp Ser Thr Leu Val Asn Gln Ile Gly Glu		
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	355 360 365		
	Thr Ala Leu Thr Tyr Tyr Leu Asp Ile Thr Asn Pro Pro Arg Thr Asn		

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	Val Leu Tyr Glu Leu Ala Gln Tyr Ala Ser Glu Pro Ser Glu Gln Glu		
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	His Leu His Lys Met Ala Ser Ser Ser Gly Glu Gly Lys Glu Leu Tyr		400
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	Leu Ser Trp Val Val Glu Ala Arg Arg His Ile Leu Ala Ile Leu Gln		
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	Asp Tyr Pro Ser Leu Arg Pro Pro Ile Asp His Leu Cys Glu Leu Leu		
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	Gln Glu Arg Ala Trp Leu Arg Glu Gln Gly Lys Glu Val Gly Glu Thr		
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	Leu Leu Tyr Tyr Gly Cys Arg Arg Ser Asp Glu Asp Tyr Leu Tyr Arg		
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	Glu Glu Leu Ala Arg Phe His Lys Asp Gly Ala Leu Thr Gln Leu Asn		
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	Val Ala Phe Ser Arg Glu Gln Ala His Lys Val Tyr Val Gln His Leu		
	595	600	605
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	610	615	620
	Ala His Ile Tyr Val Cys Gly Asp Ala Arg Asn Met Ala Lys Asp Val		
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	Gln Asn Thr Phe Tyr Asp Ile Val Ala Glu Phe Gly Pro Met Glu His		
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	145	150	155
10	Val Asp Leu Thr Gly Val Lys Tyr Ala Val Phe Gly Leu Gly Asn Lys		
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	225	230	235
20	Ser Ile Arg Gln Tyr Glu Leu Val Val His Thr Asp Met Asp Thr Ala		
	245	250	255
	Val Val Tyr Thr Gly Glu Met Gly Arg Leu Lys Ser Tyr Glu Asn Gln		
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	Lys Pro Pro Phe Asp Ala Lys Asn Pro Phe Leu Ala Val Val Thr Thr		
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	Asn Arg Lys Leu Asn Gln Gly Thr Glu Arg His Leu Met His Leu Glu		
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	Leu Asp Ile Ser Asp Ser Lys Ile Arg Tyr Glu Ser Gly Asp His Val		
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30	Ala Val Tyr Pro Ala Asn Asp Ser Ala Leu Val Asn Gln Leu Gly Glu		
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	Ile Leu Gly Thr Asp Leu Asp Ile Val Met Ser Leu Asn Asn Leu Asp		
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	Glu Glu Ser Asn Lys Arg His Pro Phe Pro Cys Pro Thr Thr Tyr Arg		
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	Thr Ala Leu Thr Tyr Tyr Leu Asp Ile Thr Asn Pro Pro Arg Thr Asn		
	370	375	380
	Val Leu Tyr Glu Leu Ala Gln Tyr Ala Ser Glu Pro Ser Glu Gln Glu		
	385	390	395
40	Gln Leu Arg Lys Met Ala Ser Ser Ser Gly Glu Gly Lys Glu Leu Tyr		
	405	410	415
	Leu Ser Trp Val Val Glu Ala Arg Arg His Ile Leu Ala Ile Leu Gln		
	420	425	430
	Asp Tyr Pro Ser Leu Arg Pro Pro Ile Asp His Leu Cys Glu Arg Leu		
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	Pro Arg Leu Gln Ala Arg Tyr Tyr Ser Ile Ala Ser Ser Ser Lys Val		
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	His Pro Asn Ser Val His Ile Cys Ala Val Val Val Glu Tyr Glu Thr		
	465	470	475
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	Lys Glu Pro Ala Gly Glu Asn Gly Arg Arg Ala Leu Val Pro Met Phe		
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	Val Arg Lys Ser Gln Phe Arg Leu Pro Phe Lys Ala Thr Thr Pro Val		
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	Ile Met Val Gly Pro Gly Thr Gly Val Ala Pro Phe Ile Gly Phe Ile		
	530	535	540

Gln Glu Arg Ala Trp Leu Gln Glu Gln Gly Lys Glu Val Gly Glu Thr
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Leu Leu Tyr Tyr Gly Cys Arg Arg Ser Asp Glu Asp Tyr Leu Tyr Arg
565 570 575
5 Glu Glu Leu Ala Gln Phe His Ala Lys Gly Ala Leu Thr Arg Leu Ser
580 585 590
Val Ala Phe Ser Arg Glu Gln Pro Gln Lys Val Tyr Val Gln His Leu
595 600 605
Leu Lys Arg Asp Lys Glu His Leu Trp Lys Leu Ile His Asp Gly Gly
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Ala His Ile Tyr Ile Cys Gly Asp Ala Arg Asn Met Ala Arg Asp Val
625 630 635 640
Gln Asn Thr Phe Cys Asp Ile Val Ala Glu Gln Gly Pro Met Glu His
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Ser Leu Asp Val Trp Ser
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30 <212> DNA
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35 <221> misc_structure
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45 <212> DNA
<213> Artificial Sequence

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<223> Top Strand of a SalI adapter

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tcgacccacg cgtccg 16

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      gccatcgat caagtccggg ccttgcttg cggccgtgaa cagcctcacc atggcgcac  600
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      agaatttgtt tattttctac ggatcgaaa ctggAACCGC tgaggactac gcctccagat 840
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	Ala Ser Arg Leu Ala Lys Glu Gly Ser Gln Arg Phe Gly Leu Lys Thr		
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15	Glu Pro Thr Asp Asn Ala Val Glu Phe Tyr Gln Phe Phe Thr Gly Asp		
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	Asp Val Ala Phe Glu Ser Ala Ser Ala Asp Glu Lys Pro Leu Ser Lys		
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	Leu Lys Tyr Val Ala Phe Gly Leu Gly Asn Asn Thr Tyr Glu His Tyr		
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	Gln Arg Ile Gly Ser Ala Gly Glu Gly Asp Asp Gly Ala Gly Thr Met		
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	Cys Val Thr Glu Asn Glu Ser Leu Ser Pro Glu Asp Glu Thr Val Tyr		
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	Ala Val Pro Phe Ser Leu Leu Ile Glu Gly Ile Thr Lys Leu Gln Pro		
	435	440	445
55	Arg Tyr Tyr Ser Ile Ser Ser Ser Ser Leu Val Gln Lys Asp Lys Ile		
	450	455	460
	Ser Ile Thr Ala Val Val Glu Ser Val Arg Leu Pro Gly Ala Ser His		
	465	470	475
			480

Met Val Lys Gly Val Thr Thr Asn Tyr Leu Leu Ala Leu Lys Gln Lys
485 490 495
Gln Asn Gly Arg Ser Leu Ser Arg Pro Ser Arg Leu Asp Leu Leu His
500 505 510
5 His Gly Pro Arg Asn Lys Tyr Asp Gly Ile His Val Pro Val His Val
515 520 525
Arg His Ser Asn Phe Lys Leu Pro Ser Asp Pro Ser Arg Pro Ile Ile
530 535 540
Met Val Gly Pro Gly Thr Gly Val Ala Pro Phe Arg Gly Phe Ile Gln
10 545 550 555 560
Glu Arg Ala Ala Leu Ala Ala Lys Gly Glu Lys Val Gly Pro Thr Val
565 570 575
Leu Phe Phe Gly Cys Arg Lys Ser Asp Glu Asp Phe Leu Tyr Lys Asp
580 585 590
15 Glu Trp Lys Thr Tyr Gln Asp Gln Leu Gly Asp Asn Leu Lys Ile Ile
595 600 605
Thr Ala Phe Ser Arg Glu Gly Pro Gln Lys Val Tyr Val Gln His Arg
610 615 620
Leu Arg Glu His Ser Glu Leu Val Ser Asp Leu Leu Lys Gln Lys Ala
20 625 630 635 640
Thr Phe Tyr Val Cys Gly Asp Ala Ala Asn Met Ala Arg Glu Val Asn
645 650 655
Leu Val Leu Gly Gln Ile Ile Ala Ala Gln Arg Gly Leu Pro Ala Glu
660 665 670
25 Lys Gly Glu Glu Met Val Lys His Met Arg Arg Arg Gly Arg Tyr Gln
675 680 685
Glu Asp Val Trp Ser
690